



Effect of CO₂ enrichment on the glucosinolate contents under different nitrogen levels in bolting stem of Chinese kale (*Brassica alboglabra* L.)^{*}

Gui-xiao LA¹, Ping FANG^{†‡1,2}, Yi-bo TENG¹, Ya-juan LI², Xian-yong LIN^{1,2}

⁽¹⁾Ministry of Education Key Lab of Environment Remediation and Ecological Health, College of Environmental and Natural Resources Science, Zhejiang University, Hangzhou 310029, China)

⁽²⁾Zhejiang Key Laboratory of Subtropical Soil Science and Plant Nutrition, College of Environmental and Natural Resources Science, Zhejiang University, Hangzhou 310029, China)

[†]E-mail: zdpfang@zju.edu.cn

Received Nov. 3, 2008; Revision accepted Mar. 31, 2009; Crosschecked Apr. 28, 2009

Abstract: The effects of CO₂ enrichment on the growth and glucosinolate (GS) concentrations in the bolting stem of Chinese kale (*Brassica alboglabra* L.) treated with three nitrogen (N) concentrations (5, 10, and 20 mmol/L) were investigated. Height, stem thickness, and dry weights of the total aerial parts, bolting stems, and roots, as well as the root to shoot ratio, significantly increased as CO₂ concentration was elevated from 350 to 800 μl/L at each N concentration. In the edible part of the bolting stem, 11 individual GSs were identified, including 7 aliphatic and 4 indolyl GSs. GS concentration was affected by the elevated CO₂ concentration, N concentration, and CO₂×N interaction. At 5 and 10 mmol N/L, the concentrations of aliphatic GSs and total GSs significantly increased, whereas those of indolyl GSs were not affected, by elevated atmospheric CO₂. However, at 20 mmol N/L, elevated CO₂ had no significant effects on the concentrations of total GSs and total indolyl GSs, but the concentrations of total aliphatic GSs significantly increased. Moreover, the bolting stem carbon (C) content increased, whereas the N and sulfur (S) contents decreased under elevated CO₂ concentration in the three N treatments, resulting in changes in the C/N and N/S ratios. Also the C/N ratio is not a reliable predictor of change of GS concentration, while the changes in N and S contents and the N/S ratio at the elevated CO₂ concentration may influence the GS concentration in Chinese kale bolting stems. The results demonstrate that high nitrogen supply is beneficial for the growth of Chinese kale, but not for the GS concentration in bolting stems, under elevated CO₂ condition.

Key words: Carbon dioxide (CO₂), *Brassica alboglabra*, Nitrogen (N), Growth, Bolting stem, Aliphatic glucosinolates, Indolyl glucosinolates, Carbon/nitrogen ratio (C/N), Nitrogen/sulfur ratio (N/S)

doi:10.1631/jzus.B0820354

Document code: A

CLC number: S635.9

INTRODUCTION

Epidemiological studies show that there is a negative relationship between Brassicaceae vegetable intake and the risk of a number of cancers (Wattenberg, 1993; Kohlmeier and Su, 1997; Price et al., 1998). Recently, it has been widely recognized that

some of the cancer-chemoprotective activities in these vegetables are attributable to their contents of glucosinolates (GSs) (Zhao et al., 1992; Wattenberg, 1993; Tawfiq et al., 1995; Fahey et al., 1997; Rosa et al., 1997; Holst and Williamson, 2004) (Fig.1). GSs are amino acid-derived secondary compounds, a characteristic of dicotyledonous plants. So far, more than 20 GSs have been identified in Brassicaceae family (Rodman, 1991). They can be grouped into aliphatic, aromatic, and indolyl GSs according to the amino acid, from which they are derived (Louda and

[‡] Corresponding author

* Project (No. 2007CB109305) supported by the National Basic Research Program (973) of China

Mole, 1991; Halkier and Du, 1997). Besides the health-promoting properties, GSs also play an important role in plant defense against insects and herbivores, and are utilized as special flavors in the food industry (Fenwick *et al.*, 1983; Chew, 1988; Gijzen *et al.*, 1989; Baik *et al.*, 2003).

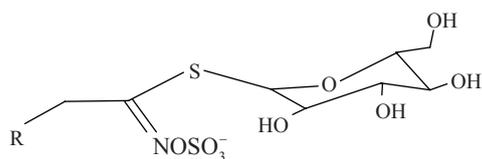


Fig.1 General structure of glucosinolates (GSs)

It is widely accepted that GS content is affected by environmental factors including climatic conditions, nutritional availability, and agronomic practices, in addition to genetic characteristics (Fenwick *et al.*, 1989). Presently, cancer incidence and global climate change are major topical issues. Because GSs exhibit cancer-chemoprotective activities, many researches are focused on the effect of climatic conditions on changes in GS content in vegetables (Schreiner, 2005). The rise in atmospheric carbon dioxide (CO₂) concentration is one of the most prominent climatic changes in recent decades (IPCC, 2007). Climate simulations indicate that the atmospheric CO₂ concentration is expected to reach 700 μl/L by the end of this century, which is double as much as the current CO₂ concentration (Caswell, 2004). Increasing atmospheric CO₂ concentration may affect natural ecosystems by directly influencing plant growth and photochemistry due to an increased photosynthetic rate, especially in C₃ plants (Islam *et al.*, 1996; Kim *et al.*, 2001; Das *et al.*, 2002). In spite of increased plant growth under elevated CO₂ concentrations (Bazzaz, 1990; Mooney *et al.*, 1991; Amthor, 2001), aerial plant parts accumulate generally less nitrogen (N), and carbon (C)/N ratio increases (Baxter *et al.*, 1994; Epron *et al.*, 1996), which could influence plant secondary metabolites synthesis and concentration. GSs, as N- and C-containing secondary metabolites, might be affected by atmospheric CO₂ enrichment owing to changes in the plant's C supply and N content (Cotrufo *et al.*, 1998). Moreover, Habash *et al.* (1995) observed that synthesis of the amino acid precursors of GSs from triosephosphates increased at an elevated CO₂ concentration. However, Karowe *et al.* (1997) reported that total foliar GS content in

mustard decreased significantly under elevated CO₂ conditions. This conflict might reflect a species-specific response to elevated CO₂ concentration (Karowe *et al.*, 1997). An increasing number of studies indicate that N availability can have a large impact on the plant response to elevated CO₂ concentration (Kimball *et al.*, 1995; 2002). Moreover, N application is one of the most important nutrient factors that significantly affect GS synthesis and content (Schnug, 1989; Zhao *et al.*, 1994; Ahmad *et al.*, 2007). In oilseed rape (*Brassica napus* L.), the GS content in the seed decreased with the higher N supply in sulfur (S)-deficient soil, but increased in S-sufficient soil (Zhao *et al.*, 1994). However, in broccoli sprouts (*Brassica oleracea* var. *italica*), N fertilization has a negative effect on GS content, even at a very low concentration (Aires *et al.*, 2006).

Therefore, it is logical to take N nutrition into account when investigating the effects of CO₂ enrichment on GS content. At present, limited information is available on the effect of elevated atmospheric CO₂ in combination with N availability on GS content in brassicaceous vegetables. It is not clear whether GS content changes consistently at different N levels. In this study, we determined the interactive effects of elevated CO₂ concentration and N availability on the contents of individual and total GSs in the edible part of the bolting stem of Chinese kale, which is a nutritionally healthy vegetable belonging to the Brassicaceae (Cruciferae) family and rich in GSs, and has spread quickly in southeastern China, Taiwan region, and Japan since the last decade (He *et al.*, 2002).

MATERIALS AND METHODS

Plant growth

Seeds of Chinese kale (*Brassica alboglabra* L. var. *Sijicutiao*) were sown in vermiculite and germinated in a greenhouse with computer-controlled growth conditions on Huajiachi campus of Zhejiang University, Hangzhou, China. The growth conditions in the greenhouse were constant day/night temperature of 23/18 °C and the natural photoperiod. After two weeks, the seedlings were irrigated with nutrient solution containing 5 mmol N/L. Four weeks later, healthy seedlings in which the third true leaf had emerged were transplanted to 1.8-L pots containing

nutrient solution with 10 mmol N/L by being fixed in a foam cavity with sponge. Each pot contained two seedlings and was covered with black plastic foil to prevent algal growth and evaporation. All the pots were transferred to four growth chambers with 65% relative humidity, constant day/night temperature of 23/18 °C, and 500 $\mu\text{mol}/(\text{m}^2\text{s})$ photosynthetically-active radiation for 16 h/d. One week after transplanting, plants were treated with 5.0 mmol N/L (low N concentration), 10 mmol N/L (medium N concentration), or 20 mmol N/L (high N concentration) (Chen *et al.*, 2005), and were grown under either ambient CO_2 [(350 \pm 20) $\mu\text{L}/\text{L}$, denoted as A] or elevated CO_2 [(800 \pm 20) $\mu\text{L}/\text{L}$, denoted as E). The N was supplied as NH_4NO_3 and the basic nutrient solution contained 1 mmol/L K_2HPO_4 , 4 mmol/L KCl, 3 mmol/L CaCl_2 , 2 mmol/L $\text{MgSO}_4\cdot 7\text{H}_2\text{O}$, 36 $\mu\text{mol}/\text{L}$ ethylene diamine tetraacetic acid (EDTA)-Fe, 46.4 $\mu\text{mol}/\text{L}$ H_3BO_3 , 9.07 $\mu\text{mol}/\text{L}$ $\text{MnCl}_2\cdot 4\text{H}_2\text{O}$, 0.765 $\mu\text{mol}/\text{L}$ $\text{ZnSO}_4\cdot 7\text{H}_2\text{O}$, 0.3 $\mu\text{mol}/\text{L}$ $\text{CuSO}_4\cdot 5\text{H}_2\text{O}$, and 0.09 $\mu\text{mol}/\text{L}$ $\text{H}_2\text{MoO}_4\cdot \text{H}_2\text{O}$ (Hoagland and Arnon, 1938). CO_2 was supplied from gas tanks for the elevated CO_2 concentration treatment. There were two replicate chambers per CO_2 concentration and three replicate pots per N treatment in each chamber. The position of every pot was rotated randomly when the solutions were renewed every three days. To avoid a potential chamber effect, the pots were switched with those from the other chamber with the same CO_2 concentration every week. The culture solutions were continuously aerated and adjusted to pH 6.0 using diluted NaOH or HCl every day until harvest. After treated for 35 d, the selected growth parameters were measured, and every plant was separated into different parts, weighed, and lyophilized. The bolting stems were ground into a powder and stored in a desiccator at $-20\text{ }^\circ\text{C}$ prior to C, N, S, and GS analyses.

Extract preparation for glucosinolate analysis

Extracts for GS analysis were prepared according to the method of Kiddle *et al.* (2001) with some modifications. Triplicate samples (0.1 g) of freeze-dried powder were each weighed in 5 ml tubes, and crude GSs were extracted with 1.5 ml 70% (v/v) methanol at 75 °C for 10 min in a water bath. The mixture was centrifuged at 5000 \times g for 10 min at 4 °C and the supernatant was decanted into another tube.

The extraction was repeated twice from residues using the same procedure. The three supernatants were combined to give a final extract volume of 5 ml. 2 ml of each GS extract was added to a mini-column filled with diethylaminoethanol (DEAE) Sephadex A-25 (80 mg as dry matter) (170170-01, Amersham Biosciences, Sweden) activated with 0.5 mol/L pyridine acetate, and desulfated by sulfatase (S9626, Sigma-Aldrich Co., MO, USA). After reaction at room temperature overnight (16 h), the desulfated glucosinolates (desulfoGSs) were eluted with 2 ml deionized water and stored at $-20\text{ }^\circ\text{C}$ prior to high-performance liquid chromatography (HPLC) analysis. 2-PropenylGS (sinigrin, S1647, Sigma-Aldrich Co., MO, USA) was used as an external standard for GS quantitative analysis.

High performance liquid chromatography

The desulfated extract (20 μL) was analyzed by HPLC (Beckman Coulter System Gold HPLC, Beckman, USA) using a Hypersil ODS2 column (250 mm \times 4.6 mm, 5 μm ; Elite, China) with a Beckman Ultrasphere ODS guard column (45 mm \times 4.6 mm, 5 μm ; Beckman, USA). The wavelength of the ultraviolet detector was set at 227 nm. The mobile phase was a mixture of deionized water (A) and acetonitrile (B) and ran at a flow rate of 1 ml/min. The elution program consisted of a linear gradient from 0 to 20% (B) in 18 min and constant 20% (B) for a further 16 min, then the column was eluted with 100% (B) for 5 min and equilibrated with 0 (B) for 6 min prior to the injection of the next sample (Macfarlane-Smith and Griffiths, 1988).

Mass spectrometry analysis

The separated compounds were identified according to the mass spectrometry (MS) data obtained by a liquid chromatography-mass spectrometry data (LC-MSD) system (Agilent 1100 LC/MSD, Agilent Co., USA). The conditions used for the electrospray source were ionspray mode, positive; capillary voltage, 4 kV; nebulizer pressure, 42184.8 Pa; fragment voltage, 100 V; curtain gas, nitrogen; drying gas flow, 13 L/min; desolvation gas temperature, 350 °C. Each individual desulfoGS was identified according to their $(\text{M}+\text{H})^+$, $(\text{M}+\text{Na})^+$, $(\text{M}+\text{K})^+$, and $(\text{M}\text{-glucosyl}+\text{H})^+$ in the MS.

Analyses of bolting stem carbon, nitrogen, and sulfur contents

Bolting stem C content was determined by titration after digestion by H₂SO₄ and potassium permanganate (Lu, 1999). Bolting stem N content was determined titrimetrically using the Kjeldahl procedure with salicylic acid, sodium thiosulfate, and zinc as catalysts (Pruden *et al.*, 1985). For measuring the S content, 0.1 g aliquot of the ground materials was digested with HNO₃ and HClO₄. The S content was determined using an inductively coupled plasma atomic emission spectrometer (ICP-MS; Agilent, 7500a, USA) (Lu, 1999).

Statistical analysis

Two-way analysis of variance (ANOVA) was performed to determine the main effects (N and CO₂ treatments) and their interactions with a significance level of $P < 0.05$. The normality of data and the homogeneity of variances were verified by Shapiro-Wilk test and Bartlett test, respectively, before using ANOVA. Differences between means were analyzed by Fisher's protected least significant difference (LSD) procedure. All statistical analysis procedures were performed by using SPSS for Windows version 12.0 (SPSS, Chicago, IL, USA).

RESULTS

Plant growth

Elevated CO₂ concentration significantly increased plant height, stem thickness, dry weights of

the total aerial parts, bolting stems, and roots, and the root-to-shoot ratio, compared with those in the ambient CO₂ treatment (Table 1). Regardless of N concentration, the height, stem thickness, dry weights of the total aerial parts, bolting stems, and roots, and root-to-shoot ratio increased by 15.64%, 11.79%, 11.91%, 15.03%, 16.34%, and 3.90%, respectively, with elevated CO₂ concentration. Nitrogen levels also significantly affected each growth parameter (Table 1). The 10 mmol N/L solution significantly increased the height, stem thickness, and dry weights of the total aerial parts, bolting stems, and roots, compared with those in the 5 mmol N/L solution in both CO₂ regimes. However, there was no significant difference between the 10 and 20 mmol N/L solution treatments for the above parameters in both CO₂ conditions, except that height and dry weight of the total aerial parts differed significantly between the two N treatments at the elevated CO₂ concentration. The root-to-shoot ratio did not differ significantly among the three N concentrations at the ambient CO₂ concentration, but there was a significant difference between the 5 and 10 mmol/L N concentrations at the elevated CO₂ concentration. Moreover, there were significant CO₂ × N interactions for plant height ($P < 0.01$) and dry weights of the total aerial parts ($P < 0.01$) and roots ($P < 0.05$), but not for bolting stem thickness, dry weight of the bolting stem, or root-to-shoot ratio (Table 1).

Glucosinolate content

A typical HPLC profile of desulfoGSs in the bolting stem is shown in Fig.2. Eleven individual GSs

Table 1 Effects of elevated CO₂ concentration on height, stem thickness, dry weights of aerial parts, bolting stems, and roots, and root-to-shoot ratio of Chinese kale at three nitrogen (N) concentrations

CO ₂ (μL/L)	N (mmol/L)	Height (cm)	Stem thickness (cm)	Dry weight (g/plant)			Root-to-shoot ratio
				Total aerial part	Bolting stem	Root	
350	5	26.13±3.00 ^d	0.72±0.02 ^d	4.84±0.10 ^c	1.63±0.10 ^d	0.45±0.03 ^d	0.093±0.006 ^c
	10	29.13±2.59 ^c	0.82±0.09 ^{bc}	6.30±0.12 ^c	2.24±0.08 ^b	0.61±0.02 ^b	0.096±0.005 ^{abc}
	20	28.25±1.98 ^{cd}	0.81±0.04 ^c	6.27±0.07 ^c	2.29±0.13 ^b	0.60±0.02 ^b	0.095±0.004 ^{bc}
800	5	32.44±2.22 ^b	0.77±0.04 ^{cd}	5.38±0.17 ^d	1.96±0.07 ^c	0.51±0.03 ^c	0.095±0.004 ^{bc}
	10	34.88±1.48 ^a	0.88±0.08 ^{ab}	6.97±0.13 ^b	2.53±0.16 ^a	0.70±0.02 ^a	0.100±0.002 ^a
	20	29.25±1.58 ^c	0.89±0.07 ^a	7.14±0.16 ^a	2.60±0.10 ^a	0.71±0.03 ^a	0.100±0.007 ^{ab}
Source of variance							
CO ₂		***	**	***	***	***	*
N		***	***	***	***	***	*
CO ₂ × N		**	NS	**	NS	*	NS

Data followed by the same superscript letter(s) indicate no significant difference at the $P < 0.05$ level. Values are the mean ± SD. Significance levels indicated by two-way ANOVA: NS, not significant; * $P < 0.05$; ** $P < 0.01$; *** $P < 0.001$

were identified according to the elution order from the HPLC column and confirmed by electrospray ionization mass spectrometry (ESI-MS) analysis based on their MS data. All GSs were identified by analyzing the chemical structure of the aglucone chain R and described according to the trivial names that have been popularly used for decades (Fig.1 and Table 2). Seven aliphatic GSs comprising glucoiberin, progoitrin, sinigrin, glucoraphanin, glucoalyssin, gluconapin, and glucoerucin, and four indolyl GSs consisting of 4-hydroxyglucobrassicin, glucobrassicin, 4-methoxyglucobrassicin, and neoglucobrassicin, were identified.

The major GSs were gluconapin, sinigrin, glucoraphanin, and glucoiberin, which constituted about 54.48%, 8.80%, 8.63%, and 5.93%, respectively, of the total GS concentration on average. The proportions

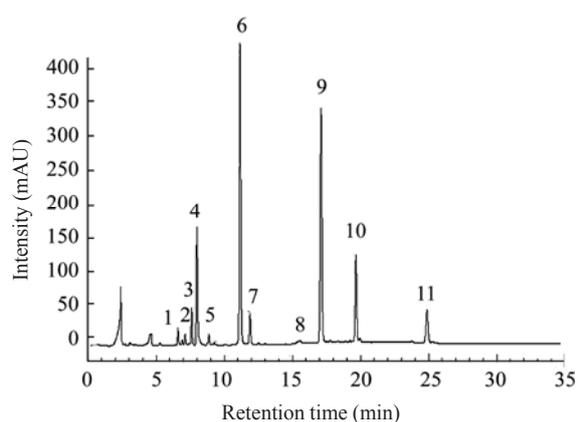


Fig.2 Typical HPLC elution profile of desulfated glucosinolates in bolting stem of Chinese kale
Peak numbers refer to GSs listed in Table 2

of the other seven individual GSs were less than 5% on average. The total GS concentration in each treatment was expressed as the sum of the 11 identified individual GS species (Fig.3). The total GS content ranged 4.82~6.41 $\mu\text{mol/g}$ DW (dry weight). CO_2 concentration significantly affected the concentrations of total GSs, total aliphatic GSs, and all individual aliphatic GSs except glucoerucin, but not the concentrations of total or individual indolyl GSs (Table 3 and Fig.3). Under elevated CO_2 , the total GS concentration increased in the 5 and 10 mmol N/L treatments by 15.59% and 18.01%, respectively, compared with those at ambient CO_2 . However, elevated CO_2 did not affect the total GS concentration at 20 mmol N/L. In the 5 and 10 mmol N/L solution treatments, all individual aliphatic GSs increased under elevated CO_2 compared with the ambient CO_2

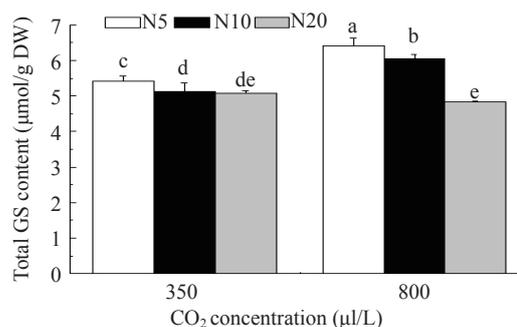


Fig.3 Comparison of the total GS concentration in bolting stems of Chinese kale grown at ambient CO_2 (350 $\mu\text{L/L}$) and elevated CO_2 (800 $\mu\text{L/L}$) concentrations under three nitrogen (N) concentrations
N5: 5 mmol N/L; N10: 10 mmol N/L; N20: 20 mmol N/L.
Columns with the same letter(s) indicate no significant difference at the $P < 0.05$ level. The bars represent the standard error

Table 2 Desulfated glucosinolates identified in bolting stems of Chinese kale

No.*	Retention time (min)	Side-chain structure	Trivial name	Desulfated molecular weight	Response factor [†]
1	6.58	$\text{CH}_3\text{-SO-(CH}_2\text{)}_3\text{-}$	Glucoiberin	343	1.07
2	7.10	$\text{CH}_2\text{=CHCH(OH)CH}_2\text{-}$	Progoitrin	309	1.09
3	7.59	$\text{CH}_2\text{=CHCH}_2\text{-}$	Sinigrin	279	1.00
4	7.98	$\text{CH}_3\text{-SO-(CH}_2\text{)}_4\text{-}$	Glucoraphanin	357	1.07
5	9.24	$\text{CH}_3\text{-SO-(CH}_2\text{)}_5\text{-}$	Glucoalyssin	371	1.07
6	11.13	$\text{CH}_2\text{=CH-(CH}_2\text{)}_2\text{-}$	Gluconapin	293	1.11
7	11.86	Indole-(4-OH)-3- $\text{CH}_2\text{-}$	4-Hydroxyglucobrassicin	384	0.28
8	15.54	$\text{CH}_3\text{-S-(CH}_2\text{)}_4\text{-}$	Glucoerucin	341	1.00 [‡]
9	17.08	Indole-3- $\text{CH}_2\text{-}$	Glucobrassicin	368	0.29
10	19.64	Indole-(4-O CH_3)-3- $\text{CH}_2\text{-}$	4-Methoxyglucobrassicin	398	0.25
11	24.86	Indole-(O CH_3)-3- $\text{CH}_2\text{-}$	Neoglucobrassicin	398	0.20

* Numbering is based on the elution order of desulfated glucosinolates from HPLC; [†] The response factors relative to the standard sinigrin were experimentally determined with HPLC by the International Organization for Standardization (ISO 9167-1) in 1992 for individual GS content in rapeseed; [‡] Not yet determined by the ISO

treatment, whereas individual indolyl GSs were not affected by the elevated CO₂ concentration. However, in the 20 mmol N/L treatment under the elevated CO₂ condition, the total aliphatic GS concentration decreased significantly ($P<0.01$), while there was no significant effect on total indolyl GS content ($P>0.05$), resulting in a slight decrease in total GS concentration ($P>0.05$). Nitrogen concentration also significantly affected the concentrations of total GSs, total aliphatic GSs, individual aliphatic GSs except glucoerucin, total indolyl GSs, and individual indolyl GSs except 4-methoxyglucobrassicin (Tables 3 and 4). The 5 mmol N/L treatment increased the total aliphatic GS concentration compared with the 10 and 20 mmol N/L treatments, but there was no significant difference in the effects of the 10 and 20 mmol N/L treatments at ambient CO₂. However, under the elevated CO₂ concentration, the total aliphatic GS concentration decreased significantly with increased N supply. Total indolyl GS concentration increased in the 20 mmol/L N treatment compared with that in the 5 mmol N/L treatment at both CO₂ concentrations, whereas the difference between the 10 and 20 mmol N/L treatments was not significant at the ambient CO₂ concentration but was significant under the elevated CO₂ concentration. Moreover, there were significant CO₂×N interactions for total GS concentration ($P<0.001$), total aliphatic GS concentration ($P<0.001$), and concentrations of all individual aliphatic GSs ($P<0.01$) except glucoerucin, but not for the concentrations of total indolyl GSs or individual indolyl GSs ($P>0.05$) except neoglucobrassicin.

Bolting stem carbon, nitrogen, and sulfur contents

With elevated CO₂, the C content in the bolting stem increased in the 5, 10, and 20 mmol N/L treatments by 11.38%, 13.62%, and 10.46%, respectively, relative to the ambient CO₂ concentration (Table 5). The effect of CO₂ on C content was strongly significant ($P<0.001$), but the effects of N concentration and CO₂×N interactions were not significant ($P>0.05$). The N content in bolting stem increased significantly with increasing N concentration in the nutrient solution under the same CO₂ regime. In contrast, the N content decreased in the three N treatments under the enriched CO₂ concentration compared with that of the ambient CO₂ treatment (Table 5). The decreases in N content in the 5, 10, and 20 mmol N/L treatments were 10.97%, 13.27%, and 4.59%, respectively. There were significant CO₂×N interactions for the C/N ratio (Table 5). The C/N ratio at the three N concentrations all increased under the elevated CO₂ concentration by 25.13%, 31.20%, and 15.73%, respectively. There were significant N concentration ($P<0.01$) and CO₂ concentration ($P<0.001$) effects on S content, but CO₂×N interactions were not significant for S content. Under the elevated CO₂ concentration, S content decreased in all of the N treatments. Because of the decreases in both N and S contents, the N/S ratio changed. The N/S ratio was significantly affected by CO₂ concentration ($P<0.05$), N concentration ($P<0.001$), and CO₂×N interactions ($P<0.01$). Under the elevated CO₂ concentration, the N/S ratio decreased significantly in the 5 and 10 mmol N/L treatments ($P<0.05$), but the decrease in N and S

Table 3 Effect of CO₂ concentration on the individual and total aliphatic GS concentrations in bolting stems of Chinese kale at three nitrogen (N) concentrations

CO ₂ (μL/L)	N (mmol/L)	GS concentration (μmol/g DW)							
		GIB	PRO	GRA	SIN	GAL	GNP	GRU	Total
350	5	0.17±0.02 ^d	0.13±0.02 ^c	0.15±0.01 ^d	0.51±0.02 ^b	0.10±0.01 ^b	3.62±0.11 ^b	0.05±0.01 ^b	4.71±0.12 ^c
	10	0.31±0.01 ^c	0.14±0.01 ^c	0.46±0.08 ^c	0.40±0.08 ^{bc}	0.10±0.01 ^b	2.89±0.12 ^d	0.06±0.01 ^{ab}	4.36±0.22 ^d
	20	0.34±0.01 ^b	0.25±0.01 ^b	0.88±0.06 ^a	0.35±0.02 ^c	0.23±0.03 ^a	2.17±0.03 ^e	0.04±0.01 ^b	4.27±0.08 ^d
800	5	0.46±0.01 ^a	0.38±0.07 ^a	0.18±0.02 ^d	0.64±0.12 ^a	0.21±0.05 ^a	3.85±0.14 ^a	0.07±0.03 ^a	5.79±0.14 ^a
	10	0.35±0.03 ^b	0.16±0.04 ^c	0.49±0.03 ^c	0.70±0.04 ^a	0.15±0.02 ^b	3.39±0.17 ^c	0.06±0.00 ^{ab}	5.31±0.09 ^b
	20	0.33±0.02 ^{bc}	0.25±0.03 ^b	0.57±0.02 ^b	0.35±0.00 ^c	0.22±0.02 ^a	2.16±0.03 ^e	0.04±0.01 ^b	3.93±0.01 ^e
Source of variance									
CO ₂		***	***	**	***	**	***	NS	***
N		NS	***	***	***	***	***	NS	***
CO ₂ ×N		***	***	***	**	**	**	NS	***

GIB: glucoiberin; PRO: progointrin; GRA: glucoraphanin; SIN: sinigrin; GAL: glucoalyssin; GNP: gluconapin; GRU: glucoerucin. Data followed by the same superscript letter(s) indicate no significant difference at $P<0.05$ level. Values are mean±SD. Significance levels indicated by two-way ANOVA: NS, not significant; * $P<0.05$; ** $P<0.01$; *** $P<0.001$

Table 4 Effect of CO₂ concentration on the individual and total indolyl GS concentrations in bolting stems of Chinese kale at three nitrogen (N) concentrations

CO ₂ (μL/L)	N (mmol/L)	GS concentration (μmol/g DW)				
		4HGB	GBS	4MGB	NGBS	Total
350	5	0.01±0.00 ^{bc}	0.18±0.04 ^b	0.26±0.03 ^a	0.24±0.01 ^b	0.70±0.06 ^{cd}
	10	0.02±0.01 ^a	0.25±0.01 ^a	0.25±0.03 ^a	0.25±0.03 ^b	0.77±0.07 ^{bc}
	20	0.02±0.00 ^a	0.28±0.00 ^a	0.27±0.01 ^a	0.25±0.01 ^b	0.81±0.01 ^{ab}
800	5	0.01±0.00 ^c	0.14±0.04 ^b	0.25±0.03 ^a	0.23±0.02 ^b	0.62±0.07 ^d
	10	0.02±0.00 ^{ab}	0.25±0.01 ^a	0.25±0.03 ^a	0.24±0.04 ^b	0.75±0.05 ^{bc}
	20	0.02±0.00 ^a	0.28±0.01 ^a	0.27±0.01 ^a	0.33±0.02 ^a	0.89±0.04 ^a
Source of variance						
CO ₂		NS	NS	NS	NS	NS
N		***	***	NS	**	***
CO ₂ ×N		NS	NS	NS	*	NS

4HGB: 4-hydroxyglucobrassicin; GBS: glucobrassicin; 4MGB: 4-methoxyglucobrassicin; NGBS: neoglucobrassicin. Data followed by the same superscript letter(s) indicate no significant difference at $P<0.05$ level. Values are mean±SD. Significance levels indicated by two-way ANOVA: NS, not significant; * $P<0.05$; ** $P<0.01$; *** $P<0.001$

Table 5 Effect of CO₂ concentration on the carbon (C), nitrogen (N), and sulfur (S) contents, C/N ratio, and N/S ratio in bolting stems of Chinese kale at three N concentrations

CO ₂ (μL/L)	N (mmol/L)	C content (%)	N content (%)	S content (%)	C/N ratio	N/S ratio
350	5	30.88±0.81 ^b	3.58±0.05 ^e	0.81±0.01 ^a	8.62±0.10 ^c	4.42±0.09 ^d
	10	30.87±1.00 ^b	4.44±0.08 ^c	0.85±0.02 ^{ab}	6.95±0.17 ^d	5.21±0.15 ^b
	20	31.81±0.37 ^b	5.06±0.11 ^a	0.83±0.02 ^b	6.29±0.18 ^e	6.09±0.19 ^a
800	5	34.39±0.12 ^a	3.19±0.03 ^f	0.77±0.02 ^b	10.79±0.12 ^a	4.16±0.12 ^e
	10	35.08±0.14 ^a	3.85±0.08 ^d	0.81±0.02 ^c	9.12±0.43 ^b	4.77±0.11 ^c
	20	35.13±0.47 ^a	4.82±0.10 ^b	0.77±0.02 ^c	7.28±0.25 ^d	6.27±0.17 ^a
Source of variance						
CO ₂		***	***	***	***	*
N		NS	***	**	***	***
CO ₂ ×N		NS	*	NS	**	**

Data followed by the same superscript letter(s) indicate no significant difference at $P<0.05$ level. Values are mean±SD. Significance levels indicated by two-way ANOVA: NS, not significant; * $P<0.05$; ** $P<0.01$; *** $P<0.001$

contents at the ambient and elevated CO₂ concentrations did not differ significantly in the 20 mmol N/L treatment.

DISCUSSION

Little is known about the interactive effect of CO₂ enrichment and N availability on GS content in vegetables. In the present study, the effect of an elevated CO₂ concentration at three different N concentrations (5, 10, and 20 mmol/L) on GS content in the bolting stem of Chinese kale was investigated. We observed that the CO₂ concentration and N concentration in the nutrient solution showed significant interactive effects on the height and dry weights of the aerial parts, bolting stems, and roots. The maximum stem thickness and dry weights of the aerial parts,

bolting stems, and roots were obtained at 20 mmol N/L under the elevated CO₂ condition, which confirms the importance of N availability in determining the vegetable's response to elevated CO₂. Similar conclusions have also been reported for the grass *Bromus mollis*, rice, wheat, and tomato seedlings (Larigauderie et al., 1988; Kim et al., 2001; Kobayashi et al., 2001; Li et al., 2007).

In this study, 11 individual GSs were detected in bolting stems of Chinese kale, of which the major GSs were gluconapin, sinigrin, glucoiberin, and glucoraphanin. These results are in agreement with a previous study on Chinese kale (La et al., 2008). Glucoraphanin, sinigrin, glucoiberin, and glucobrassicin are reported to be the most important GSs for the hydrolysis products serving as the most powerful agents protecting human and animal cells against carcinogenesis (Bones and Rossiter, 1996; Fahey et

al., 1997; Nilsson *et al.*, 2006). We detected all the GSs in bolting stem, and the concentrations of sinigrin, glucoiberin, and glucobrassicin were similar to those reported for broccoli (Kushad *et al.*, 1999; Padilla *et al.*, 2007). A high concentration of progoitrin in vegetables is a latent problem because it causes goiter and other harmful effects on animals, such as depressed growth, poor egg production, and liver damage (Heaney and Fenwick, 1995). However, there is no evidence that *Brassica* consumption has any goitrogenic effects on humans (Mithen *et al.*, 2000) and also there is no normative limit issued yet for progoitrin concentration in vegetables. Fortunately, the concentration of progoitrin detected in the edible part of Chinese kale was relatively low, ranging 0.12~0.37 $\mu\text{mol/g DW}$.

The elevated CO_2 concentration increased the total GS concentration as a result of a strong increase in aliphatic GSs, whereas there was no significant effect on the concentrations of indolyl GSs at 5 or 10 mmol N/L, compared with those at the ambient CO_2 concentration. This is in agreement with a previous study on broccoli inflorescences (Schonhof *et al.*, 2007). However, at 20 mmol N/L, the difference between the ambient and elevated CO_2 treatments was not significant.

To our knowledge, no corresponding previous study has reported on the interactive effect of CO_2 and N concentrations on GS content. Bryant *et al.* (1983) advanced the carbon/nutrient balance hypothesis to predict the change of N- and C-containing compounds under elevated CO_2 conditions. These authors pointed out that, under an elevated CO_2 concentration, because of the increased C supply and N limitation, the concentrations of N- and C-containing compounds increased. Studies on the changes in condensed tannin, soluble phenolic polymer, and GS concentrations in the oilseed rape (*Brassica napus*) leaves under CO_2 enrichment are consistent with this hypothesis (Peñuelas and Estiarte, 1998; Himanen *et al.*, 2008). However, some studies of the effect of CO_2 enrichment on GS content are not consistent with the prediction of the carbon/nutrient balance hypothesis (Karowe *et al.*, 1997; Reddy *et al.*, 2004; Schonhof *et al.*, 2007). In the present study, under the elevated CO_2 condition, the C content in bolting stem of Chinese kale increased while the N content decreased in all of the three N treatments, which resulted in an

increase in the C/N ratio; however, GS concentration did not change consistently with the increase in C/N ratio. Clearly, the change in C/N ratio was not a reliable predictive tool for the change in GS composition and concentration in Chinese kale bolting stems.

N and S are not only two essential elements that are constituents of amino acids, but also the main factors that affect GS content in bolting stems of Chinese kale. Under the elevated CO_2 concentration, besides the decrease in N content, S content simultaneously decreased in all of the N treatments. However, in broccoli inflorescences, N content decreased while the S content was unaffected by a rise in CO_2 concentration because of the unchanged fresh and dry weights of broccoli between CO_2 regimes (Schonhof *et al.*, 2007). In young pedunculate oak (*Quercus robur* L.) trees, sulfate uptake was enhanced under CO_2 enrichment (650 $\mu\text{L/L}$) (Seegmüller *et al.*, 1996), which indicated that there are genotypic differences in the sulfate absorption response to elevated atmospheric CO_2 concentration. In the present study, because of the changes in N and S contents, the N/S ratio decreased in the 5 and 10 mmol N/L treatments under the elevated CO_2 concentration, but was not affected significantly by the 20 mmol N/L solution because of the similar change in N and S contents under enriched CO_2 concentration. Hesse *et al.* (2004) reported that decrease in the N/S ratio due to diminished N content caused increased synthesis of sulfurous cysteine as a precursor of methionine. Moreover, cysteine and methionine act as effective sulfur donors in thiohydroxamate formation in the syntheses of aliphatic and indolyl GSs (Mikkelsen *et al.*, 2002). However, in our study, at the 5 and 10 mmol N/L concentrations, the aliphatic GS concentration increased with the decrease in N/S ratio, but indolyl GS concentration was not affected by the reduced N/S ratio under the elevated CO_2 concentration, which agreed with previous results of the effect of elevated CO_2 on the GS content of broccoli inflorescences (Schonhof *et al.*, 2007). Although indolyl GS concentrations were unchanged under the elevated CO_2 concentration, indolyl GS content increased with the increasing N supply, which is in agreement with the findings of Shattuck and Wang (1993) and Kim *et al.* (2002).

Besides the effect of N on GS concentration and composition, plant species is thought to be taken into

account as they experience increasing atmospheric CO₂ levels. For broccoli, which is a cultivated derivative of *B. oleracea*, aliphatic GS concentration increased while indole GS concentration decreased under elevated CO₂ conditions (Schonhof *et al.*, 2007). In a recent study, elevated CO₂ (720 µl/L) increased the concentrations of total aliphatic GSs and aromatic GSs and decreased the indole GS concentration in leaves of both transgenic and wild-type oilseed rapeseed (*Brassica rapa* subsp. *oleifera*) (Himanen *et al.*, 2008). However, in *Arabidopsis thaliana* CO₂ enrichment did not significantly influence the GS concentration (Bidart-Bouzat *et al.*, 2005). Karowe *et al.* (1997) suggested that responses in GS content to elevated CO₂ concentration appeared to be species-specific, as in the same experiment total GS concentrations in both young and old mustard (*Brassica juncea*) leaves decreased, whereas GS concentrations in radish (*Raphanus sativus* L.) and turnip (*Brassica rapa* subsp. *rapa*) appeared to be unaffected by CO₂ enrichment (724 µl/L). Furthermore, Reddy *et al.* (2004) and Bidart-Bouzat *et al.* (2005) observed that the significant changes in individual GS concentrations were not consistent among cultivars of oilseed rape and *Arabidopsis thaliana* under elevated CO₂ conditions, supporting the hypothesis that, in general, the response to elevated CO₂ differs among cultivars. Moreover, in Chinese kale bolting stems, the concentrations of aliphatic GS glucorucin and all individual indolyl GSs were not affected by the elevated CO₂ concentration, indicating that the concentrations of individual GSs within a specific GS group (i.e., aliphatic, aromatic, or indolyl) did not change consistently, which agreed with the conclusion that the response to elevated CO₂ also depends on the individual GS type (Bidart-Bouzat *et al.*, 2005).

CONCLUSION

The elevated CO₂ concentration promoted the growth of Chinese kale in the three N treatments and a high N concentration is beneficial for the growth of Chinese kale. GS concentration was affected by CO₂ concentration, N concentration, and CO₂×N interactions. Under the elevated CO₂ concentration, total GS concentration increased as a result of the increase in aliphatic GS concentration in the 5 and 10 mmol N/L

treatments, but there was no significant difference in the 20 mmol N/L treatment, compared with GS concentration under the ambient CO₂ concentration. The maximum total GS concentration was recorded in the 5 mmol N/L treatment under the elevated CO₂ concentration. Because of the increase in C content and decrease in N and S contents, the C/N ratio was increased significantly at each N level and N/S ratio decreased significantly in the 5 and 10 mmol N/L treatments. However, changes in the C/N ratio were not a reliable predictor of changes in GS concentration in Chinese kale bolting stems. Changes in N and S contents and the N/S ratio could contribute to the change in GS concentration in the bolting stem of Chinese kale. These results indicate that at an elevated CO₂ concentration high N availability promoted the growth of Chinese kale, but reduced the total GS content in bolting stems.

References

- Ahmad, G., Jan, A., Arif, M., Jan, M.T., Khattak, R.A., 2007. Influence of nitrogen and sulfur fertilization on quality of canola (*Brassica napus* L.) under rainfed conditions. *J. Zhejiang Univ. Sci. B*, **8**(10):731-737. [doi:10.1631/jzus.2007.B0731]
- Aires, A., Rosa, E., Carvalho, R., 2006. Effect of nitrogen and sulfur fertilization on glucosinolates in the leaves and roots of broccoli sprouts (*Brassica oleracea* var. *italica*). *J. Sci. Food Agric.*, **86**(10):1512-1516. [doi:10.1002/jsfa.2535]
- Amthor, J.S., 2001. Effects of atmospheric CO₂ concentration on wheat yield: review of results from experiments using various approaches to control CO₂ concentration. *Field Crops Res.*, **73**(1):1-34. [doi:10.1016/S0378-4290(01)00179-4]
- Baik, H.Y., Juvik, J.A., Jeffery, E.H., Wallig, M.A., Kushad, M., Klein, B.P., 2003. Relating glucosinolate content and flavor of broccoli cultivars. *J. Food Sci.*, **68**(3):1043-1050. [doi:10.1111/j.1365-2621.2003.tb08285.x]
- Baxter, R., Ashenden, T.W., Farrar, J.F., 1994. Effects of elevated carbon dioxide on three grass species from montane pasture. II. Nutrient uptake, allocation and efficiency of use. *J. Exp. Bot.*, **45**(9):1267-1278. [doi:10.1093/jxb/45.9.1267]
- Bazzaz, F.A., 1990. The response of natural ecosystems to the rising global CO₂ levels. *Annu. Rev. Ecol. Syst.*, **21**(1):167-196. [doi:10.1146/annurev.es.21.110190.001123]
- Bidart-Bouzat, M.G., Mithen, R., Berenbaum, M.R., 2005. Elevated CO₂ influences herbivory-induced defense responses of *Arabidopsis thaliana*. *Oecologia*, **145**(3):415-424. [doi:10.1007/s00442-005-0158-5]
- Bones, A.M., Rossiter, J.T., 1996. The myrosinase glucosinolates system, its organization and biochemistry. *Plant*

- Physiol.*, **97**(1):194-208. [doi:10.1111/j.1399-3054.1996.tb00497.x]
- Bryant, J.P., Chapin, F.S., Klein, D.R., 1983. Carbon/nutrient balance of boreal plants in relation to vertebrate herbivory. *Oikos*, **40**(3):357-368. [doi:10.2307/3544308]
- Caswell, H., 2004. *Advances in Ecological Research*. Academic Press, New York.
- Chen, R.Y., Liu, H.C., Song, C.Z., Sun, G.W., 2005. Effect of nitrogen nutrient on the growth and quality of Chinese kale. *Trans. CSAE*, **21**(S):143-146 (in Chinese).
- Chew, F.S., 1988. *Biologically Active Natural Products*. American Chemical Society Symposium, Washington DC.
- Cotrufo, M.F., Ineson, P., Scott, A., 1998. Elevated CO₂ reduces the nitrogen concentration of plant tissues. *Glob. Change Biol.*, **4**(1):43-54. [doi:10.1046/j.1365-2486.1998.00101.x]
- Das, M., Pal, M., Zaidi, P.H., Raj, A., Sengupta, U.K., 2002. Stage sensitivity of mung bean (*Vigna radiata* L. Wilczek) to an elevated level of carbon dioxide. *J. Agron. Crop Sci.*, **188**(4):219-224. [doi:10.1046/j.1439-037X.2002.00556.x]
- Epron, D., Liozon, R., Mousseau, M., 1996. Effects of elevated CO₂ concentration on leaf characteristics and photosynthetic capacity of beech (*Fagus sylvatica*) during the growing season. *Tree Physiol.*, **16**:425-432.
- Fahey, J.W., Zhang, Y., Talalay, P., 1997. Broccoli sprouts: an exceptionally rich source of inducers of enzymes that protect against chemical carcinogens. *Proc. Natl. Acad. Sci. USA*, **94**(19):10367-10372. [doi:10.1073/pnas.94.19.10367]
- Fenwick, G.R., Heaney, R.K., Mullin, W.J., 1983. Glucosinolates and their breakdown products in food and food plants. *CRC Crit. Rev. Food Sci. Nutr.*, **18**(2):123-201.
- Fenwick, G.R., Heaney, R.K., Mawson, R., 1989. *Toxicants of Plant Origin*. CRC Press, Florida.
- Gijzen, M., McGregor, I., Seguin-Swartz, G., 1989. Glucosinolate uptake by developing rapeseed embryos. *Plant Physiol.*, **89**(1):260-263. [doi:10.1104/pp.89.1.260]
- Habash, D.Z., Paul, M.J., Parry, M.A., Keys, A.J., Lawlor, D.W., 1995. Increased capacity for photosynthesis in wheat grown at elevated CO₂: the relationship between electron-transport and carbon metabolism. *Planta*, **197**(3):482-489. [doi:10.1007/BF00196670]
- Halkier, B.A., Du, L., 1997. The biosynthesis of glucosinolates. *Trends Plant Sci.*, **2**(11):425-431. [doi:10.1016/S1360-1385(97)90026-1]
- He, H.J., Chen, H., Schnitzler, W.H., 2002. Glucosinolate composition and contents in *Brassica* vegetables. *Sci. Agric. Sinica*, **35**(2):192-197 (in Chinese).
- Heaney, R.K., Fenwick, G.R., 1995. Natural toxins and protective factors in *Brassica* species, including rapeseed. *Nat. Toxins*, **3**(4):233-237. [doi:10.1002/nt.2620030412]
- Hesse, H., Nikiforova, V., Gakiere, B., Hoefgen, R., 2004. Molecular analysis and control of cysteine biosynthesis: integration of nitrogen and sulphur metabolism. *J. Exp. Bot.*, **55**(401):1283-1292. [doi:10.1093/jxb/erh136]
- Himanen, S.J., Nissinen, A., Auriola, S., Poppy, G.M., Stewart, C.N., Holopainen, J.K., Nerg, A.M., 2008. Constitutive and herbivore-inducible glucosinolate concentrations in oilseed rape (*Brassica napus*) leaves are not affected by *Bt CryIAc* insertion but change under elevated atmospheric CO₂ and O₃. *Planta*, **227**(2):427-437. [doi:10.1007/s00425-007-0629-5]
- Hoagland, D.R., Arnon, D.I., 1938. *The Water Culture Method for Growing Plants Without Soil*, Circ 347. California Agricultural Experiment Station, Berkley.
- Holst, B., Williamson, G., 2004. A critical review of the bioavailability of glucosinolates and related compounds. *Nat. Prod. Rep.*, **21**(3):425-447. [doi:10.1039/b204039p]
- IPCC (Intergovernmental Panel on Climate Change), 2007. *Climate Change 2007*. Cambridge University Press, Cambridge.
- Islam, M.S., Matsui, T., Yoshida, Y., 1996. Effect of carbon dioxide enrichment on physico-chemical and enzymatic changes in tomato fruits at various stages of maturity. *Sci. Hortic.*, **65**(2-3):137-149. [doi:10.1016/0304-4238(95)00867-5]
- Karowe, D.N., Seimens, D.H., Mitchell-olds, T., 1997. Species-specific response of glucosinolate content to elevated atmospheric CO₂. *J. Chem. Ecol.*, **23**(11):2569-2582. [doi:10.1023/B:JOEC.0000006667.81616.18]
- Kiddle, G., Bennett, R.N., Botting, N.P., Davidson, N.E., Robertson, A.A.B., Wallsgrove, R.M., 2001. High performance liquid chromatography separation of natural and synthetic desulfoglucosinolates and their chemical validation by spectroscopic, NMR and CI-MS methods. *Phytochem. Anal.*, **12**(4):226-242. [doi:10.1002/pca.589]
- Kim, H.Y., Lieffering, M., Miura, S., Kobayashi, K., Okada, M., 2001. Growth and nitrogen uptake of CO₂-enriched rice under field conditions. *New Phytol.*, **150**(2):223-229. [doi:10.1046/j.1469-8137.2001.00111.x]
- Kim, S.J., Matsuo, T., Watanabe, M., Watanabe, Y., 2002. Effect of nitrogen and sulphur application on the glucosinolate content in vegetable turnip rape (*Brassica rapa* L.). *Soil Sci. Plant Nutr.*, **48**(1):43-49.
- Kimball, B.A., Pinter, J.P.J., Garcia, R.L., LaMorte, R.L., Wall, G.W., Hunsaker, D.J., Wechsung, G., Wechsung, F., Kartschall, T., 1995. Productivity and water use of wheat under free-air CO₂ enrichment. *Global Change Biol.*, **1**(6):429-442. [doi:10.1111/j.1365-2486.1995.tb00041.x]
- Kimball, B.A., Zhu, J.G., Cheng, L., Kobayashi, K., Bindi, M., 2002. Responses of agricultural crops to free-air CO₂ enrichment. *J. Appl. Ecol.*, **13**(10):1323-1338 (in Chinese).
- Kobayashi, K., Lieffering, M., Kim, H.Y., 2001. *Structure and Function in Agroecosystem Design and Management*. CRC Press, Florida.
- Kohlmeier, L., Su, L., 1997. Cruciferous vegetable consumption and colorectal cancer risk: meta-analysis of the epidemiological evidence. *FASEB J.*, **11**(3):2141.
- Kushad, M.M., Brown, A.F., Kurilich, A.C., Juvik, J.A., Klein, B.P., Wallig, M.A., Jerrery, E.H., 1999. Variation of glucosinolates in vegetable crops of *Brassica oleracea*. *J. Agric. Food Chem.*, **47**(4):1541-1548. [doi:10.1021/jf980985s]

- La, G.X., Fang, P., Li, Y.J., Wang, Y., 2008. Determination of desulpho-glucosinolates in bolting stems of Chinese kale by liquid chromatography-mass spectrometry. *Journal of Zhejiang University (Agric. Life Sci.)*, **34**(5):557-563 (in Chinese). [doi:10.3785/j.issn.1008-9209.2008.05.012]
- Larigauderie, A., Hilbert, D.W., Oechel, W.C., 1988. Effect of CO₂ enrichment and nitrogen availability on resource acquisition and resource allocation in a grass, *Bromus mollis*. *Oecologia*, **77**(4):544-549. [doi:10.1007/BF00377272]
- Li, J., Zhou, J.M., Duan, Z.Q., Du, C.W., Wang, H.Y., 2007. Effect of CO₂ enrichment on the growth and nutrient uptake of tomato seedlings. *Pedosphere*, **17**(3):343-351. [doi:10.1016/S1002-0160(07)60041-1]
- Louda, S., Mole, S., 1991. *Herbivores: Their Interactions with Secondary Plant Metabolites*, 2nd Ed. Academic Press, New York.
- Lu, R.K., 1999. *Analytical Methods of Soil Agro-chemistry*. Chinese Agriculture Science and Technology Press, Beijing (in Chinese).
- Macfarlane-Smith, W.H., Griffiths, D.W., 1988. A time-course study of glucosinolates in the ontogeny of forage rape (*Brassica napus* L.). *J. Sci. Food Agric.*, **43**(2):121-134. [doi:10.1002/jsfa.2740430203]
- Mikkelsen, M.D., Petersen, B., Olsen, C., Halkier, B.A., 2002. Biosynthesis and metabolic engineering of glucosinolates. *Amino Acids*, **22**(3):279-295. [doi:10.1007/s00726020014]
- Mithen, R.F., Dekker, M., Verkerk, R., Rabot, S., Johnson, I.T., 2000. The nutritional significance, biosynthesis and bioavailability of glucosinolates in human foods. *J. Sci. Food Agric.*, **80**(7):967-984. [doi:10.1002/(SICI)1097-0010(20000515)80:7<967::AID-JSFA597>3.0.CO;2-V]
- Mooney, H.A., Drake, B.G., Luxmoore, R.J., Oechel, W.C., Pitelka, L.F., 1991. Predicting ecosystem responses to elevated CO₂ concentrations. *Bioscience*, **41**(2):96-104. [doi:10.2307/1311562]
- Nilsson, J., Olsson, K., Engqvist, G., Ekvall, J., Olsson, M., Nyman, M., Kesson, B., 2006. Variation in the content of glucosinolates, hydroxycinnamic acids, carotenoids, total antioxidant capacity and low-molecular-weight carbohydrates in *Brassica* vegetables. *J. Sci. Food Agric.*, **86**(4):528-538. [doi:10.1002/jsfa.2355]
- Padilla, G., Cartea, M.E., Velasco, P., de Haro, A., Prdás, A., 2007. Variation of glucosinolates in vegetable crop of *Brassica rapa*. *Phytochemistry*, **68**(4):536-545. [doi:10.1016/j.phytochem.2006.11.017]
- Peñuelas, J., Estiarte, M., 1998. Can elevated CO₂ affect secondary metabolism and ecosystem function? *Trends Ecol. Evol.*, **13**(1):20-24. [doi:10.1016/S0169-5347(97)01235-4]
- Price, K.R., Casuscelli, F., Colquhoun, I.J., Rhodes, M.J.C., 1998. Composition and content of flavonol glycosides in broccoli florets (*Brassica oleracea*) and their fate during cooking. *J. Sci. Food Agric.*, **77**(4):468-472. [doi:10.1002/(SICI)1097-0010(199808)77:4<468::AID-JSFA66>3.0.CO;2-B]
- Pruden, G., Kalembasa, S.J., Jenkinson, D.S., 1985. Reduction of nitrate of prior to Kjeldahl digestion. *J. Sci. Food Agric.*, **36**(2):71-73. [doi:10.1002/jsfa.2740360203]
- Reddy, G.V.P., Tossavainen, P., Nerg, A.M., Holopainen, J.K., 2004. Elevated atmospheric CO₂ affects the chemical quality of *Brassica* plants and the growth rate of the specialist *Plutella xylostella*, but not the generalist, *Spodoptera littoralis*. *J. Agric. Food Chem.*, **52**(13):4185-4191. [doi:10.1021/jf049358v]
- Rodman, J.E., 1991. A taxonomic analysis of glucosinolate producing plants, part 1: Phenetics. *Syst. Bot.*, **16**(4):598-618. [doi:10.2307/2418864]
- Rosa, E., Heaney, R.K., Fenwick, G.R., Portas, C.A.M., 1997. Glucosinolates in crop plants. *Hort. Rev.*, **19**:99-215.
- Schnug, E., 1989. Double low oilseed rape in West Germany, sulphur nutrition and glucosinolate levels. *Aspects Appl. Biol.*, **23**:67-82.
- Schonhof, I., Kläring, H.P., Krumbein, A., Schreiner, M., 2007. Interaction between atmospheric CO₂ and glucosinolates in Broccoli. *J. Chem. Ecol.*, **33**(1):105-114. [doi:10.1007/s10886-006-9202-0]
- Schreiner, M., 2005. Vegetable crop management strategies to increase the quantity of phytochemicals. *Eur. J. Nutr.*, **44**(2):85-94. [doi:10.1007/s00394-004-0498-7]
- Seegmüller, S., Schulte, M., Herschbach, C., Rennenberg, H., 1996. Interactive effects of mycorrhization and elevated atmospheric CO₂ on sulphur nutrition of young pedunculate oak (*Quercus robur* L.) trees. *Plant Cell Environ.*, **19**(4):418-426. [doi:10.1111/j.1365-3040.1996.tb00333.x]
- Shattuck, V.I., Wang, W., 1993. Nitrogen dioxide fumigation alters the glucosinolate and nitrate levels in pak choy (*Brassica campestris* ssp. *Chinensis*). *Sci. Horticul.*, **56**(2):87-100. [doi:10.1016/0304-4238(93)90010-N]
- Tawfiq, N., Heaney, R.K., Pulumb, J.A., Fenwick, G.R., Musk, S.R., Williamson, G., 1995. Dietary glucosinolates as blocking agents against carcinogenesis: glucosinolate breakdown products assessed by induction of quinine reductase activity in murine hepalc1c7 cells. *Carcinogenesis*, **16**(5):1191-1194. [doi:10.1093/carcin/16.5.1191]
- Wattenberg, L.W., 1993. *Food and Cancer Prevention: Chemical and Biological Aspects*. Royal Society of Chemistry, London.
- Zhao, F., Evans, E.J., Bilsborrow, P.E., Schnug, E., Syers, J.K., 1992. Correction for protein content in the determination of the glucosinolate content of rapeseed by the XRF method. *J. Sci. Food Agric.*, **58**(3):431-433. [doi:10.1002/jsfa.2740580319]
- Zhao, F., Evans, E.J., Bilsborrow, P.E., Schnug, E., Syers, J.K., 1994. Influence of nitrogen and sulphur on the glucosinolate profiles of rapeseed (*Brassica napus* L.). *J. Sci. Food Agric.*, **64**(3):295-304. [doi:10.1002/jsfa.2740640309]