

Electronic supplementary materials

## Activation of anthrachamycin biosynthesis in *Streptomyces chattanoogensis* L10 by site-directed mutagenesis of *rpoB*

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**Table S1 Strains and plasmids used in this study**

Strains	Discription	Reference
<i>S.chattanoogaensis</i> L10	Wild type	(Du <i>et al.</i> 2011)
<i>S.chattanoogaensis</i> ZJUY1	The <i>RpoB</i> (H437Y) mutant strain	This study
<i>S.chattanoogaensis</i> ZJUY2	The <i>RpoB</i> (R440C) mutant strain	This study
<i>S.chattanoogaensis</i> ZJUY3	The <i>RpoB</i> (S433P) mutant strain	This study
<i>S.chattanoogaensis</i> ZJUY4	The <i>RpoB</i> (D427N) mutant strain	This study
<i>S.chattanoogaensis</i> ZJUY5	The <i>RpsL</i> (R86L) mutant strain	This study
<i>S.chattanoogaensis</i> ZJUY6	The <i>RpsL</i> (V87K) mutant strain	This study
<i>S.chattanoogaensis</i> ZJUY7	The <i>RpsL</i> (K88E) mutant strain	This study
<i>S.chattanoogaensis</i> ZJUY8	The <i>RpsL</i> (D89R)mutant strain	This study
<i>S.chattanoogaensis</i> ZJUY9	The <i>RpsL</i> (L90K)mutant strain	This study
<i>S.chattanoogaensis</i> ZJUY10	The <i>RpsL</i> (G92D)mutant strain	This study
<i>S.chattanoogaensis</i> ZJUY11	The KS $\alpha$ subunit gene <i>chaA</i> disrupted in ZJUY1	This study
<i>S.chattanoogaensis</i> ZJUY12	The gene <i>chal</i> disrupted in ZJUY1	This study
<i>E.coli</i> DH5 $\alpha$	General cloning strain	This study
<i>E.coli</i> BL21(DE3)	Strain for recombinant protein expression	Invitrogen, USA
<i>E.coli</i> ET12567/PUZ8002	Methylation-deficient <i>E. coli</i> for conjugation with the helper plasmid	(Mao <i>et al.</i> 2015)
Plasmids	Discription	Reference
pKC1139	Temperature-sensitive shuttle vector for gene disruption	(Muth <i>et al.</i> 1989)
pKC1139- <i>rpsL</i>	<i>rpsL</i> and flanking regions cloned in pKC1139 vector	This study
pKC1139- <i>rpoB</i>	<i>rpoB</i> and flanking regions cloned in pKC1139 vector	This study
pKCmu1	The <i>RpoB</i> (H437Y) mutant plasmid	This study
pKCmu2	The <i>RpoB</i> (R440C) mutant plasmid	This study
pKCmu3	The <i>RpoB</i> (S433P) mutant plasmid	This study
pKCmu4	The <i>RpoB</i> (D427N) mutant plasmid	This study
pKCmu5	The <i>RpsL</i> (R86L) mutant plasmid	This study
pKCmu6	The <i>RpsL</i> (V87K) mutant plasmid	This study
pKCmu7	The <i>RpsL</i> (K88E) mutant plasmid	This study
pKCmu8	The <i>RpsL</i> (D89R) mutant plasmid	This study
pKCmu9	The <i>RpsL</i> (L90K) mutant plasmid	This study
pKCmu10	The <i>RpsL</i> (G92D) mutant plasmid	This study
pKC1139- $\Delta$ <i>chaA</i>	<i>chaA</i> disruption plasmid based on pKC1139	This study
pKC1139- $\Delta$ <i>chal</i>	<i>chal</i> disruption plasmid based on pKC1139	This study
pET32a	Expression vector	This study
pET32a- <i>adpA<sub>ch</sub></i>	<i>adpA<sub>ch</sub></i> ORF cloned in <i>EcoRI/HindIII</i> site of pET32a	This study
pClone 007 Blunt Simple Vector	General clone vector	TsingKe, China
pClone1	<i>chal</i> promoter ( <i>chalp</i> ) in pClone 007 Blunt Simple Vector	This study

**Table S2 Oligonucleotide primers used in this study**

Primers	Sequence (5'→3')	Description
F1	<u>GATAT</u> CCGTCTGTGATGAGAAGATGG	For amplification of flanking regions of <i>rpsL</i>
R1	CGGA <u>ATT</u> CCGGCGGACCAGACCAGGG	
F2	<u>GATAT</u> CTCTTGGCCGCCTCGCGCAAC	For amplification of flanking regions of <i>rpoB</i>
R2	CGGA <u>ATT</u> CAAGTTGACGTCGAGCACTAT	
M13F	GTAAAACGACGGCCAGT	M13 universal primers
M13R	CAGGAAACAGCTATGACAT	
F3	AGGTCCTTACAAGGCCACCACGC	For amplification of changed Arg-86 to Leu in RpsL
F4	GCAGGTCCTTCTTACGGCCACC	For amplification of changed Val-87 to Lys in RpsL
F5	CACCCGGCAGGTCCCCACACGGCC	For amplification of changed Lys-88 to Glu in RpsL
F6	GAACACCCGGCAGGCGCTTACACG	For amplification of changed Asp-89 to Arg in RpsL
F7	CGAACACCCGGCTTGTCTTACAC	For amplification of changed Leu-90 to Lys in RpsL
F8	GTAGCGAACATCCGGCAGGTCC	For amplification of changed Gly-92 to Asp in RpsL
F9	GGGTCTGACCTACAAGCGCCGT	For amplification of changed His-437 to Tyr in RpoB
F10	CCACAAGCGCTGTCTGTCCGGCG	For amplification of changed Arg-440 to Cys in RpoB
F11	CAACCCGCTGCCGGGTCTGACC	For amplification of changed Ser-433 to Phe in RpoB
F12	CCAGTTCATGAACCAGACCAAC	For amplification of changed Asp-427 to Asn in RpoB
RmuI	GGCCAGTGCCTAGCTTGGGCT	For amplification fragments without <i>Hind</i> III restriction site
F13	<u>AAGCT</u> TACCGAACCGCCCGCTACG	For amplification of 5' flanking region of <i>chaA</i>
R13	TCTAGAGACCCGTCGCCGGTCACTG	
F14	<u>TCTAG</u> ACCCGAGAGGAGGACGGCATG	For amplification of 3' flanking region of <i>chaA</i>
R14	<u>GATAT</u> CGCCCGGGTCACCAGGTCGAT	
F15	CCGGA <u>ATT</u> CCGGATGAGCCAGGACTCCGCT	For amplification of <i>adpA<sub>ch</sub></i>
R15	CCA <u>AAGCT</u> TGGGTCACCCTACGGGGCGTTC	
F16	FAM-GTAAAACGACGGCCAGTGTC	For amplification of 5'-FAM labeled probe
R16	CACAGGAAACAGCTATGACCAT	
F17	CC <u>ATAT</u> GGGCGCATGGAGATAGCGGATA	For amplification of <i>chal</i> promoter
R17	CC <u>ATAT</u> GGGCGCAGGTGAAATGCGACAC	
F18	TGGCTGCGGCAGTTGCTC	gene expression of <i>relA</i> for qRT PCR
R18	GAAGTCGACGGGCGTGCC	
F19	TCGTCACCAAGACTCCGC	gene expression of <i>relC</i> for qRT PCR

R19	CGTTCAGGTCGGGCATCT	
F20	CTGCAACGCCTTCCACATG	gene expression of <i>chaA</i> for qRT PCR
R20	GCAGGTCGTTCTGCTTGGTG	
F21	CCTGGTCTACAACGGGGTGC	gene expression of <i>chaGT1</i> for qRT PCR
R21	AGCAGATGGAGCGGGACA	
F22	CCACCTGGCGGGCATGAT	gene expression of <i>chaM</i> for qRT PCR
R22	ACCGCAGCAACCCGAGGG	
F23	CGTTCAGTACACCCCGGAGA	gene expression of <i>chaS</i> for qRT PCR
R23	CCTCGATGACCAGGTAGCG	
F24	GCACGACACTCAGGTCAAGG	gene expression of <i>chal</i> for qRT PCR
R24	GGTGCGGGCGATATTGGA	
F25	CAGAATCGTCGGGCTGTGC	gene expression of <i>adpA<sub>ch</sub></i> for qRT PCR
R25	GACGAAGAGCTCACGGGGAT	
F26	AGGGCAACCTCGGTCTGA	gene expression of <i>hrdB</i> for qRT PCR
R26	ATGTGGACGGGGATACGG	
F27	GCTCTAGAGCGAATGGATCATCACCGTG	For amplification of 5' flanking region of <i>chal</i>
R27	CCAAGCTTGGGAACGCGGTTACCTGAC	
F28	GCTCTAGAGCCCCCGGAGAGAACAGCA	For amplification of 3' flanking region of <i>chal</i>
R28	CCGGAATTCGGGAACCCCGCGGCGGGAG	

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**Table S3**  $^1\text{H}$  and  $^{13}\text{C}$  NMR data of anthrachamycin in  $\text{DMSO-}d_6$  (600 MHz)

Anthrachamycin		
Position	$^1\text{H}$ NMR ( $J$ Hz)	$^{13}\text{C}$ NMR
1	—	154.4, qC
2	7.01 (1H, s)	112.5, CH
3	—	141.6, qC
4	7.31 (1H, s)	119.5, CH
4a	—	139.7, qC
5	7.50 (1H, s)	116.25, CH
6	—	155.61, qC
6a	—	119.2, qC
7	—	191.5, qC
7a	—	115.5, qC
8	—	160.3, qC
9	7.29(1H, d, 7.9)	122.8, CH
10	7.80 (1H, t, 7.8)	137.6 CH
11	7.46 (1H, d, 7.2)	117.9, CH
11a	—	137.2, qC
12	—	185.3 qC
12a	—	137.8, qC
12b	—	115.8, qC
13	2.43 (3H, s)	21.9, $\text{CH}_3$
1'	4.92 (1H, d, 5.9)	100.6, CH
2'	3.44, (1H, m)	72.0, CH
3'	3.44, (1H, m)	87.8, CH
4'	3.26, (1H, m)	68.2, CH
5'	3.41, (1H, m)	76.6, CH
6'	3.68, 3.37, m	60.6, $\text{CH}_2$
1''	4.35 (1H, d, 7.8)	104.0, CH
2''	3.08 (1H, m)	73.7, CH
3''	3.17 (1H, m)	76.0, CH
4''	3.05 (1H, m)	70.1, CH
5''	3.19 (1H, m)	76.9, CH
6''	3.69, 3.39, m	61.1, $\text{CH}_2$

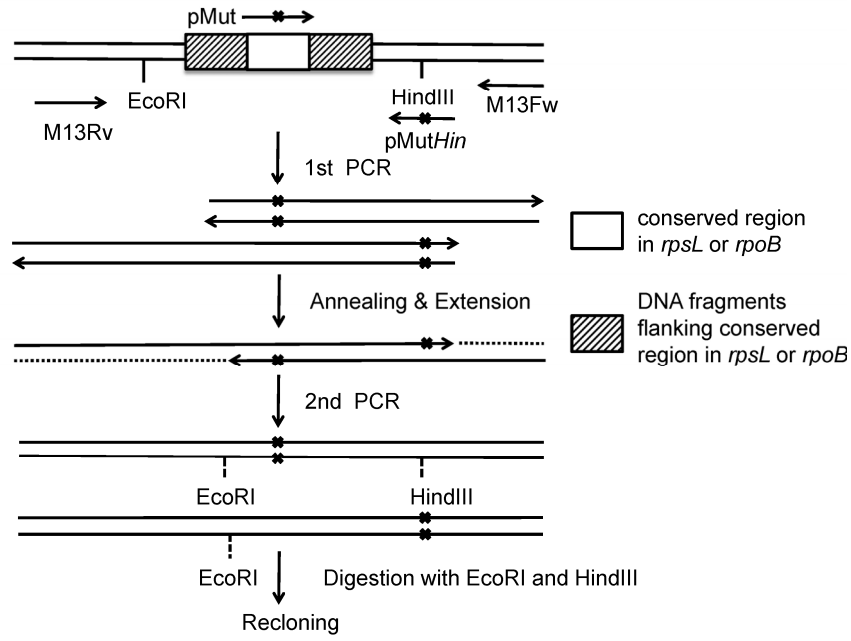


Fig. S1 Schematic representation of overlapping PCR strategy for introducing mutations into *rpsL* or *rpoB*

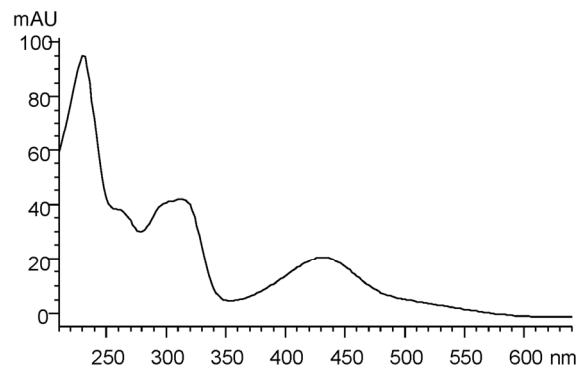


Fig. S2 UV absorption of anthrachamycin



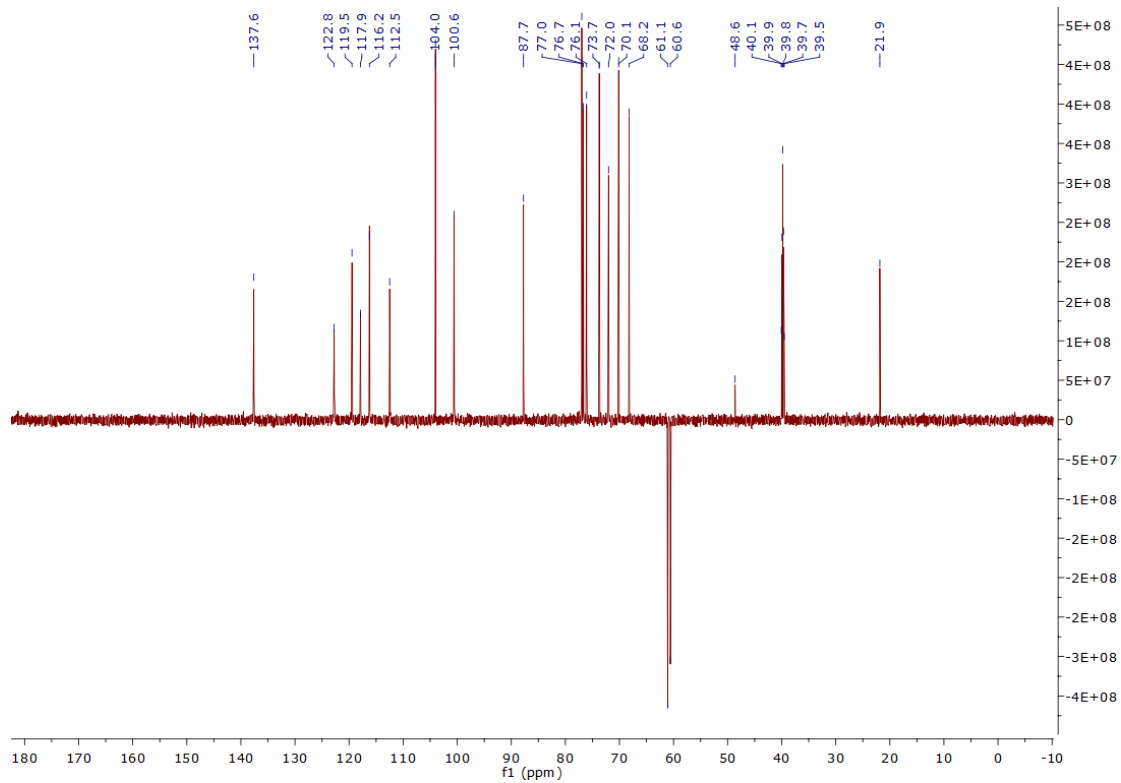


Fig. S5 Distortionless enhancement by polarization transfer (DEPT135) spectrum of anthrachamycin

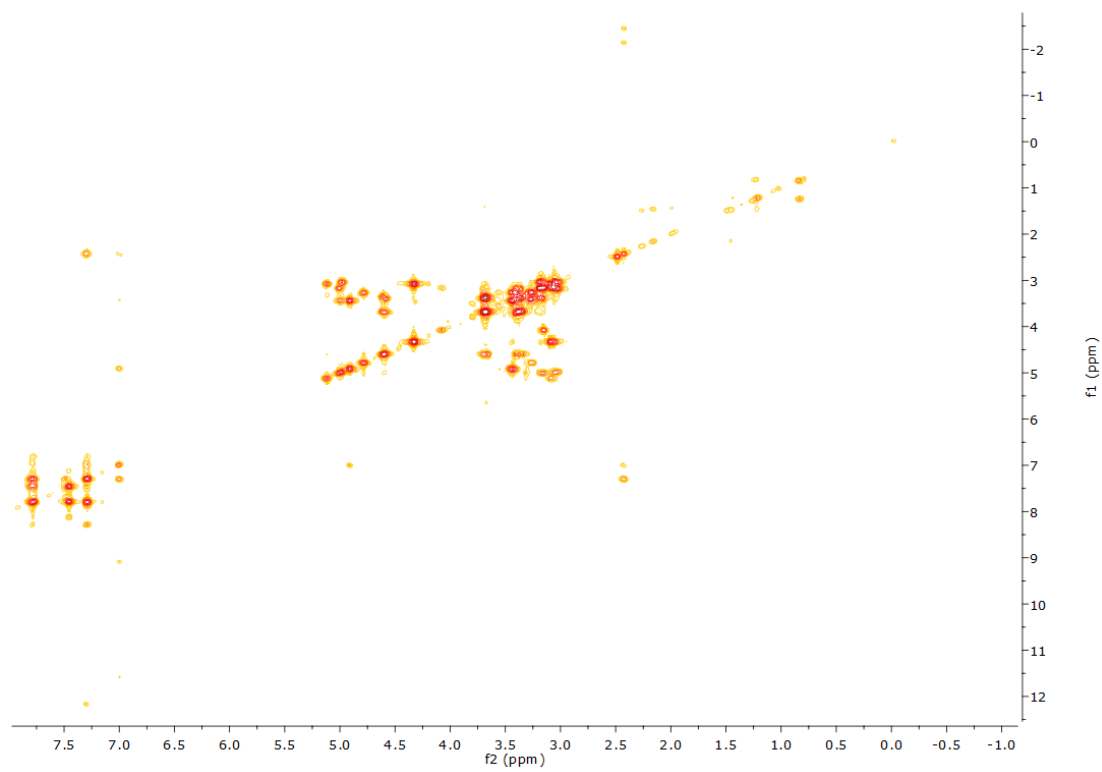
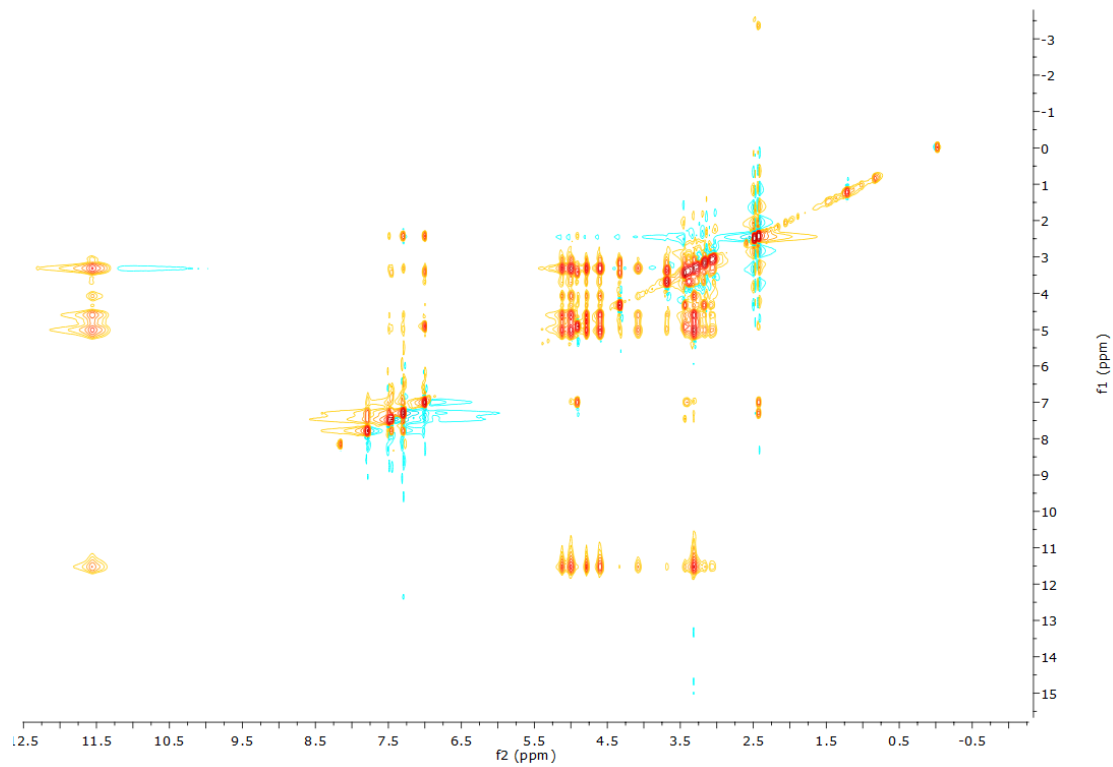
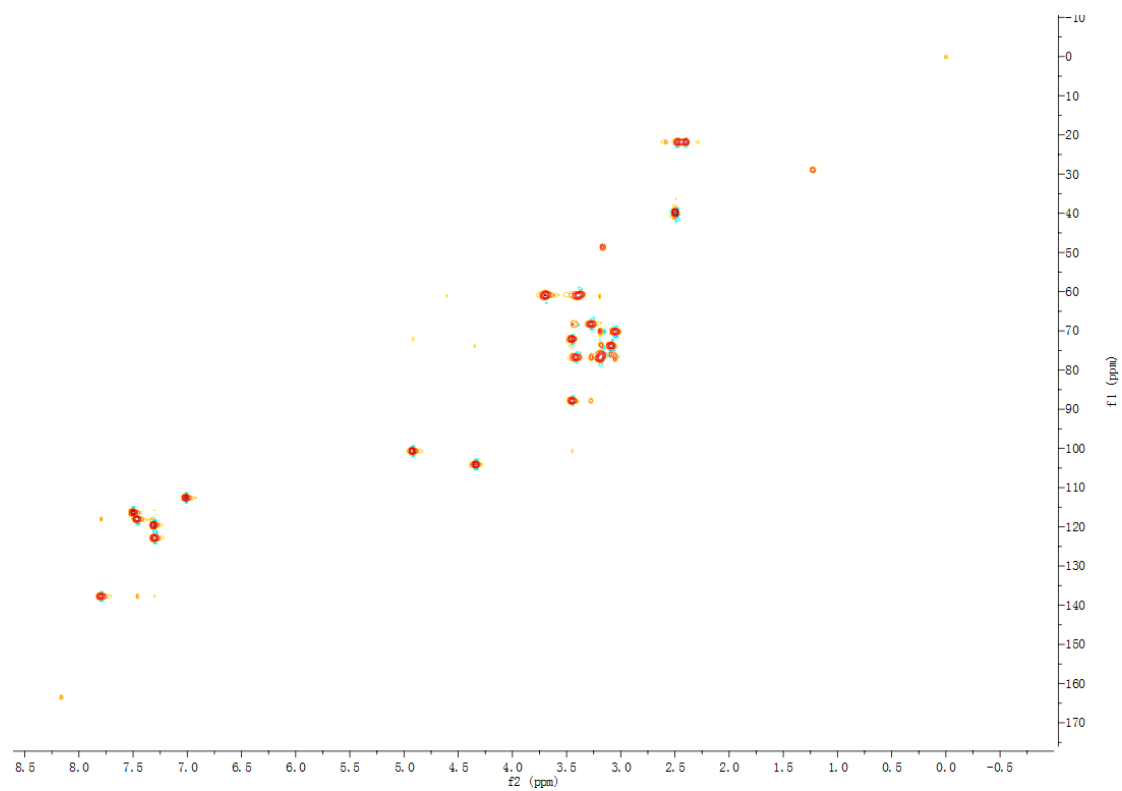


Fig. S6 Correlated spectroscopy (COSY) spectrum of anthrachamycin

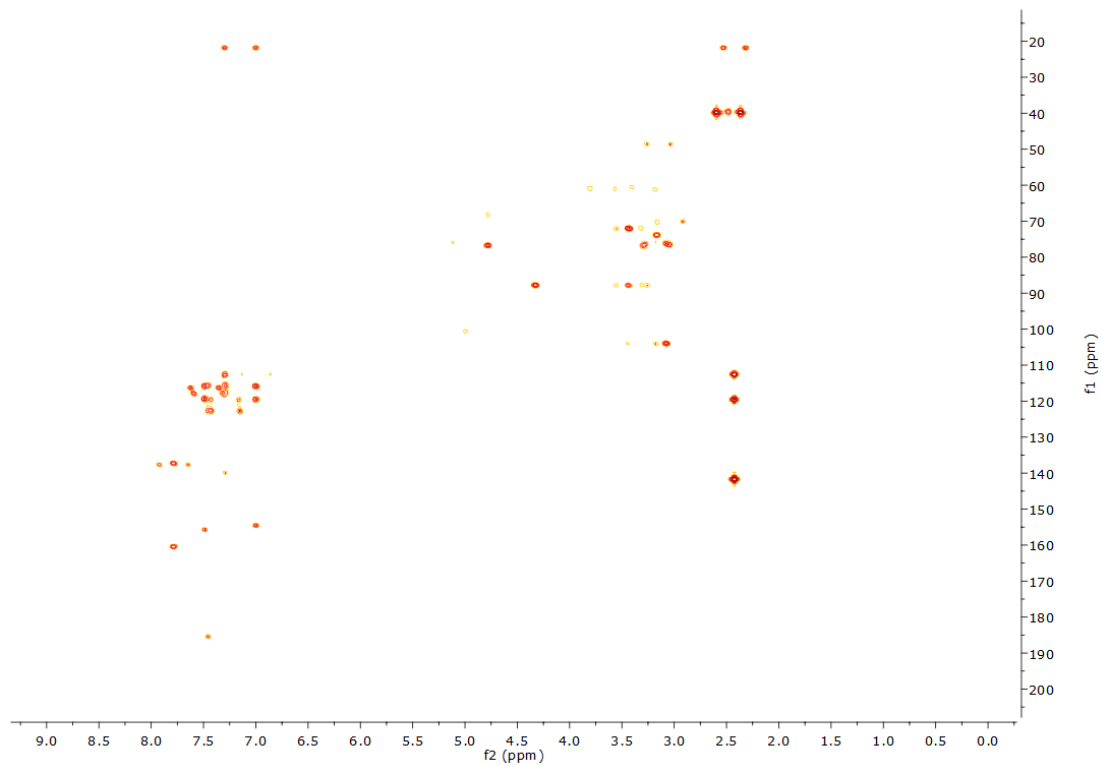




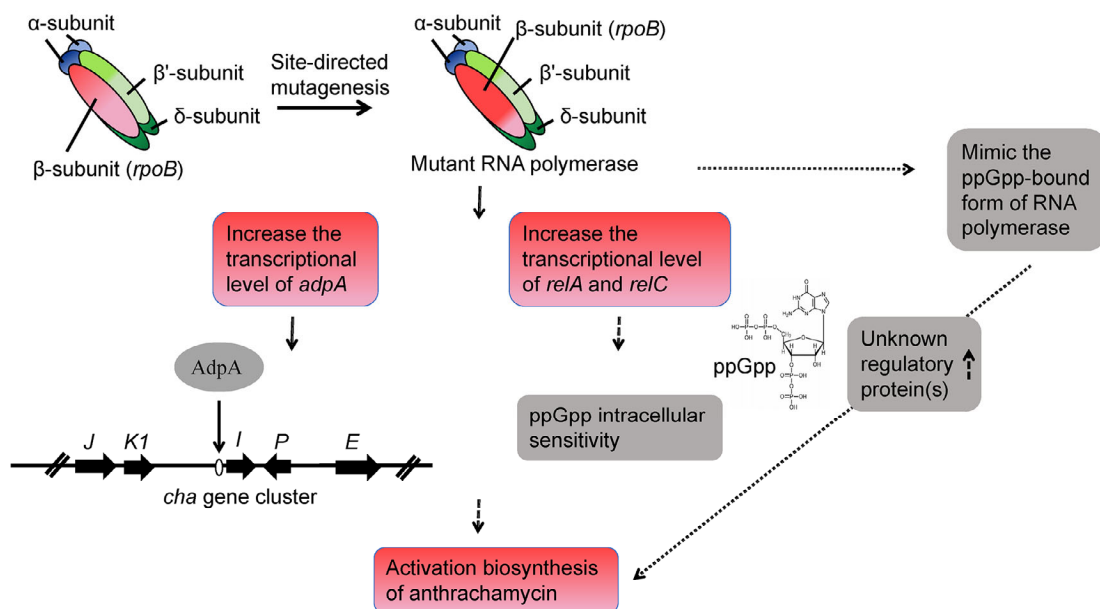
**Fig. S7** Nuclear overhauser enhancement spectroscopy (NOESY) spectrum of anthrachamycin



**Fig. S8** Heteronuclear single quantum coherence (HSQC) spectrum of anthrachamycin



**Fig. S9 Heavy mental binding capacity (HMBC) spectrum of anthrachamycin**



**Fig. S10 Speculated mechanism for activating anthrachamycin biosynthesis**

Arrows indicate activation and dashed lines denote that the precise mechanism is under investigation

## References

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