



Leucine and histidine independently regulate milk protein synthesis in bovine mammary epithelial cells via mTOR signaling pathway*

Hai-na GAO^{1,2,3,4}, Han HU^{1,3,4}, Nan ZHENG^{†‡1,3,4}, Jia-qi WANG^{1,3,4}

(¹Ministry of Agriculture-Milk Risk Assessment Laboratory, Institute of Animal Science, Chinese Academy of Agricultural Sciences, Beijing 100193, China)

(²College of Animal Science and Technology, Gansu Agricultural University, Lanzhou 730070, China)

(³Ministry of Agriculture-Milk and Dairy Product Inspection Center, Beijing 100193, China)

(⁴State Key Laboratory of Animal Nutrition, Institute of Animal Science, Chinese Academy of Agricultural Sciences, Beijing 100193, China)

†E-mail: zhengnan_1980@126.com

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Abstract: The aim of this study is to investigate the effects of leucine (Leu) and histidine (His) on the expression of both the mammalian target of rapamycin (mTOR) signaling pathway-related proteins and caseins in immortalized bovine mammary epithelial cells (CMEC-H), using a single supplement through Western blotting. The Earle's balanced salt solution (EBSS) was set as the control group and other treatment groups, based on the EBSS, were added with different concentrations of Leu or His, respectively. The results showed that, compared with the control group, the expression of caseins and the phosphorylation of mTOR (Ser²⁴⁸¹), Raptor (Ser⁷⁹²), eIF4E (Ser²⁰⁹), and eEF2 (Thr⁵⁶) increased with the Leu concentrations ranging from 0.45 to 10.80 mmol/L ($P < 0.01$). The P-4EBP1 (Thr³⁷) at 10.80 mmol/L Leu, and P-RPS6 (Ser^{235/236}) at 5.40 to 10.80 mmol/L Leu all decreased. Similarly, the His supplementation from 0.15 to 9.60 mmol/L increased the expression of α s2-casein, β -casein, κ -casein, P-mTOR (Ser²⁴⁸¹), P-Raptor (Ser⁷⁹²), P-S6K1 (Thr³⁸⁹), P-4EBP1 (Thr³⁷), P-eIF4E (Ser²⁰⁹), and P-eEF2 (Thr⁵⁶) ($P < 0.01$) in CMEC-H, whereas the α s1-casein expression was only reduced at 9.60 mmol/L His, G protein β subunit-like protein (G β L) at 0.15 and 9.60 mmol/L His, and P-RPS6 at 4.80 to 9.60 mmol/L His. Our linear regression model assay suggested that the α s1-casein expression was positively correlated with P-mTOR ($P < 0.01$), P-S6K1 ($P < 0.01$), and P-eEF2 ($P < 0.01$) for the addition of Leu, while the expressions of β -casein ($P < 0.01$) and κ -casein ($P < 0.01$) were positively correlated with P-eEF2 for the addition of His. In conclusion, the milk protein synthesis was up-regulated through activation of the mTOR pathway with the addition of Leu and His in CMEC-H.

Key words: Bovine mammary epithelial cells, Leucine, Histidine, Western blotting, mTOR, Casein

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1 Introduction

Dairy cow nutrient requirement systems regard essential amino acid (EAA) requirements in aggregate as metabolizable protein (MP), and presume a fixed efficiency of MP use for milk protein (Arriola Apelo *et al.*, 2014b). The composition of EAAs in MP vastly affects milk protein synthesis. Since the exact amount of limiting amino acids (LAAs) in dairy cow's diets was unknown, cows were easily overfed to meet the balance of MP requirements, which leads to a waste of amino acids (AAs) and poor N efficiency

‡ Corresponding author

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ORCID: Hai-na GAO, <http://orcid.org/0000-0003-1327-4003>; Nan ZHENG, <http://orcid.org/0000-0002-8469-4243>

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(Bionaz *et al.*, 2012; Arriola Apelo *et al.*, 2014a). In current dairy production systems, an average of 25% of dietary N is captured in milk, which results in approximately 75% of the dietary N being excreted in urine and feces (Hristov *et al.*, 2004; Arriola Apelo *et al.*, 2014b). Feeding animals with low-protein diets (Kalscheur *et al.*, 2006) and supplementation of those deficient EAAs (lysine (Lys), methionine, leucine (Leu), isoleucine, valine, and histidine (His)) may maintain production, which can improve the total N efficiency (Kim *et al.*, 2001; Wang *et al.*, 2010; Appuhamy *et al.*, 2011a; 2012), which is in accordance with Liebig's hypothesis on developing the concept of the order of LAAs. This concept is commonly described by using the analogy of a water barrel with broken staves (Appuhamy *et al.*, 2012). The utilization of AAs is important for mammary protein synthesis in lactating cows (Appuhamy *et al.*, 2012; Arriola Apelo *et al.*, 2014c). Previous research on AA metabolism primarily focused on optimizing balanced diets, which is essential for maintaining and enhancing milk protein synthesis in dairy cattle (Hanigan *et al.*, 2001).

AAs not only serve as a precursor for protein synthesis but also as signaling molecules that regulate the protein synthesis in mammals (Kimball, 2002; Arriola Apelo *et al.*, 2014c). A great number of reports have shown that AA-induced stimulation of milk protein synthesis is partially mediated by the mammalian target of rapamycin (mTOR), a protein kinase present in the rapamycin-sensitive mTOR complex 1 (mTORC1) (Yang *et al.*, 2006; Prizant and Barash, 2008; Appuhamy *et al.*, 2012; Arriola Apelo *et al.*, 2014c). mTORC1 consists of mTOR, regulatory associated protein of TOR (Raptor), and a G protein β subunit-like protein (G β L) (Kim, 2009). A great deal of evidence indicated that P-mTOR at Ser²⁴⁴⁸ was an indicator of mTOR pathway activity in immortalized bovine mammary epithelial cells (MAC-T) (Appuhamy *et al.*, 2011b; 2012). Prizant and Barash (2008) showed that P-mTOR in Ser²⁴⁴⁸ was completely inhibited by His supplementation, but phosphorylation of the ribosomal protein S6 kinase 1 (S6K1) and eukaryotic initiation factor 4E binding protein (4EBP1) could still be detected. AA supplementation may have contributed to other existent phosphorylation sites of mTOR that were not detected, resulting in phosphorylation of downstream effectors. Peterson *et al.* (2000) and Gerasimovskaya *et al.*

(2005) demonstrated that the P-mTOR at Ser²⁴⁸¹ was also a good biomarker for mTOR pathway activation. Cheng *et al.* (2004) demonstrated that the P-mTOR in Thr²⁴⁴⁶ acted as a nutrient-regulated phosphorylation site located in the mTOR catalytic domain. We cannot exclude the possibility that phosphorylation of mTOR in Ser²⁴⁴⁸ was important, yet it was not the only one regulating mTOR activity in bovine mammary epithelial cells (CMECs). Kim *et al.* (2002) confirmed that P-Raptor is associated with mTOR and modulated by AAs, which regulate the mTOR kinase activity. When activated by AAs, mTORC1 in turn catalyzed S6K1 and 4EBP1 phosphorylation, followed by phosphorylating the ribosomal protein S6 (RPS6) (Wang and Proud, 2006). S6K1 was a target of mTOR activated in Thr³⁸⁹ whose phosphorylation status was a main indicator of the AA-induced mTORC1-activated signaling pathway (Moshel *et al.*, 2006; Prizant and Barash, 2008; Appuhamy *et al.*, 2012; Arriola Apelo *et al.*, 2014c). In addition, phosphorylation of 4EBP1 by mTOR increased the initial reaction rates (Beugnet *et al.*, 2003). When 4EBP1 was activated, the expression of a regulatory protein binding into the mRNA cap-binding protein eukaryotic initiation factor 4E (eIF4E) was also promoted. When 4EBP1 was unphosphorylated, cap-dependent translation was inhibited by competitively blocking the binding of eukaryotic translation initiation factor 4G (eIF4G) to eIF4E (Gingras *et al.*, 1999; Harris and Lawrence, 2003). The elongation step of mRNA translation was enhanced by eukaryotic elongation factor 2 (eEF2) (Wang and Proud, 2006) and was inhibited by its phosphorylation on Thr⁵⁶, which suggested that eEF2 may be a limiting factor in milk protein synthesis (Christophersen *et al.*, 2002; Appuhamy *et al.*, 2011b).

These studies have shown that Leu and His in addition to Lys and Met are main LAAs in forage diets (Kim *et al.*, 2001; Korhonen *et al.*, 2002). Leu, a branched chain amino acid (BCAA), is currently the most prevalent EAA in the diet (Korhonen *et al.*, 2002; Appuhamy *et al.*, 2012). According to previous reports, Leu was involved in cell growth and differentiation through the mTOR pathway (Proud, 2007; Stipanuk, 2007). Moreover, Leu produced a dose-dependent regulation of mTOR and served as a key intracellular regulatory factor of the mTORC1 pathway in CMECs (Moshel *et al.*, 2006; Prizant and Barash,

2008; Appuhamy *et al.*, 2012; Durán and Hall, 2012).

Kim *et al.* (2001) found that His was the first limiting factor for milk protein secretion. Some traditional experiments indicated that the perfusion of His promoted milk protein synthesis (Bequette *et al.*, 2000). Likewise, some cell experiments demonstrated that the addition of His activated the mTOR pathway, which up-regulated phosphorylation of the downstream protein and ultimately promoted milk protein synthesis (Appuhamy *et al.*, 2012). Prizant and Barash (2008) also reported that His was a key intracellular regulatory factor of the mTORC1 pathway in L-1 cells, a cloned cell line derived from lactating bovine mammary gland, however, negatively regulated P-S6K1, and decreased the β -casein synthesis rate. So, it is meaningful to explore the mechanism of how His influenced the mTOR pathway.

We hypothesize that AAs affect milk protein synthesis in CMEC-H through phosphorylation of translation factors of the mTOR signaling pathway. Although the effects of EAAs on mTOR cell signaling and milk protein synthesis have been extensively explored (Moshel *et al.*, 2006; Burgos and Cant, 2010; Toerien *et al.*, 2010), effects of different levels of Leu or His on phosphorylation of Raptor on Ser⁷⁹², mTOR on Ser²⁴⁸¹, and G β L expression involved in casein protein synthesis were not clear in bovine mammary cells. Our study investigated the regulatory effects of different concentrations of Leu and His on the mTOR signaling pathway and the relationships between these signaling protein activities with milk protein synthesis.

2 Materials and methods

In the present study, all animal care and experimental procedures were approved and conducted under the established standards of the Institute of Animal Science, Chinese Academy of Agricultural Sciences, Beijing, China.

Unless otherwise stated, all components were purchased from the Sigma-Aldrich Shanghai Trading Co. Ltd. (Shanghai, China). The Dulbecco's modified Eagle's medium (DMEM)/Ham's F12 (diluted 1:1 (v/v), 1439945/1491066, Invitrogen Trading Co. Ltd., China) was used as the basal growth medium, supplemented with 10% fetal bovine serum (FBS)

(Invitrogen Trading Co. Ltd., China) and 100 μ g/ml penicillin-streptomycin solution (C0222, Beyotime Institute of Biotechnology, China). The differentiation medium contained bovine insulin, progesterone, bovine holo-transferrin, hydrocortisone, bovine epithelial growth factor, and bovine estradiol (Sigma-Aldrich, Cat. Nos. I4434, T1283, P8783, H0888, E4127, and E2758, respectively).

Individual AAs (L-Leu and L-His, Cat. Nos. L8912 and H5659, respectively) were purchased from the Sigma-Aldrich Shanghai Trading Co. Ltd., China. The non- and site-specific phosphorylated antibodies against mTOR (Ser²⁴⁸¹, YT2913/YP1134), 4EBP1 (Thr³⁷, YT0018/YP0001), RPS6 (Ser^{235/236}, YT4139/YP0832), and eIF4E (Ser²⁰⁹, YT1516/YP0094) were purchased from Immuno Way. The total and site-specific phosphorylated antibodies against Raptor (Ser⁷⁹², No. 2280/2083), S6K1 (Thr³⁸⁹, No. 9202/9205), eEF2 (Thr⁵⁶, No. 2332/2331), and G β L (No. 3274) were purchased from Cell Signaling Technology (Danvers, MA, USA).

The α s1-casein antibody (SAB1401093) and κ -casein antibody (SAB1401094) were purchased from Sigma-Aldrich Shanghai Trading Co., Ltd. China, the α s2-casein antibody (bs-10034R) was purchased from Bioss (Beijing Biosynthesis Biotechnology Co., Ltd., China) and the β -casein antibody (orb18512) was purchased from Biorbyt (Cambridge, UK). β -Actin (ab8226), used as a loading control, was purchased from Abcam Trading (Shanghai) Company Ltd. (China).

2.1 Cell culture and experimental design

The immortalized bovine mammary epithelial cell line (CMEC-H) was established in our previous work (Hu *et al.*, 2014). The cells were cultured at 37 °C with 5% CO₂ and digested with 0.25% (2.5 g/L) trypsin-0.02% etilendiaminetetraacetic acid (EDTA) (C0203, Beyotime Institute of Biotechnology, China).

The Earle's balanced salt solution (EBSS, CC0043, Leagene Biotechnology, China) was used as a substitute of medium without AAs (Table 1), and was set as a negative control in the experimental design. The EBSS was supplemented with 10% FBS and was used as a positive control. The concentrations of Leu and His in the DMEM/F12 medium were 0.45 and 0.15 mmol/L, respectively. EAAs were added according to the concentrations described in Tables 2

and 3. The CMEC-H cells were cultured in 9-cm plates (172958, Thermo Scientific, China) to 80% confluence and induced to be differentiated by the differentiation medium for 24 h. The differentiated CMEC-H cells were incubated in serum-free, complete DMEM/F12 media overnight. Leu or His was supplemented individually for 6 h at the indicated concentrations. Each experiment was triplicated and each of the whole experiments was repeated three times on three different days.

Table 1 Composition of the Earle's balanced salt solution (EBSS) medium *

Component	Concentration (mmol/L)
Anhydrous CaCl ₂	1.050
CuSO ₄ ·5H ₂ O	5.2×10 ⁻⁶
Fe(NO ₃) ₃ ·9H ₂ O	1.238×10 ⁻⁴
FeSO ₄ ·7H ₂ O	0.002
Anhydrous MgCl ₂	0.301
Anhydrous MgSO ₄	0.407
KCl	4.157
NaHCO ₃	14.286
NaCl	120.612
Anhydrous Na ₂ HPO ₄	0.500
NaH ₂ PO ₄ ·H ₂ O	0.453
ZnSO ₄ ·7H ₂ O	0.002
D-Glucose (dextrose)	17.506
Phenol red	0.022

* Modified from Gao et al. (2015)

Table 2 Leu levels in the medium for the *in vitro* culture of CMEC-H from dairy cattle

Group	EBSS	Leu (mmol/L)	10% FBS
Negative control	+		
Positive control	+		+
Treatment	+	0.45	
	+	1.35	
	+	5.40	
	+	10.80	

DMEM/F12 base media contained leucine (0.45 mmol/L) at a level sufficient for normal cellular processes. +: add

Table 3 His levels in the medium for the *in vitro* culture of CMEC-H from dairy cattle

Group	EBSS	His (mmol/L)	10% FBS
Negative control	+		
Positive control	+		+
Treatment	+	0.15	
	+	1.20	
	+	4.80	
	+	9.60	

DMEM/F12 base media contained histidine (0.14999 mmol/L) at a level sufficient for normal cellular processes. +: add

2.2 Protein immunoblot analysis

The cell samples were homogenized in an ice-cold radio immunoprecipitation assay (RIPA) buffer (P0013, Beyotime Institute of Biotechnology, China) containing 1 mmol/L phenylmethylsulfonyl fluoride (PMSF), 1% (v/v) protease, and a phosphatase inhibitor cocktail (78430, Thermo Scientific). Cell lysates were centrifuged at 2000×g for 3 min and the supernatants were transferred to new tubes. The protein concentrations of the samples were determined by bicinchoninic acid (BCA) assay kits (P0010S, Beyotime Institute of Biotechnology, China). The lysates (30 μg of protein) were loaded onto gels, separated by 10% sodium dodecyl sulfate polyacrylamide gel electrophoresis (SDS-PAGE), and blotted onto polyvinylidene fluoride (PVDF) transfer membranes (0.45 and 0.2 μmol/L immobilon-P, IPVH00010 and ISEQ00010, Millipore). The membranes were blocked for 1 h (blot-PO buffer, WBAVDP001, Millipore) and incubated overnight at 4 °C in primary antibodies (appropriately diluted 1:1000 in phosphate-buffered saline (PBS)) against total and site-specific phosphorylated mTOR (Ser²⁴⁸¹), 4EBP1 (Thr³⁷), RPS6 (Ser^{235/236}), eIF4E (Ser²⁰⁹), Raptor (Ser⁷⁹²), S6K1 (Thr³⁸⁹), eEF2 (Thr⁵⁶), and GβL. The amounts of synthesized milk proteins were determined by antibodies against α1-casein (1:1000), α2-casein (1:400), β-casein (1:500), and κ-casein (1:1000). The blots were washed three times with Tris-buffered saline containing 0.02% (v/v) Tween-20 (TBST), and then incubated with a second antibody (anti-rabbit IgG, A9169, Sigma; anti-goat IgG, API06P, Millipore; anti-Mouse IgG, A9044, Sigma) and diluted at 1:5000 in PBS.

Immunoreactive bands were visualized by Pierce ECL Western Blotting Substrate kits (32106, Thermo Scientific) according to the manufacturer's instructions and were then exposed to X-ray film. The protein band densities were determined by the Image J2x 2.1.4.7 Analyzer (Rawak Software, Inc., Germany). The relative values from the AA-supplemented cells were related to their AA-deprived controls.

2.3 Statistical analysis

At least three independent experiments were done with identical results and all data were tested using the Duncan's test for post-hoc multiple comparisons of treatment means by SAS software (SAS

9.2 Inst., Cary, NC, USA). Differences between experimental groups were considered significant at a P -value of <0.05 . The random effect in the one-way analysis of variance (ANOVA) model was replicated and the fixed effect was the levels of Leu and His. Meaningful relationships among the phosphorylation state of the signaling proteins and casein protein expression in CMEC-H were quantified with simple linear regression models using the REG procedure of SAS.

3 Results

3.1 Effects of Leu or His supplementation on the expression of caseins

Compared with the negative control group, our data demonstrated that the addition of Leu to EBSS

generated dose-responsive effects on the casein expression. Figs. 1a and 1b show that Leu had stimulatory effects on the casein expression in the range of 0.45 to 10.80 mmol/L. Specific to each casein, α 1-casein, α 2-casein, β -casein, and κ -casein reached their highest expression at 0.45, 1.35, 5.40, and 10.80 mmol/L Leu with an increase of 260%, 66%, 150%, and 270%, respectively.

Figs. 1c and 1d show the effects of His supplementation on the casein expression. Compared with the negative control, the expression of casein was all promoted by His in the range of 0.15 to 9.60 mmol/L, except that α 1-casein expression was decreased at 9.6 mmol/L His. In addition, each casein, α 1-casein, α 2-casein, β -casein, and κ -casein reached their highest expression at 4.80, 1.20, 0.15, and 0.15 mmol/L His with 1.6-, 1.8-, 1.8-, and 1.8-fold changes, respectively.

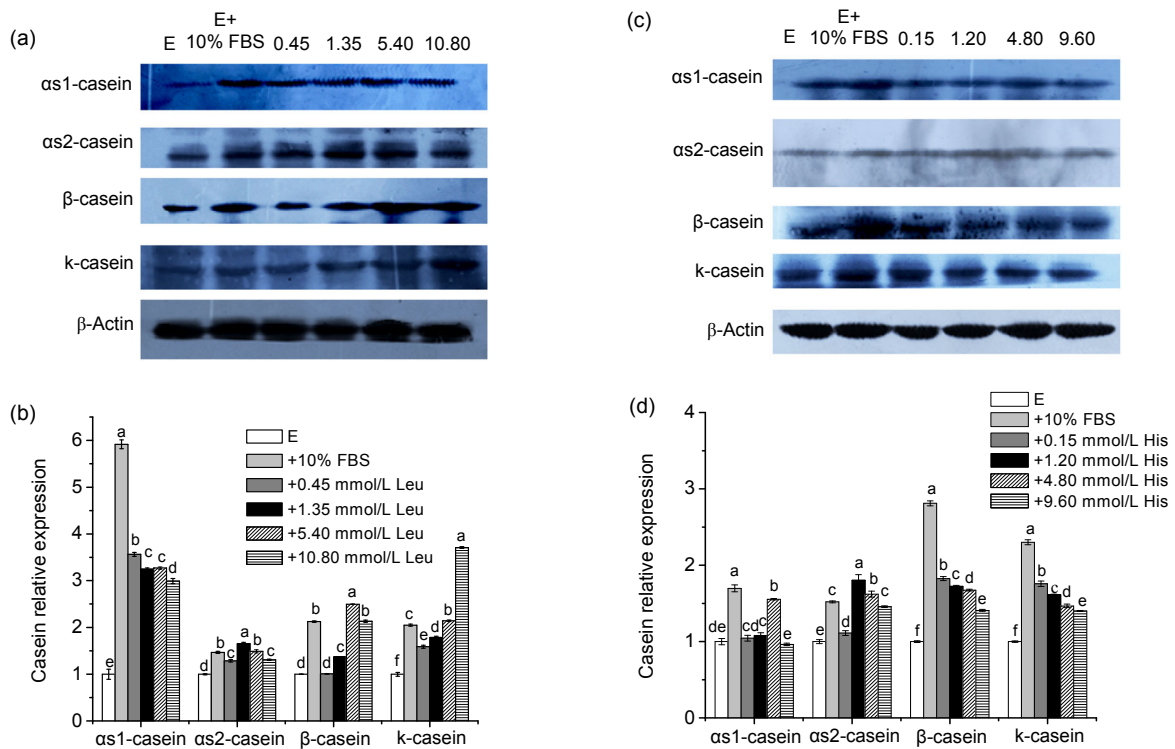


Fig. 1 Effects of Leu and His on the expression of caseins in CMEC-H

(a) Leu was supplemented for 6 h at the indicated concentrations. Casein levels were determined by immunoblot analysis. Numbers above the lanes refer to the levels (mmol/L) of the supplemented Leu relative to that in the Earle's balanced salt solution. (b) Densitometric analysis of signals obtained from the casein immunoblot (a). (c) His was supplemented for 6 h at the indicated concentrations. Casein levels were determined by immunoblot analysis. Numbers above the lanes refer to the levels (mmol/L) of the supplemented His relative to that in the Earle's balanced salt solution. (d) Densitometric analysis of signals obtained from the casein immunoblot (c). β -Actin was assessed as a loading control. A representative blot and quantitation of three independent experiments were shown. In all panels, data represent the mean \pm SD. E: Earle's balanced salt solution; E+10% FBS: Earle's balanced salt solution supplemented with 10% fetal bovine serum. Data in the same concentration marked with different letters represent a significant difference ($P < 0.05$)

3.2 Effects of Leu or His supplementation on the expression of mTORC1

Our results indicated that the Leu supplementation affected the phosphorylation of mTORC1 in a dose-dependent manner. Figs. 2a and 2b show that, compared with the negative control group, only the P-Raptor was markedly increased with the addition of Leu from 0.45 to 10.80 mmol/L. The G β L expression at 10.80 mmol/L Leu and mTOR phosphorylation at 0.45 and 10.80 mmol/L Leu were not significantly changed, and they were significantly enhanced at other concentrations. The changes of mTORC1 all reached their summit at 5.40 mmol/L Leu.

Figs. 2c and 2d show the influence of His supplementation on mTORC1. P-Raptor was dramatically increased from 0.15 to 9.60 mmol/L His. The P-mTOR and G β L expression were slightly enhanced in the

full concentration range, except that the G β L expression was subtly reduced at 0.15 and 9.60 mmol/L His.

3.3 Effects of Leu or His level on the expression of mTOR downstream signaling proteins

Figs. 3a and 3b show that Leu had obvious stimulatory effects on phosphorylation of eEF2 and eIF4E compared to the negative control group, while S6K1 phosphorylation with the addition of Leu was not significantly changed except at 0.45 mmol/L Leu with 2.2-fold changes. For 4EBP1 phosphorylation, its enhancement was observed from 0.45 to 5.40 mmol/L Leu, while 10.80 mmol/L Leu impaired its phosphorylation. Leu supplementation had a minor enhancing effect on the RPS6 phosphorylation level between 0.45 and 1.35 mmol/L, while the phosphorylation level was inhibited over 1.35 mmol/L Leu.

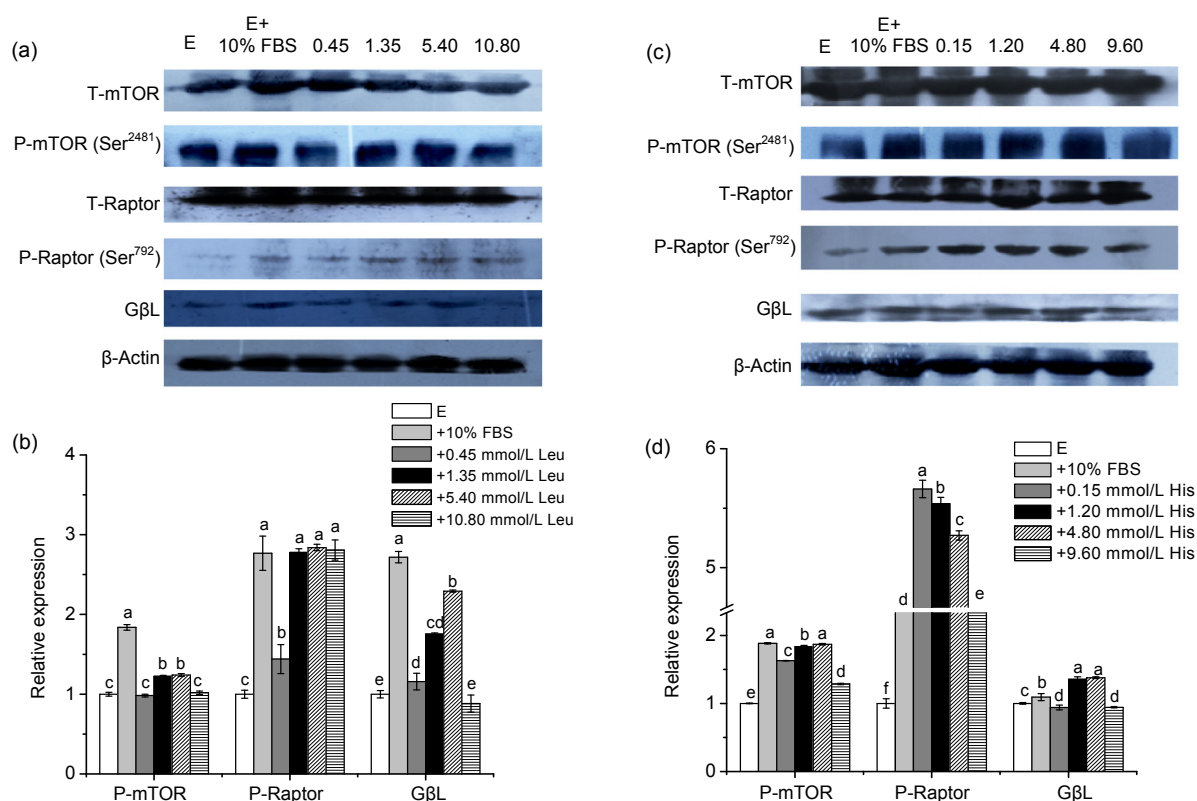


Fig. 2 Effects of Leu and His on the expression of mTORC1 in CMEC-H

(a) Leu was supplemented for 6 h at the indicated concentrations. mTORC1 levels were determined by immunoblot analysis. Numbers above the lanes refer to the levels (mmol/L) of the supplemented Leu relative to that in the Earle's balanced salt solution. (b) Densitometric analysis of signals obtained from the mTORC1 immunoblot (a). (c) His was supplemented for 6 h at the indicated concentrations. mTORC1 levels were determined by immunoblot analysis. Numbers above the lanes refer to the levels (mmol/L) of the supplemented His relative to that in the Earle's balanced salt solution. (d) Densitometric analysis of signals obtained from the mTORC1 immunoblot (c). β -Actin was assessed as a loading control. A representative blot and quantitation of three independent experiments were shown. In all panels, data represent the mean \pm SD. E: Earle's balanced salt solution; E+10% FBS: Earle's balanced salt solution supplemented with 10% fetal bovine serum. Data in the same concentration marked with different letters represent a significant difference ($P < 0.05$)

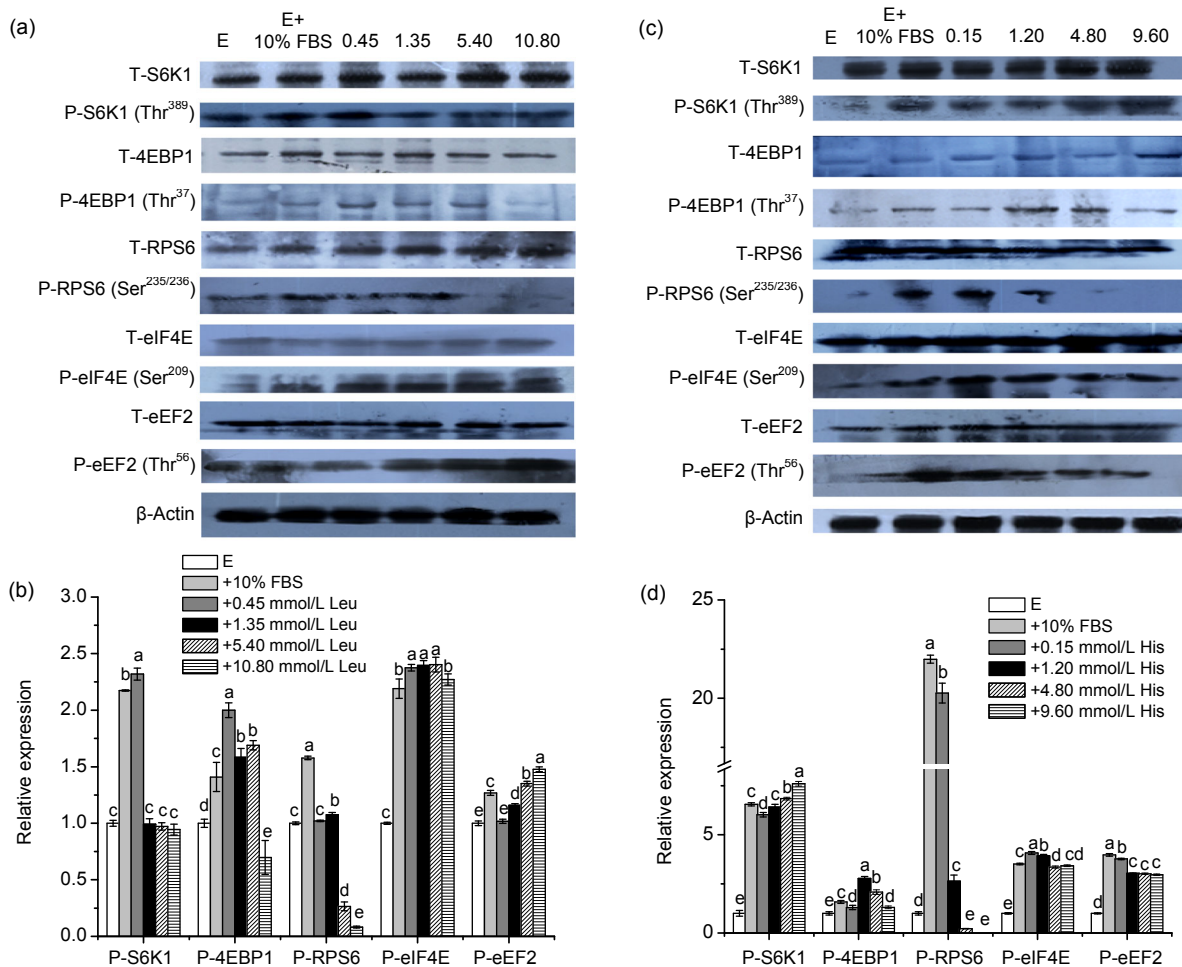


Fig. 3 Effects of Leu and His on the expression of mTOR downstream signaling proteins in CMEC-H (a) Leu was supplemented for 6 h at the indicated concentrations. mTOR downstream signaling protein levels were determined by immunoblot analysis. Numbers above the lanes refer to the levels (mmol/L) of the supplemented Leu relative to that in the Earle's balanced salt solution. (b) Densitometric analysis of signals obtained from the mTOR downstream signaling protein immunoblot (a). (c) His was supplemented for 6 h at the indicated concentrations. mTOR downstream signaling protein levels were determined by immunoblot analysis. Numbers above the lanes refer to the levels (mmol/L) of the supplemented His relative to that in the Earle's balanced salt solution. (d) Densitometric analysis of signals obtained from the mTOR downstream signaling protein immunoblot (c). β -Actin was assessed as a loading control. A representative blot and quantitation of three independent experiments were shown. In all panels, data represent the mean \pm SD. E: Earle's balanced salt solution; E+10% FBS: Earle's balanced salt solution supplemented with 10% fetal bovine serum. Data in the same concentration marked with different letters represent a significant difference ($P < 0.05$)

Figs. 3c and 3d suggest that His supplementation up-regulated all of the studied mTOR downstream proteins in the full concentration range except P-RPS6 at 9.60 mmol/L His and S6K1 phosphorylation which increased in a dose-dependent manner. In addition, at 9.60 mmol/L His, P-S6K1 was nearly 7 times greater than that of the control group. The phosphorylation of 4EBP1 reached its peak ($P < 0.01$) at 1.20 mmol/L His. It was noteworthy that P-RPS6 at 0.15 mmol/L His was 20 times greater than that of the control group.

3.4 Relationships between casein expression and mTOR pathway-related protein changes with Leu supplementation

A simple linear regression model assay demonstrated that α s1-casein expression was positively associated with phosphorylation of mTOR, G β L, Raptor, S6K1, 4EBP1, eEF2, RPS6, and eIF4E ($P < 0.01$), respectively (Table 4). With the addition of Leu, a 1-fold increase in α s1-casein expression was related

to 4.85-, 2.04-, 1.15-, 2.48-, 2.44-, 8.14-, 1.41-, and 2.12-fold changes in mTOR, GβL, Raptor, S6K1, 4EBP1, eEF2, RPS6, and eIF4E phosphorylation, respectively. A notably positive relationship ($R^2=0.7820, 0.5815, 0.7881, 0.7835, \text{ and } 0.6164; P<0.01$) was found between α1-casein expression and mTOR, GβL, S6K1, eEF2 and eIF4E phosphorylation, respectively. No phosphorylation of other mTOR signaling proteins explained the further variation in α1-casein protein expression in CMEC-H (Table 4).

Table 4 Regression analyses for relationships among phosphorylation ratios of signaling proteins and α1-casein, α2-casein, β-casein, κ-casein in bovine mammary epithelial cells cultured in media with Leu for 6 h

Dependent variable	Independent variable	P-value	Regression coefficient	R ²
α1-casein	mTOR	0.0001	4.84738	0.7820
	GβL	0.0001	2.03558	0.5815
	Raptor	0.0052	1.15177	0.3572
	S6K1	0.0001	2.48016	0.7881
	4EBP1	0.0010	2.44002	0.4710
	eEF2	0.0001	8.14314	0.7835
	RPS6	0.0087	1.40988	0.3179
	eIF4E	0.0001	2.11775	0.6164
	α2-casein	mTOR	0.0285	0.41805
GβL		0.0046	0.24768	0.3663
Raptor		0.0001	0.21865	0.6179
S6K1		0.0008	0.29594	0.4823
4EBP1		0.0172	0.28417	0.2630
eEF2		0.0341	0.68184	0.2043
RPS6		0.3133	0.08850	0.0049
eIF4E		0.0021	0.26688	0.4225
β-casein		mTOR	0.0170	1.20726
	GβL	0.0013	0.73002	0.4534
	Raptor	0.0001	0.60842	0.6670
	S6K1	0.0068	0.68103	0.3370
	4EBP1	0.4660	0.25321	-0.0267
	eEF2	0.0188	2.00066	0.2556
	RPS6	0.4630	-0.17444	-0.0262
	eIF4E	0.0013	0.73944	0.4531
	κ-casein	mTOR	0.2831	0.82785
GβL		0.6733	0.15885	-0.0504
Raptor		0.0018	0.71268	0.4338
S6K1		0.0475	0.74591	0.1751
4EBP1		0.2856	-0.52169	0.0128
eEF2		0.2485	1.48963	0.0249
RPS6		0.0626	-0.60010	0.1503
eIF4E		0.0017	1.03420	0.4375

The α2-casein expression was positively correlated with phosphorylation of mTOR, GβL, Raptor, S6K1, 4EBP1, eEF2, and eIF4E ($P<0.05$), respectively (Table 4). A 1-fold increase in α2-casein protein expression was related to 0.42-, 0.25-, 0.22-, 0.30-, 0.28-, 0.68-, and 0.27-fold changes in mTOR, GβL, Raptor, S6K1, 4EBP1, eEF2, and eIF4E phosphorylation, respectively. However, Raptor phosphorylation appeared to be the primary driver when computed in a multivariate model, which explained 61.79% of the observed variation (Table 4).

The β-casein expression was positively associated with phosphorylation of mTOR, GβL, Raptor, S6K1, eEF2, and eIF4E ($P<0.05$), respectively (Table 4). A 1-fold increase in the β-casein protein expression was related to 1.21-, 0.73-, 0.61-, 0.68-, 2.00-, and 0.74-fold changes in mTOR, GβL, Raptor, S6K1, eEF2, and eIF4E phosphorylation, respectively. A notably positive relationship ($R^2=0.6670, P<0.01$) was found between the β-casein expression and P-Raptor, while no correlations were found between the expression of β-casein and phosphorylation of 4EBP1 ($P=0.4660$) or RPS6 ($P=0.4630$) in CMEC-H (Table 4).

The κ-casein expression was positively associated with phosphorylation of Raptor, S6K1, and eIF4E ($P<0.05$), respectively (Table 4). No correlations were found between the κ-casein expression and phosphorylation of mTOR ($P=0.2831$), GβL ($P=0.6733$), 4EBP1 ($P=0.2856$), eEF2 ($P=0.2485$), or RPS6 ($P=0.0626$) in CMEC-H (Table 4).

3.5 Relationships between casein expression and mTOR pathway-related protein changes with His supplementation

The expression of α1-casein was positively correlated with phosphorylation of mTOR, GβL, and eEF2 ($P<0.05$), respectively (Table 5). A 1-fold increase in the α1-casein expression was related to 0.59-, 0.60-, and 0.14-fold changes in mTOR, GβL, and eEF2 phosphorylation, respectively. A notable positive relationship ($R^2=0.5133, P<0.01$) was found between the α1-casein expression and eEF2 phosphorylation.

The α2-casein expression was positively associated with phosphorylation of mTOR, GβL, S6K1, 4EBP1, and eIF4E ($P<0.01$), respectively (Table 5). A 1-fold increase in the α2-casein expression was related to 0.61-, 0.74-, 0.22-, 0.18-, and 0.14-fold changes in mTOR, GβL, S6K1, 4EBP1, and eIF4E

phosphorylation, respectively. A positive relationship ($R^2=0.5790$, $P<0.01$) was found between the α s2-casein expression and P-4EBP1.

The β -casein expression was positively associated with phosphorylation of mTOR, G β L, eEF2, RPS6, and eIF4E ($P<0.05$), respectively (Table 5). A 1-fold increase in the β -casein expression was related to 1.23-, 1.11-, 0.34-, and 0.27-fold changes in mTOR, G β L, eEF2, and eIF4E phosphorylation, respectively. A notable positive relationship ($R^2=0.5341$, 0.9638, and 0.5368, $P<0.01$) was found between the β -casein expression and mTOR, eEF2 and RPS6 phosphorylation, respectively.

Table 5 Regression analyses for relationships among phosphorylation ratios of signaling proteins and α s1-casein, α s2-casein, β -casein, κ -casein in bovine mammary epithelial cells cultured in media with His for 6 h

Dependent variable	Independent variable	P-value	Regression coefficient	R ²
α s1-casein	mTOR	0.0023	0.58689	0.4149
	G β L	0.0234	0.60173	0.2370
	Raptor	0.5879	0.02210	-0.0424
	S6K1	0.1926	0.10670	0.0476
	4EBP1	0.3503	0.05559	-0.0044
	eEF2	0.0005	0.13542	0.5133
	RPS6	0.3110	0.01383	0.0055
	eIF4E	0.1374	0.08256	0.0784
	α s2-casein	mTOR	0.0007	0.60885
G β L		0.0018	0.74109	0.4307
Raptor		0.0598	0.07033	0.1544
S6K1		0.0020	0.21580	0.4267
4EBP1		0.0001	0.17724	0.5790
eEF2		0.1536	0.06191	0.0682
RPS6		0.9829	0.00029	-0.0625
eIF4E		0.0029	0.14346	0.3995
β -casein		mTOR	0.0003	1.22687
	G β L	0.0263	1.10642	0.2271
	Raptor	0.3081	0.07720	0.0063
	S6K1	0.0628	0.27713	0.1500
	4EBP1	0.2005	0.14074	0.0440
	eEF2	0.0001	0.33806	0.9638
	RPS6	0.0003	0.07675	0.5368
	eIF4E	0.0049	0.26810	0.3624
	κ -casein	mTOR	0.0006	0.85113
G β L		0.0544	0.69444	0.1629
raptor		0.1936	0.06931	0.0472
S6K1		0.0501	0.20637	0.1704
4EBP1		0.2484	0.09071	0.0250
eEF2		0.0001	0.23346	0.9048
RPS6		0.0001	0.05838	0.6228
eIF4E		0.0015	0.20877	0.4464

The κ -casein expression was positively associated with phosphorylation of mTOR, eEF2, RPS6, and eIF4E ($P<0.01$), respectively (Table 5). A 1-fold increase in the κ -casein expression was related to 0.85-, 0.23-, and 0.21-fold changes in mTOR, eEF2, and eIF4E phosphorylation, respectively. A notable positive relationship ($R^2=0.5050$, 0.9048, and 0.6228, $P<0.01$) was found between the κ -casein expression and mTOR, eEF2 and RPS6 phosphorylation, respectively.

4 Discussion

In this paper, the functions and potential molecular mechanisms of Leu and His effects on the mTOR signaling pathway in the regulation of milk protein synthesis were studied. In order to eliminate the impact of the other AAs on CMEC-H, all AAs were removed from EBSS and Leu or His was then individually added to it.

In our study, all four forms of caseins were up-regulated by Leu or His supplementation with the exception of 9.60 mmol/L His on α s1-casein, which supported the notion that both Leu and His are key limiting factors for milk protein synthesis.

It is widely accepted that mTOR is a key regulator of milk protein synthesis and most reports were concerned with the role of AAs in the regulation of P-mTOR on Ser²⁴⁴⁸ in milk protein synthesis (Prizant and Barash, 2008; Appuhamy *et al.*, 2012; Arriola Apelo *et al.*, 2014c). Here we identify another mTOR phosphorylation site on Ser²⁴⁸¹ as an indicator of activated mTOR pathway in CMEC-H, which was increased by the addition of Leu or His in the full concentration range. An exception to this was the supplementation of Leu at 0.45 mmol/L with no obvious change. This data indicated that P-mTOR on Ser²⁴⁸¹ could act as an alternative switch to integrate signals from Leu and His to regulate protein translation. The effects of Leu and His supplementation on two other mTORC1 components, Raptor and G β L, were also studied. The association between Raptor and mTOR was manipulated by AAs. Raptor acts as a scaffold protein regulating the assembly, localization, and substrate binding of mTORC1 (Laplante and Sabatini, 2012), and it is also used for the mTOR-catalyzed phosphorylation of 4EBP1 and mediates

TOR action *in vivo* (Hara *et al.*, 2002). With AA deprivation, the mTOR kinase activity is inactive (Kim *et al.*, 2013). The addition of Leu and His activates mTOR kinase to increase the Raptor expression (Kim *et al.*, 2002). GβL also plays a positive role in mTOR activation by AA supplementation. The binding of GβL to the mTOR kinase domain stabilized the interaction between the Raptor and mTOR (Kim *et al.*, 2003). Our results indicated that Raptor notably increased (1.8 and 4.7 times) in comparison with the control group, when added Leu and His, respectively. GβL expression increased in the range of 0.45 and 5.40 mmol/L of Leu with a 2.5-fold change in comparison with the control, whereas His supplementation slightly enhanced GβL expression in the same range. Both GβL and P-mTOR demonstrated the highest changes at 5.40 mmol/L Leu or 4.80 mmol/L His. Previous studies indicated that GβL positively correlated with P-mTOR on Ser²⁴⁴⁸ (Kim *et al.*, 2003), and that there was a strong relationship between GβL and P-mTOR on Ser²⁴⁸¹.

S6K1, a major downstream target of mTOR, serves as an indicator of activation of the mTORC1 pathway (Yang *et al.*, 2006; Prizant and Barash, 2008; Appuhamy *et al.*, 2012; Arriola Apelo *et al.*, 2014c). As most reports showed, 0.45 mmol/L Leu dramatically increased S6K1 phosphorylation.

The effect of Leu concentrations greater than 0.45 mmol/L on S6K1 phosphorylation was monitored, and there was no obvious change in comparison with the control group. P-S6K1 increased along with His supplementation in the full concentration range. This was in contrast to the negative effects of His on P-S6K1 in CMECs reported by Prizant and Barash (2008). However, as a whole, P-S6K1 has a dose-dependent effect from His concentration. However, our results support the traditional perfused experiment in which His was positively associated with P-S6K1 in dairy cows (Toerien *et al.*, 2010). The difference between those two studies was potentially due to the different time response of the His addition. In the study of Prizant and Barash (2008), CMECs were incubated in His for 10 min; however, in our study 6 h was used for incubation. We thought after starvation, the CMECs were in a stress period responding to the His addition in a short time, because they needed a period to repair the damages resulting from the devoid of AAs. So the addition of His may

first maintain the cell survival instead of proving the milk synthesis.

4EBP1 is another major downstream target of mTOR (Li *et al.*, 2005). Activated mTOR regulates protein translation by directly inducing P-4EBP1. Our results indicated that the addition of Leu and His to the EBSS increased P-4EBP1 in CMEC-H. Furthermore, only the highest concentrations of Leu inhibited its phosphorylation, which further validated the notion that 4EBP1 was up-regulated by mTOR.

RPS6 is phosphorylated by S6K1. In our study, Leu supplementation slightly improved RPS6 phosphorylation levels between 0.45 and 1.35 mmol/L, while the phosphorylation level was inhibited over 1.35 mmol/L. As a substrate of S6K1, RPS6 was positively correlated with S6K1 phosphorylation between 0.45 and 1.35 mmol/L Leu, which was consistent with the results of Appuhamy *et al.* (2012) that focused on 0.45 mmol/L Leu. On the other hand, His supplementation vastly enhanced the RPS6 phosphorylation level at 0.15 mmol/L, followed by a sharply decreased phosphorylation level between 1.20 and 9.60 mmol/L. Our study revealed that there may have been a negative feedback between RPS6 and S6K1 beyond 1.35 mmol/L Leu or 0.15 mmol/L His supplementation.

The eIF4E is involved in directing ribosomes to the cap structure of mRNAs (Stipanuk, 2007). The mTORC1 phosphorylates 4EBP1 to release eIF4E from the 4EBP1-eIF4E complex, thus promoting eIF4E-dependent translation initiation. The stimulatory effects of His and Leu supplementation on the phosphorylation of eIF4E in CMECs were notably enhanced, which was in accordance with P-4EBP1 changes.

The eEF2 was known to catalyze the mRNA translocation step in the elongation process during protein synthesis (Kaul *et al.*, 2011). Compared with initiation, eEF2 may be an important factor and is likely to be a rate-limiting enzyme for milk protein synthesis under AA-deprived conditions in the mammary gland of cows only in late-lactation because it consumes more energy in elongation (Merrick, 1992; Wilde *et al.*, 1997; Christophersen *et al.*, 2002). Direct spatially-controlled inhibition of eEF2 phosphorylation induces local translational activation (Li *et al.*, 2005; Kaul *et al.*, 2011). Our study demonstrated that both Leu and His ($P < 0.01$)

enhanced eEF2 phosphorylation levels in comparison with a negative control. However, Leu supplementation had enhanced eEF2 phosphorylation by no more than a 20% increase.

A linear regression model was employed to analyze the relationships among EAA, milk protein and mTOR pathway-related proteins. Appuhamy *et al.* (2014) reported that intracellular EAA tended ($P=0.09$) to positively affect mTOR phosphorylation, and the phosphorylation of mTOR ($P<0.01$) was positively associated with fractional rates of mammary protein synthesis (FSR) in bovine mammary tissue slices. Consistently, in the current work, a 1-fold increase in mTOR phosphorylation and G β L expression was associated with a remarkable increase in α s1-casein, α s2-casein, and β -casein expression ($P<0.05$), provision of extracellular Leu and His in CMEC-H. However, Leu supplementation on four casein expressions explained 115%, 22%, 60%, and 71% ($P<0.01$), respectively, of the variability in Raptor phosphorylation. Alternatively, the addition of His to casein protein expression was insignificantly associated with Raptor phosphorylation ($P=0.59, 0.06, 0.31, \text{ and } 0.19$, respectively). Both Leu and His significantly increased mTOR phosphorylation. Leu significantly activated phosphorylation of S6K1, which enhanced four casein expressions in CMEC-H ($P<0.05$). However, when the addition of His was analyzed with the same regression model, the S6K1 phosphorylation had a notable association with α s2-casein and κ -casein ($P<0.05$).

With the addition of Leu or His, α s1-casein was positively related to eEF2 phosphorylation ($P<0.01$) in CMEC-H. In previous experiments, P-eEF2 was associated with P-mTOR, which tended ($P=0.10$) to impose a negative effect on casein synthesis rates, independent of mTOR phosphorylation (Appuhamy *et al.*, 2014). Therefore, the inconsistent results of the effects of Leu and His on eEF2 phosphorylation could be due to different substitutes of the medium. In our study, we did not contain AAs and insulin, while insulin and nonessential amino acids (NEAAs) were present in the medium of the study of Appuhamy *et al.* (2012). It is possible that the potential presence of greater nutrients in substituting in the medium may have partly masked the single AA effects on the cellular status in CMECs. We believe that the interaction of AAs and insulin is more significant than the influence of adding a single AA on mTOR pathways.

5 Conclusions

In summary, the results of this study demonstrated that Leu or His supplementation stimulated α s-casein, β -casein, and κ -casein expression. Casein expression, P-mTOR (Ser²⁴⁸¹), P-Raptor (Ser⁷⁹²), P-eIF4E (Ser²⁰⁹), and P-eEF2 (Thr⁵⁶) increased with Leu concentration in the range of 0.45 to 10.80 mmol/L. Leu supplementation suppressed P-4EBP1 (Thr³⁷) at 10.80 mmol/L, and inhibited P-RPS6 (Ser^{235/236}) over 5.40 mmol/L. Supplementing His from 0.15 to 9.60 mmol/L increased the α s2-casein, β -casein, and κ -casein expression and phosphorylation of S6K1 (Thr³⁸⁹), 4EBP1 (Thr³⁷), eIF4E (Ser²⁰⁹), and eEF2 (Thr⁵⁶) in CMEC-H cells, at 0.15 and 9.60 mmol/L reduced the G β L expression, and over 4.80 mmol/L inhibited RPS6 (Ser^{235/236}). When adding Leu, the α s1-casein was positively correlated with P-mTOR (Ser²⁴⁸¹) ($R^2=0.7820, P<0.01$) and P-S6K1 (Thr³⁸⁹) ($R^2=0.7881, P<0.01$), respectively. With the addition of His, eEF2 was positively correlated with β -casein ($R^2=0.9638, P<0.01$) and κ -casein ($R^2=0.9048, P<0.01$), respectively. Our results provide basic information for further study to clarify the regulation mechanism of Leu and His on casein expression through the mTOR pathway in the dairy cattle mammary gland.

Compliance with ethics guidelines

Hai-na GAO, Han HU, Nan ZHENG, and Jia-qi WANG declare that they have no conflict of interest.

All institutional and national guidelines for the care and use of laboratory animals were followed.

References

- Appuhamy, J.A., Knapp, J.R., Becvar, O., *et al.*, 2011a. Effects of jugular-infused lysine, methionine, and branched-chain amino acids on milk protein synthesis in high-producing dairy cows. *J. Dairy Sci.*, **94**(4):1952-1960. [doi:10.3168/jds.2010-3442]
- Appuhamy, J.A., Bell, A.L., Nayananjalie, W.A., *et al.*, 2011b. Essential amino acids regulate both initiation and elongation of mRNA translation independent of insulin in MAC-T cells and bovine mammary tissue slices. *J. Nutr.*, **141**(6):1209-1215. [doi:10.3945/jn.110.136143]
- Appuhamy, J.A., Knoebel, N.A., Nayananjalie, W.A., *et al.*, 2012. Isoleucine and leucine independently regulate mTOR signaling and protein synthesis in MAC-T cells and bovine mammary tissue slices. *J. Nutr.*, **142**(3):484-491. [doi:10.3945/jn.111.152595]
- Appuhamy, J., Nayananjalie, W.A., England, E.M., *et al.*, 2014. Effects of AMP-activated protein kinase (AMPK)

- signaling and essential amino acids on mammalian target of rapamycin (mTOR) signaling and protein synthesis rates in mammary cells. *J. Dairy Sci.*, **97**(1):419-429. [doi:10.3168/jds.2013-7189]
- Arriola Apelo, S.I., Bell, A.L., Estes, K., et al., 2014a. Effects of reduced dietary protein and supplemental rumen-protected essential amino acids on the nitrogen efficiency of dairy cows. *J. Dairy Sci.*, **97**(9):5688-5699. [doi:10.3168/jds.2013-7833]
- Arriola Apelo, S.I., Knapp, J.R., Hanigan, M.D., 2014b. Invited review: current representation and future trends of predicting amino acid utilization in the lactating dairy cow. *J. Dairy Sci.*, **97**(7):4000-4017. [doi:10.3168/jds.2013-7392]
- Arriola Apelo, S.I., Singer, L.M., Lin, X.Y., et al., 2014c. Isoleucine, leucine, methionine, and threonine effects on mammalian target of rapamycin signaling in mammary tissue. *J. Dairy Sci.* **97**(2):1047-1056. [doi:10.3168/jds.2013-7348]
- Bequette, B.J., Hanigan, M.D., Calder, A.G., et al., 2000. Amino acid exchange by the mammary gland of lactating goats when histidine limits milk production. *J. Dairy Sci.*, **83**(4):765-775. [doi:10.3168/jds.S0022-0302(00)74939-3]
- Beugnet, A., Tee, A.R., Taylor, P.M., et al., 2003. Regulation of targets of mTOR (mammalian target of rapamycin) signalling by intracellular amino acid availability. *Biochem. J.*, **372**(Pt 2):555-566. [doi:10.1042/BJ20021266]
- Bionaz, M., Hurley, W., Loor, J., 2012. Milk protein synthesis in the lactating mammary gland: insights from transcriptomics analyses. In: Hurley, W. (Ed.), *Milk Protein*. InTech, Chapter 11, p.285-324. [doi:10.5772/46054]
- Burgos, S.A., Cant, J.P., 2010. IGF-1 stimulates protein synthesis by enhanced signaling through mTORC1 in bovine mammary epithelial cells. *Domest. Anim. Endocrinol.*, **38**(4):211-221. [doi:10.1016/j.domaniend.2009.10.005]
- Cheng, S.W., Fryer, L.G., Carling, D., 2004. Thr²⁴⁴⁶ is a novel mammalian target of rapamycin (mTOR) phosphorylation site regulated by nutrient status. *J. Biol. Chem.*, **279**(16):15719-15722. [doi:10.1074/jbc.C300534200]
- Christophersen, C.T., Karlsen, J., Nielsen, M.O., et al., 2002. Eukaryotic elongation factor-2 (eEF-2) activity in bovine mammary tissue. *J. Dairy Res.*, **69**(2):205-212. [doi:10.1017/S0022029902005526]
- Durán, R.V., Hall, M.N., 2012. Leucyl-tRNA synthetase: double duty in amino acid sensing. *Cell Res.*, **22**(8):1207-1209. [doi:10.1038/cr.2012.68]
- Gao, H.N., Hu, H., Wang, J.Q., et al., 2015. Effects of leucine and histidine on milk protein synthesis via mammalian target of rapamycin signaling pathway in the bovine mammary epithelial cells. *Chin. J. Anim. Nutr.*, **27**(4):1124-1134 (in Chinese). [doi:10.3969/j.issn.1006-267x.2015.04.016]
- Gerasimovskaya, E.V., Tucker, D.A., Stenmark, K.R., 2005. Activation of phosphatidylinositol 3-kinase, Akt, and mammalian target of rapamycin is necessary for hypoxia-induced pulmonary artery adventitial fibroblast proliferation. *J. Appl. Physiol.*, **98**(2):722-731. [doi:10.1152/jappphysiol.00715.2004]
- Gingras, A.C., Raught, B., Sonenberg, N., 1999. eIF4 initiation factors: effectors of mRNA recruitment to ribosomes and regulators of translation. *Annu. Rev. Biochem.*, **68**(1):913-963. [doi:10.1146/annurev.biochem.68.1.913]
- Hanigan, M.D., Crompton, L.A., Metcalf, J.A., et al., 2001. Modelling mammary metabolism in the dairy cow to predict milk constituent yield, with emphasis on amino acid metabolism and milk protein production: model evaluation. *J. Theor. Biol.*, **213**(2):223-239. [doi:10.1006/jtbi.2001.2417]
- Hara, K., Maruki, Y., Long, X., et al., 2002. Raptor, a binding partner of target of rapamycin (TOR), mediates TOR action. *Cell*, **110**(2):177-189. [doi:10.1016/S0092-8674(02)00833-4]
- Harris, T.E., Lawrence, J.C.Jr., 2003. TOR signaling. *Sci. STKE*, **212**:re15. [doi:10.1126/stke.2122003re15]
- Hristov, A.N., Price, W.J., Shafii, B., 2004. A meta-analysis examining the relationship among dietary factors, dry matter intake, and milk and milk protein yield in dairy cows. *J. Dairy Sci.*, **87**(7):2184-2196. [doi:10.3168/jds.S0022-0302(04)70039-9]
- Hu, H., Zheng, N., Dai, W.T., 2014. Immortalization of a primary bovine mammary epithelial cell line by the SV40 large T-antigen gene. The American Dairy Science Association. Kansas City, MO. [doi:10.3168/jds.2014-8051]
- Kalscheur, K.F., Baldwin VI, R.L., Glenn, B.P., et al., 2006. Milk production of dairy cows fed differing concentrations of rumen-degraded protein. *J. Dairy Sci.*, **89**(1):249-259. [doi:10.3168/jds.S0022-0302(06)72089-6]
- Kaul, G., Pattan, G., Rafeequi, T., 2011. Eukaryotic elongation factor-2 (eEF2): its regulation and peptide chain elongation. *Cell Biochem. Funct.*, **29**(3):227-234. [doi:10.1002/cbf.1740]
- Kim, C.H., Choung, J.J., Chamberlain, D.G., 2001. Estimates of the efficiency of transfer of L-histidine from blood to milk when it is the first-limiting amino acid for secretion of milk protein in the dairy cow. *J. Sci. Food Agric.*, **81**(12):1150-1155. [doi:10.1002/jsfa.923]
- Kim, D.H., Sarbassov, D.D., Ali, S.M., 2002. mTOR interacts with raptor to form a nutrient-sensitive complex that signals to the cell growth machinery. *Cell*, **110**(2):163-175. [doi:10.1016/S0092-8674(02)00808-5]
- Kim, D.H., Sarbassov, D.D., Ali, S.M., et al., 2003. GβL, a positive regulator of the rapamycin-sensitive pathway required for the nutrient-sensitive interaction between raptor and mTOR. *Mol. Cell*, **11**(4):895-904. [doi:10.1016/S1097-2765(03)00114-X]
- Kim, E., 2009. Mechanisms of amino acid sensing in mTOR signaling pathway. *Nutr. Res. Pract.*, **3**(1):64-71. [doi:10.4162/nrp.2009.3.1.64]
- Kim, S.G., Buel, G.R., Blenis, J., 2013. Nutrient regulation of the mTOR complex 1 signaling pathway. *Mol. Cells*, **35**(6):463-473. [doi:10.1007/s10059-013-0138-2]
- Kimball, S.R., 2002. Regulation of global and specific mRNA

- translation by amino acids. *J. Nutr.*, **132**(5):883-886.
- Korhonen, M., Vanhatalo, A., Huhtanen, P., 2002. Evaluation of isoleucine, leucine, and valine as a second-limiting amino acid for milk production in dairy cows fed grass silage diet. *J. Dairy Sci.*, **85**(6):1533-1545 [doi:10.3168/jds.S0022-0302(02)74223-9]
- Laplante, M., Sabatini, D.M., 2012. mTOR signaling in growth control and disease. *Cell*, **149**(2):274-293. [doi:10.1016/j.cell.2012.03.017]
- Li, X., Alafuzoff, I., Soininen, H., et al., 2005. Levels of mTOR and its downstream targets 4E-BP1, eEF2, and eEF2 kinase in relationships with tau in Alzheimer's disease brain. *FEBS J.*, **272**(16):4211-4220. [doi:10.1111/j.1742-4658.2005.04833.x]
- Merrick, W.C., 1992. Mechanism and regulation of eukaryotic protein synthesis. *Microbiol. Rev.*, **56**(2):291-315.
- Moshel, Y., Rhoads, R.E., Barash, I., 2006. Role of amino acids in translational mechanisms governing milk protein synthesis in murine and ruminant mammary epithelial cells. *J. Cell. Biochem.*, **98**(3):685-700. [doi:10.1002/jcb.20825]
- Peterson, R.T., Beal, P.A., Comb, M.J., et al., 2000. FKBP12-rapamycin-associated protein (FRAP) autophosphorylates at serine 2481 under translationally repressive conditions. *J. Biol. Chem.*, **275**(10):7416-7423. [doi:10.1074/jbc.275.10.7416]
- Prizant, R.L., Barash, I., 2008. Negative effects of the amino acids Lys, His, and Thr on S6K1 phosphorylation in mammary epithelial cells. *J. Cell. Biochem.*, **105**(4):1038-1047. [doi:10.1002/jcb.21904]
- Proud, C.G., 2007. Amino acids and mTOR signalling in anabolic function. *Biochem. Soc. Trans.*, **35**(5):1187-1190. [doi:10.1042/BST0351187]
- Stipanuk, M.H., 2007. Leucine and protein synthesis: mTOR and beyond. *Nutr. Rev.*, **65**(3):122-129. [doi:10.1111/j.1753-4887.2007.tb00289.x]
- Toerien, C.A., Trout, D.R., Cant, J.P., 2010. Nutritional stimulation of milk protein yield of cows is associated with changes in phosphorylation of mammary eukaryotic initiation factor 2 and ribosomal S6 kinase 1. *J. Nutr.*, **140**(2):285-292. [doi:10.3945/jn.109.114033]
- Wang, C., Liu, H.Y., Wang, Y.M., et al., 2010. Effects of dietary supplementation of methionine and lysine on milk production and nitrogen utilization in dairy cows. *J. Dairy Sci.*, **93**(8):3661-3670. [doi:10.3168/jds.2009-2750]
- Wang, X., Proud, C.G., 2006. The mTOR pathway in the control of protein synthesis. *Physiology (Bethesda)*, **21**:362-369.
- Wilde, C.J., Quarrie, L.H., Tonner, E., et al., 1997. Mammary apoptosis. *Livest. Prod. Sci.*, **50**(1-2):29-37. [doi:10.1016/S0301-6226(97)00070-5]
- Yang, Q., Inoki, K., Kim, E., et al., 2006. TSC1/TSC2 and Rheb have different effects on TORC1 and TORC2 activity. *PNAS*, **103**(18):6811-6816. [doi:10.1073/pnas.0602282103]

中文概要

题目: 亮氨酸和组氨酸通过mTOR信号通路调控奶牛乳腺上皮细胞中酪蛋白的合成

目的: 以体外培养的永生化奶牛乳腺上皮细胞(CMEC-H)为模型,利用蛋白免疫印迹法(Western blotting)检测酪蛋白和哺乳动物雷帕霉素靶蛋白(mTOR)信号路径元件的蛋白磷酸化表达。进一步探索亮氨酸和组氨酸通过mTOR信号途径调控酪蛋白合成机制的重要意义。

创新点: 首次在CMEC-H模型中研究不同浓度的亮氨酸和组氨酸通过mTOR复合物1(mTORC1)中mTOR(Ser²⁴⁸¹)、mTOR调控蛋白(Raptor, Ser⁷⁹²)和G蛋白β亚基样蛋白(GβL)对酪蛋白表达调控的影响。研究证明mTOR(Ser²⁴⁸¹)并不是在乳腺上皮细胞中激活mTOR信号通路的唯一磷酸化位点, mTOR(Ser²⁴⁸¹)同样可作为激活mTOR信号通路的生物标记。

方法: 以厄尔平衡溶液代替培养基,设为阴性对照,分别添加不同浓度的亮氨酸或组氨酸,利用Western blotting检测酪蛋白和mTOR信号路径元件的蛋白表达。

结论: 与阴性对照组相比,当在CMEC-H细胞中添加0.45~10.80 mmol/L亮氨酸6 h时,4种酪蛋白的表达和mTOR(Ser²⁴⁸¹)、Raptor(Ser⁷⁹²)、真核翻译起始因子4E(eIF4E, Ser²⁰⁹)和真核细胞翻译延伸因子(eEF2, Thr⁵⁶)的磷酸化表达均显著上调($P<0.01$)。而当亮氨酸浓度在10.80 mmol/L时,真核翻译起始因子4E结合蛋白1(4EBP1, Thr³⁷)的磷酸化表达被抑制;在5.40~10.80 mmol/L时,核糖体蛋白S6(RPS6, Ser^{235/236})的磷酸化被抑制。当添加0.15~9.60 mmol/L组氨酸6 h时,α₂-酪蛋白、β-酪蛋白、κ-酪蛋白的表达和mTOR(Ser²⁴⁸¹)、Raptor(Ser⁷⁹²)、核糖体S6蛋白激酶(S6K1, Thr³⁸⁹)、4EBP1(Thr³⁷)、eIF4E(Ser²⁰⁹)和eEF2(Thr⁵⁶)的磷酸化表达均显著上调($P<0.01$)。而当组氨酸浓度在9.60 mmol/L时,α₁-酪蛋白的表达量降低;在0.15和9.60 mmol/L时,GβL被抑制;在4.80~9.60 mmol/L时,RPS6磷酸化被抑制。线性回归模型显示,当添加亮氨酸时,α₁-酪蛋白的表达与mTOR、S6K1和eEF2的磷酸化表达成显著的正相关($P<0.01$;表4);当添加组氨酸,β-酪蛋白和κ-酪蛋白均与eEF2磷酸化表达成显著的正相关($P<0.01$;表5)。综上所述,在乳腺上皮细胞中,亮氨酸和组氨酸能通过mTOR信号通路促进酪蛋白基因的表达。

关键词: 奶牛乳腺上皮细胞;亮氨酸;组氨酸;蛋白免疫印迹; mTOR; 酪蛋白