Research Article
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Neuroprotection of Engineered *Clostridium butyricum*-pMTL007-GLP-1 in A53T α-Syn Mouse Model via PI3K/AKT/GSK-3β

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Abstract: Background: Parkinson's disease (PD) is a prevalent neurodegenerative disorder with limited therapeutic options and no cure, underscoring the urgent need for novel treatment strategies. Our previous work demonstrated that an engineered strain of *Clostridium butyricum*-pMTL007-GLP-1 (*C. butyricum*-GLP-1) alleviated PD symptoms by enhancing mitophagy, though the exact molecular mechanisms remained incompletely understood. Methods: In this study, we further investigated the neuroprotective effects and underlying mechanisms of this engineered strain using an A53T α-synuclein transgenic mouse model of PD. Specifically, we evaluated its impact on motor function, gut α-synuclein expression, intestinal barrier function, gut microbial composition, and neuropathological changes, with a focus on the PI3K/AKT/GSK-3β signaling pathway. Results: Our findings revealed that *C. butyricum*-GLP-1 ameliorated motor deficits in PD mice by reducing intestinal α-synuclein accumulation, restoring gut barrier function, and modulating microbial diversity—notably increasing the relative abundance of of *Prevotella* at the genus level. Furthermore, the engineered strain attenuated neuropathological alterations by decreasing phosphorylated α-synuclein (p-α-syn) in the substantia nigra while upregulating tyrosine hydroxylase (TH), dopamine-transporter (DAT), and glucagon-like peptide-1-receptor (GLP-1R) expression. These neuroprotective effects were associated with suppressed proinflammatory responses and enhanced anti-inflammatory and anti-apoptotic signaling, likely mediated through PI3K/AKT/GSK-3β pathway activation. Conclusions: *C. butyricum*-pMTL007-GLP-1 exerts significant neuroprotective effects in PD mice by reshaping gut microbiota composition and activating the PI3K/AKT/GSK-3β pathway. These findings provide further theoretical support for the potential application of probiotic-based therapies in Parkinson's disease treatment.

Key words: Parkinson's disease; *Clostridium butyricum*-pMTL007-GLP-1; Glucagon-like peptide-1 (GLP-1); Gut microbiota; PI3K/AKT/GSK-3β pathway

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1 Introduction

Parkinson's disease (PD) is a progressive neurodegenerative disorder characterized by four cardinal motor symptoms: resting tremor, bradykinesia, muscle rigidity, and postural instability. The disease's neuropathological hallmarks include the accumulation of misfolded α-synuclein (α-syn) aggregates, progressive degeneration of dopaminergic neurons in the substantia nigra pars compacta, and the formation of intraneuronal Lewy bodies (Tripodi et al., 2024; Mulvaney et al., 2020). Epidemiologically, PD affects approximately 1% of the global population over 60 years of age, with prevalence increasing dramatically to 3-4% among octogenarians. This represents a growing public health challenge with substantial socioeconomic implications (Tarsy, 2012). Despite decades of research, PD remains a complex multifactorial disorder whose exact etiopathogenesis continues to elude complete understanding (Lv et al., 2020). Current gold-standard pharmacotherapies, principally Levodopa, dopamine-receptor agonists, and monoamine-oxidase-B inhibitors, offer primarily symptomatic relief by targeting dopaminergic pathways but fail to modify the underlying disease progression (Chang et al., 2022). This critical therapeutic gap underscores the pressing need to develop novel treatment strategies that can both alleviate symptoms and fundamentally alter the neurodegenerative course of PD, thereby improving long-term patient outcomes and quality of life.

Current evidence highlights the critical role of the PI3K/AKT signaling pathway in PD, where it regulates cell survival and modulates inflammatory responses (Malagelada et al., 2009; Yang et al., 2014). Downstream of this pathway, glycogen synthase kinase-3β (GSK-3β) drives α-synuclein aggregation and neuroinflammation. Its activity is regulated through inhibitory phosphorylation at Ser9 (Golpich et al., 2015; Kozikowski et al., 2006), with accumulating studies implicating GSK-3β as a key contributor to PD pathogenesis (Lei et al., 2011). Pharmacological interventions targeting GSK-3β have demonstrated therapeutic potential. For instance, chlorogenic acid reduces rotenone-induced phosphorylated α-synuclein levels by inhibiting GSK-3β activity. Similarly, PNU-120596 attenuates neuroinflammation in murine models by suppressing the JAK2/NF-κB/GSK3β axis (Gowayed et al., 2022; Sharma et al., 2022). These findings suggest that modulating GSK-3β to enhance PI3K/AKT signaling represents a promising strategy for PD treatment.

Glucagon-like peptide-1 (GLP-1), a gut-derived hormone secreted by intestinal L cells, and its receptor agonists (such as exendin-4) have demonstrated well-documented neuroprotective effects in multiple rodent models of PD (Holst, 2007; Hölscher, 2018; Reich and Hölscher, 2022). Mechanistically, GLP-1 receptor (GLP-1R) activation initiates PI3K/AKT signaling through its canonical G-Protein-Coupled Receptor (GPCR) pathway involving sequential β-arrestin recruitment, adenylate cyclase stimulation, and PKA-dependent PI3K activation. This signaling cascade's critical role in GSK-3β modulation has been consistently demonstrated across multiple disease models (AD, PD, and trophoblasts), with PI3K inhibitors reliably abolishing these effects (Yang et al., 2016; Meng et al., 2016; Wang et al., 2017; Wu et al., 2018). However, the therapeutic application of GLP-1 is severely constrained by rapid enzymatic degradation once in the bloodstream, resulting in an extremely short half-life of merely 1-2 minutes (Lorenz et al., 2013). While synthetic GLP-1 analogs have been developed to circumvent this limitation, their clinical utility remains hampered by the necessity for prolonged subcutaneous administration and high treatment costs, which collectively diminish patient compliance and impose significant socioeconomic burdens. Consequently, developing strategies to improve the pharmacokinetic and pharmacoeconomic profile of GLP-1-based therapies represents a critical unmet need in PD treatment.

Emerging evidence underscores the potential of microbial interventions in promoting healthy aging, with particular emphasis on the gut microbiota's pivotal role in PD pathogenesis (Xu et al., 2024; Wang et al., 2021; Chidambaram et al., 2021). Preclinical investigations and clinical observations have consistently shown that probiotic administration can effectively modulate the gut microbiota, strengthen intestinal barrier integrity, and attenuate neuroinflammatory responses, consequently mitigating PD-related symptoms (Snigdha et al., 2022). A notable example is *Clostridium butyricum* (*C. butyricum*), which has been demonstrated to improve motor dysfunction in PD murine models through the gut microbiota-GLP-1 signaling axis (Sun et al., 2021). Nevertheless, large-scale, well-controlled clinical trials remain imperative to conclusively validate the

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therapeutic efficacy of probiotics and elucidate their precise mechanisms of action in PD management.

In our previous studies, we successfully developed two genetically engineered strains — *Lactococcus lactis* MG1363-pMG36e-GLP-1 and *Escherichia coli* (*E. coli*) Nissle 1917-GLP-1 — both of which demonstrated significant neuroprotective effects in murine models of PD (Fang et al., 2020; Wu et al., 2022). *Clostridium butyricum*, a Generally Recognized As Safe (GRAS)-certified strain, outperforms *E. coli* (virulence risks) and *Lactobacillus* (low transformation efficiency, 10^4-10^5 CFU/μg DNA) with its gastric acid resistance (>90% survival), superior intestinal colonization (10^8 CFU/g), and high transformation efficiency (10^7 CFU/μg DNA). Its therapeutic potential in PD further supports its use as an ideal engineered probiotic vector (Stoeva et al., 2021; Zhang et al., 2024; Sun et al., 2021). Building upon these findings, we subsequently engineered the C. butyricum-pMTL007-GLP-1 strain. This novel construct combines the neuroprotective properties of GLP-1 with the therapeutic potential of *C. butyricum*, and has shown promising neuroprotective outcomes in MPTP-induced PD mouse models (Wang et al., 2023). While these results are encouraging, the exact molecular mechanisms mediating these neuroprotective effects remain to be fully elucidated. In the current study, we employed a well-characterized A53T-SNCA transgenic PD mouse model to systematically evaluate the therapeutic potential of *C. butyricum*-pMTL007-GLP-1. Our comprehensive approach combines behavioral assessments, molecular analyses, and gut microbiota profiling to uncover the underlying mechanisms of action. We anticipate that this study will provide valuable mechanistic insights and robust experimental evidence to advance PD therapeutics, potentially contributing to the development of next-generation engineered bacterial medications.

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2 Methods

2.1 Animals and experimental design

Thirty-two 2-month-old male A53T α-synuclein transgenic mice and eight wild-type C57BL/6 mice were purchased from Changzhou Cavens Laboratory Animal Co., LTD (Changzhou, China) and maintained under specific pathogen-free (SPF) conditions with controlled environmental parameters (12:12 light-dark cycle, 50-55% humidity, 22-24℃) and *ad libitum* access to food and water. All experimental procedures were conducted between 9:00 - 12:00 a.m. to minimize circadian variability (see treatment schedule in Fig. 1a). Following a 7-day acclimatization period, we confirmed successful model establishment through standardized behavioral assessments. Wild-type C57BL/6 mice (n=8, Group C) received daily oral gavage of 100 μL gelatin-saline solution (0.1% gelatin in 0.9% saline, w/v) for 30 days, while the transgenic mice were randomly divided into four treatment groups (n=8/group). The AM group (disease model) received daily oral gelatin-saline (100 μL); the AL group (positive control) received daily intraperitoneal injections of 0.4 mg/kg liraglutide (GLP-1 receptor agonist) to circumvent potential oral degradation; the ACB group received daily oral administration of 10^8 CFU/mL *C. butyricum* in 0.01% gelatin-saline (100 μL); and the ACBG group received equivalent dosing of *C. butyricum*-pMTL007-GLP-1 via identical administration protocol. All animal procedures strictly adhered to National Institutes of Health guidelines and were approved by the Animal Experimental Ethical Inspection Committee of Nanchang Royo Biotechnology Co., Ltd., Nanchang, China (Ethical Approval Number: RyE2021070912).

2.2 Behavioral assessment

All mice were habituated to the behavioral chamber environment for 30 minutes 24 hours prior to testing. Behavioral assessments were then conducted following established protocols from our previous study (Wang et al., 2023). These included three standardized tests: (1) The pole test involved a cotton-wrapped vertical pole (50 cm long, 1 cm in diameter), with descent time (from top to base) recorded as the primary outcome measure; (2) The open-field test took place in a 40×40 cm arena where total distance traveled and central zone exploration (20×20 cm center area) were automatically tracked over a 10-minute session using video analysis software (ANY-maze V6.3, Stoelting Co., USA), and the arena was wiped with 70% (v/v) ethanol between trials; (3) The hanging wire test evaluated neuromuscular coordination with mice positioned on a suspended wire (50 cm long, 2 mm in diameter) and scored limb engagement (0-4 points based on number of paws maintaining grip, with

immediate 0 score upon falling). All tests were performed in a dedicated behavioral suite with 15-minute inter-test intervals to minimize stress interference, and final scores represented the mean of three consecutive trials per animal conducted on separate days to ensure reliability.

2.3 Sample collection

Following behavioral assessment, mice were humanely euthanized via isoflurane overdose. Blood samples were collected via cardiac puncture, allowed to coagulate at 37°C for 2 hours, then centrifuged at 3000 g for 15 min at 4°C to obtain serum, which was aliquoted and stored at -80°C until analysis. Fresh fecal pellets were immediately flash-frozen in liquid nitrogen and maintained at -80°C for subsequent 16S rRNA sequencing and microbiota profiling. Brain tissues (including substantia nigra and striatum) and intestinal segments (duodenum, jejunum, and colon) were either snap-frozen in liquid nitrogen for molecular analyses or immersion-fixed in 4% paraformaldehyde (PFA) in 0.1 mol/L phosphate buffer (pH 7.4) for 24 h at 4°C prior to paraffin embedding and histological processing.

2.4 Gut microbiota analysis

Gut microbiota composition was analyzed using 16S rRNA high-throughput sequencing following established protocols (Chen et al., 2018). Genomic DNA was extracted from the mouse fecal samples and subjected to PCR amplification targeting the V3-V4 hypervariable regions of the 16S rRNA gene using universal primers 338F (5′-ACTCCTACGGGAGGCAGCAG-3′) and 806R (5′-GGACTACHVGGGTWTCTAAT-3′; where H = A/T/C, V = G/A/C, and W = A/T) (Personalbio, Shanghai, China). We sequenced the resulting amplicons on an Illumina platform to generate raw sequencing data, which were subsequently processed through the QIIME2 pipeline (version 2019.4). Sequence analysis was performed using the UPARSE software package, and sequences sharing ≥97% similarity were clustered into operational taxonomic units (OTUs). Following OTU clustering, read counts were normalized by rarefaction to the minimum sequencing depth across all samples to ensure comparability. Alpha diversity metrics were then calculated to assess microbial community complexity. We taxonomically classified representative sequences from each OTU, based on alignment with the Ribosomal Database Project (RDP) reference database. The resulting species-abundance matrix was used for downstream analyses. All sequencing data were deposited in the NCBI Sequence Read Archive under accession number PRJNA1091924.

2.5 Immunofluorescence (IF) and Immunohistochemistry (IHC)

Brain and colon tissues were embedded in paraffin and sectioned into 5-μm-thick slices. Following deparaffinization and rehydration, endogenous peroxidase activity was blocked by incubating the sections with 3% (v/v) H_2O_2, followed by blocking with 5% (v/v) goat serum at room temperature. The sections were then incubated overnight at 4°C with primary antibodies (detailed in Supplementary Table S1). After washing, bound primary antibodies were detected using appropriate secondary antibodies. Immunoreactivity was visualized under a light microscope (Higashi et al., 2007).

2.6 Western blot analysis

Brain and colon tissues were homogenized in radioimmunoprecipitation assay (RIPA) lysis buffer (Solarbio Life Science, China, Cat# R0010) containing protease and phosphatase inhibitors to preserve protein integrity during extraction. Following homogenization, lysates were centrifuged at 12,000g for 10 min at 4°C, and the resulting supernatants were collected for protein quantification using a bicinchoninic acid (BCA) assay kit (Thermo Fisher Scientific, Cat# A53226). Equal amounts of protein were resolved by 10% — 12% (0.1 — 0.12 g/ml) SDS-PAGE and electrophoretically transferred onto polyvinylidene fluoride (PVDF) membranes. After blocking with 5% (v/v) non-fat milk in Tris-buffered saline containing 0.1% (v/v) Tween-20 (TBST) for 1 hour at room temperature, membranes were probed overnight at 4 °C with primary antibodies (see Supplementary Table S1 for details). Following three washes with TBST, membranes were incubated with horseradish peroxidase (HRP)-conjugated secondary antibodies diluted in 1% (v/v) milk-TBST for 1 h at room temperature. Protein-antibody complexes were visualized using an enhanced chemiluminescence (ECL) detection system (Thermo Fisher Scientific, USA), and band intensities were quantified by densitometric analysis using ImageJ software (NIH, USA) (Zeng et al., 2017).

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2.7 Real-Time PCR

Total RNA was isolated from brain tissue using TRIzol reagent (Invitrogen, USA) and reverse-transcribed into complementary DNA (cDNA) using a PrimeScript RT reagent kit (Takara, Japan, Cat# 639506). Quantitative real-time PCR (qRT-PCR) was performed on an ABI 7900HT Fast Real-Time PCR System (Applied Biosystems, USA) using SYBR Green chemistry. Each 20 μL reaction mixture contained: 10 μL of 2×SYBR Premix EX Taq II (Takara, Japan, Cat# RR420A), 0.4 μL of ROX Reference Dye II (50×) (Takara, Japan, Cat# RR420A), 1.0 μL of cDNA template, 0.8 μL each of forward and reverse primers (final concentration 0.4 μM), and 7 μL of nuclease-free water. The thermal cycling protocol consisted of an initial denaturation at 95 °C for 10 min, followed by 40 cycles of 95°C for 30 s, 60°C for 30 s, and 72°C for 30 s. Gene-expression levels of target genes (IL-1β, IL-6, TNF-α, and GLP-1R) were normalized to the endogenous control GAPDH and calculated using the comparative threshold cycle ($2^{-\Delta\Delta Ct}$) method. All primer sequences are provided in Supplementary Table S2.

2.8 Measurement of DA, IL-1β, IL6, TNF-α, and GLP-1 in serum

Dopamine (DA) levels in the substantia nigra (SN) and serum concentrations of IL-1β, IL6, TNF-α, and GLP-1 were quantified using commercial assay kits (Nanjing Jiancheng Bioengineering Institute, Nanjing, China, Cat. No. H170-1-2; 4A BIOTECH, Cat. No. CME0015, CME0006, CME0004; and IBL, Tokyo, Japan, Cat. No. 27788) following the manufacturer's protocols. During sample collection, we did not use peptidase inhibitors (including DPP-4 inhibitors), as our experimental design accounted for potential GLP-1 degradation by relying on sustained bacterial GLP-1 production to maintain stable levels.

2.9 Statistical analysis

Statistical analysis was performed using GraphPad Prism 7.0 (GraphPad Software, San Diego, CA, USA). Data were analyzed by one-way analysis of variance (ANOVA) with Tukey's post hoc test for multiple comparisons. Results are presented as mean ± standard deviation (SD), and a p-value < 0.05 was considered statistically significant.

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3 Results

3.1 *C. butyricum*-pMTL007-GLP-1 improved motor impairments in PD mice

To assess the therapeutic effects of *C. butyricum*-pMTL007-GLP-1 in PD mice, we evaluated locomotor function through a series of behavioral tests (Fig. 1a). In the open-field test, PD mice showed characteristic deficits including: reduced center exploration time (anxious behavior), decreased total distance traveled (AM: 1532 cm *vs.* C: 3728 cm, $p<0.01$), and fewer center entries (AM: 13.38 *vs.* C: 30, $p<0.01$) — all indicative of bradykinesia and impaired exploratory motivation. These impairments were substantially ameliorated by *C. butyricum*-pMTL007-GLP-1 treatment (ACBG), showing comparable efficacy to liraglutide (Fig. 1b-1d). Motor-coordination assessments demonstrated parallel improvements, with ACBG normalizing both the prolonged pole-test descent time (ACBG: 9.875 s *vs.* AM: 15.79 s, $p<0.01$) and shortened hanging-test latency (ACBG: 107.7 s *vs.* AM: 50.33 s, $p<0.01$). Notably, ACBG outperformed the parental strain (ACB) in both tests (pole: 9.875 s *vs.* 12.92 s; hanging: 107.7 s *vs.* 78.1 s, $p<0.05$) while matching liraglutide's efficacy (Figure 1e and 1f).

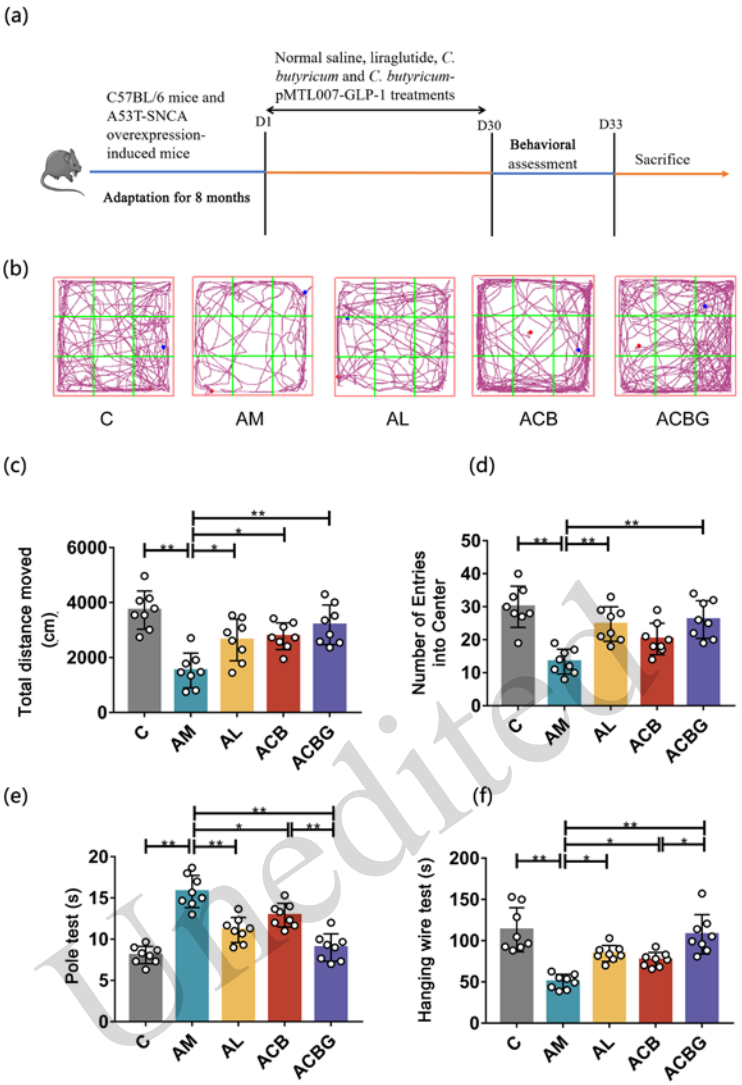

Fig. 1 *C. butyricum*-pMTL007-GLP-1 alleviated motor abnormalities in PD mice. (a) Schematic diagram of the experimental design. (b) Movement trajectories of the five groups of mice in the open-field test. The blue dot indicates the starting point, and the red dot indicates the endpoint. (c) Cumulative distance traveled in the open-field test. (d) Number of entries into the center in the open-field test. (e) Descent time in the pole test. (f) Drop latency in the hanging wire test. C: Normal mice (n = 8); AM: PD mice (n = 8); AL: Liraglutide-treated PD mice (n = 8); ACB: *C. butyricum*-treated PD mice (n = 8); ACBG: *C. butyricum*-pMTL007-GLP-1-treated PD mice (n = 8). Data were analyzed by one-way analysis of variance (ANOVA) with Tukey's post hoc test for multiple comparisons. Data are presented as mean ± standard deviation (SD). *$p < 0.05$, **$p < 0.01$.

3.2 *C. butyricum*-pMTL007-GLP-1 restored disrupted gut microbiota in PD mice

To investigate the effects of *C. butyricum*-pMTL007-GLP-1 on the gut microbiota composition in PD mice, we performed 16S rRNA high-throughput sequencing. Our findings demonstrated that α-diversity indices, including Chao1 and observed species richness, were significantly reduced in PD mice compared to controls, but were partially restored following *C. butyricum*-pMTL007-GLP-1 treatment (Fig. 2a and 2b). Principal Coordinate Analysis (PCoA) based on unweighted UniFrac distances revealed distinct clustering patterns in the C and AM groups, with the ACBG group showing an intermediate microbial profile (Fig. 2c). Venn diagram analysis identified 320 OTUs shared across all groups, while the number of unique OTUs was 565 (C group), 339 (AM group), 437 (AL group), 419 (ACB group), and 492 (ACBG group) (Fig. 2d). Notably, the ACBG group exhibited the highest degree of OTU similarity with the control group, suggesting microbiota restoration.

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Taxonomic profiling at the phylum level showed that Bacteroidetes, Firmicutes, Proteobacteria, and Verrucomicrobia constituted the four most abundant phyla across all experimental groups (Fig. 2e). Of particular interest, microbiome analysis revealed PD-associated dysbiosis marked by significant reductions in *Prevotella* (2.5 ± 1% *vs.* control 6.1 ± 2%, $p<0.05$), a mucin-producing genus essential for gut-barrier integrity, and near elimination of the pro-inflammatory taxon *AF12* (0.0 ± 0.1% *vs.* control 1.1 ± 0.4%, $p<0.05$). C. butyricum-pMTL007-GLP-1 treatment not only normalized but surpassed physiological *Prevotella* levels (10 ± 3%, $p < 0.05$) while restoring *AF12* to baseline (1.0 ± 0.2%, $p < 0.05$) (Fig. 2f-2j). These results demonstrate that *C. butyricum*-pMTL007-GLP-1 treatment can restore gut microbial diversity and composition in PD mice, potentially contributing to the observed therapeutic effects on PD symptoms.

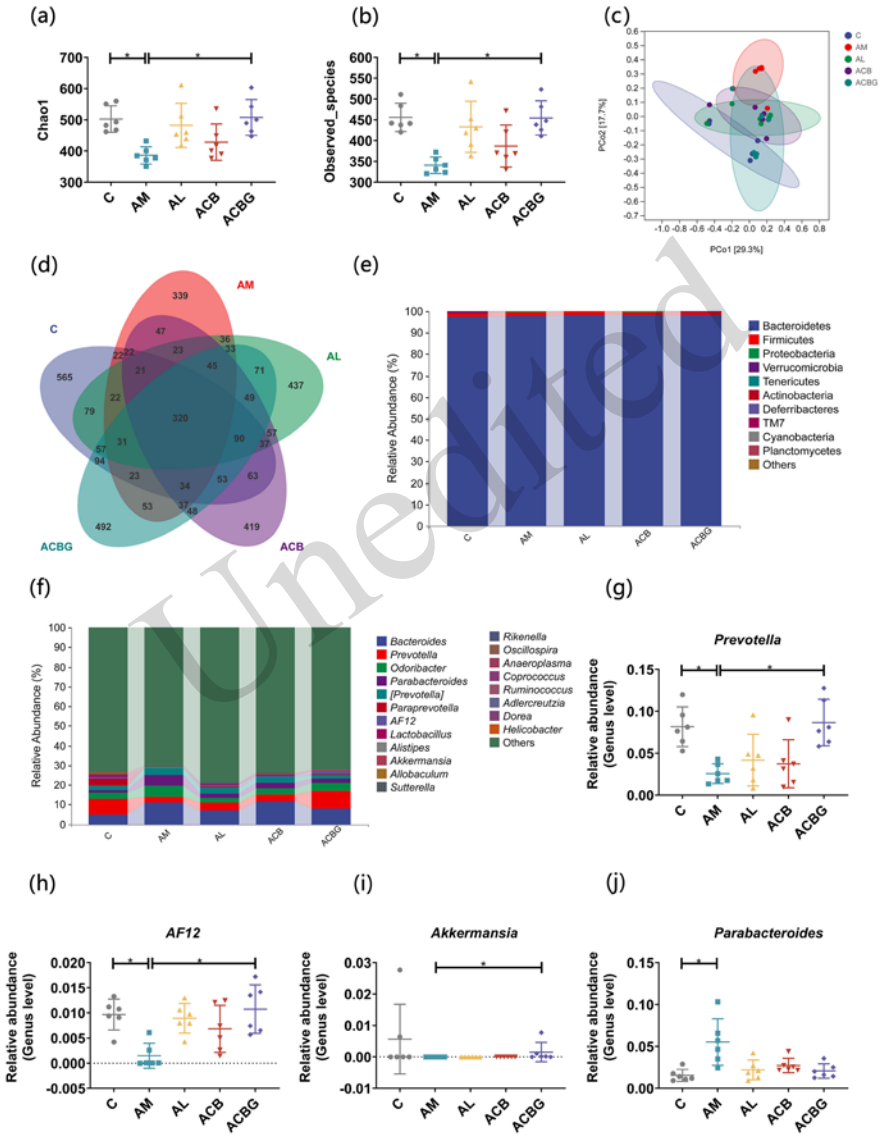

Fig. 2 *C. butyricum*-pMTL007-GLP-1 restored gut microbiota diversity in PD mice. (a) Chao1 index. (b) Observed species index. (c) PCoA plot of β-diversity index. (d) Venn diagram of OTUs. (e) Microbial species composition at the phylum level. (f) Microbial species composition at the genus level. Relative Abundance of *Prevotella* (g), *AF12* (h), *Akkermansia* (i), and *Parabacteroides* (j). C: Normal mice (n = 6); AM: PD mice (n = 6); AL: Liraglutide-treated PD mice (n = 6); ACB: *C. butyricum*-treated PD mice (n = 6); ACBG: *C. butyricum*-pMTL007-GLP-1-treated PD mice (n = 6). Data were analyzed by one-way analysis of variance (ANOVA) with Tukey's post hoc test for multiple comparisons. Data are presented as mean ± standard deviation (SD). *$p < 0.05$, **$p < 0.01$.

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3.3 *C. butyricum*-pMTL007-GLP-1 attenuated intestinal α-syn accumulation and improved intestinal-barrier integrity in PD mice

To assess the effects of *C. butyricum*-pMTL007-GLP-1 on intestinal pathophysiology in PD mice, we first examined GLP-1 secretion using IF. The AM group exhibited significantly fewer GLP-1-positive cells compared to the C group, while *C. butyricum*-pMTL007-GLP-1 treatment substantially restored GLP-1-positive cell populations (Fig. 3a, quantitative analyses in Supplementary Fig. S1). We next evaluated α-synuclein pathology via IHC. *C. butyricum*-pMTL007-GLP-1 treatment significantly reduced both p-α-synuclein and α-synuclein accumulation while increasing p-GSK-3β levels in intestinal tissues (Fig. 3b). To investigate intestinal barrier function, we performed Western blotting analysis of colonic tight-junction proteins. Compared to controls, PD mice showed markedly decreased expression of occludin and zonula occludens-1 (ZO-1), key components of intestinal tight junctions. Importantly, *C. butyricum*-pMTL007-GLP-1treatment significantly restored the expression of these barrier proteins (Fig. 3c-3e). These results demonstrated that *C. butyricum*-pMTL007-GLP-1 exerted dual beneficial effects by reducing pathological α-synuclein accumulation in the gut and enhancing intestinal barrier integrity through GLP-1-mediated mechanisms. These findings suggest that the therapeutic effects of *C. butyricum*-pMTL007-GLP-1 on PD symptoms may be mediated, at least in part, through gut-brain axis modulation.

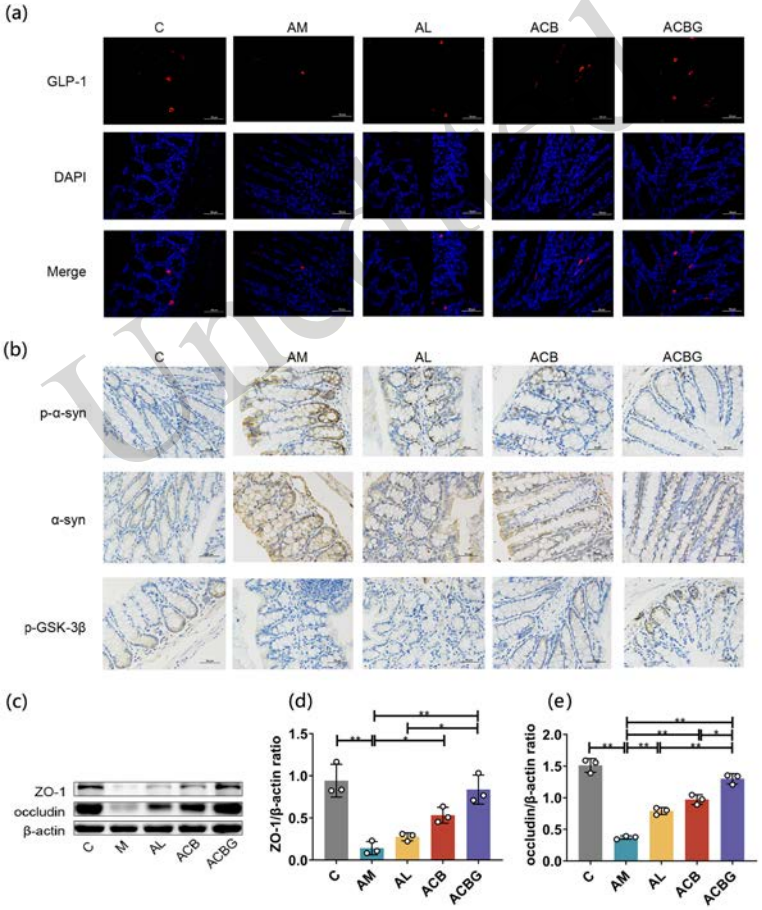

Fig. 3 *C. butyricum*-pMTL007-GLP-1 reduced intestinal α-syn expression and enhanced intestinal-barrier function by promoting GLP-1 secretion in PD mice. (a) Immunofluorescence analysis of GLP-1 in the colon (magnification: ×400, scale bar = 50 μm). (b) Immunohistochemistry analysis of α-synuclein, p-α-synuclein, and p-GSK-3β in the colon (magnification: ×400, scale bar = 50 μm). (c-e) Western blot results showing expression of ZO-1 and occludin (with β-actin as the internal reference) in the colon. C: Normal mice (n = 3); AM: PD mice (n = 3); AL: Liraglutide-treated PD mice (n = 3); ACB: *C. butyricum*-treated PD mice (n = 3); ACBG: *C. butyricum*-pMTL007-GLP-1-treated PD mice (n = 3). Data were analyzed by one-way analysis of variance (ANOVA) with Tukey's post hoc test for multiple comparisons. Data are presented as mean ± standard deviation (SD). *$p < 0.05$, **$p < 0.01$.

3.4 *C. butyricum*-pMTL007-GLP-1 mitigated neuropathological alterations in PD mice

To evaluate dopaminergic neuron integrity, we first measured striatal DA levels by ELISA. PD mice exhibited significantly reduced DA content compared to controls (C *vs.* AM: 530.4 ng/mL *vs.* 405.9 ng/mL, $p < 0.01$). Notably, all treatment groups showed substantial DA restoration (ACB: 479.5 ng/mL; AL: 488.8 ng/mL; ACBG: 520.9 ng/mL; all $p < 0.01$ *vs.* AM), with *C. butyricum* (ACB) and *C. butyricum*-pMTL007-GLP-1 (ACBG) demonstrating efficacy comparable to liraglutide (Fig. 4a). Western blot analysis revealed concomitant changes in key dopaminergic markers. Both dopamine-transporter (DAT) and GLP-1-receptor (GLP-1R) expression were significantly downregulated in PD mice but restored following *C. butyricum*-pMTL007-GLP-1 treatment (Fig. 4b-4d). Immunofluorescence and immunoblotting further demonstrated that tyrosine hydroxylase (TH)-positive neurons, which were heavily depleted in PD mice, were significantly preserved across treatment groups, with maximal protection observed in the ACBG group (Fig. 4e and 4f). α-Synuclein pathology analysis yielded complementary findings. Both p-α-syn and α-syn immunoreactivity were substantially increased in PD mice, but significantly attenuated by *C. butyricum*-pMTL007-GLP-1 treatment (Fig. 4g-4i; quantitative analyses in Supplementary Fig. S1). Western blot quantification confirmed these morphological observations, showing reduced α-synuclein accumulation in treated animals.

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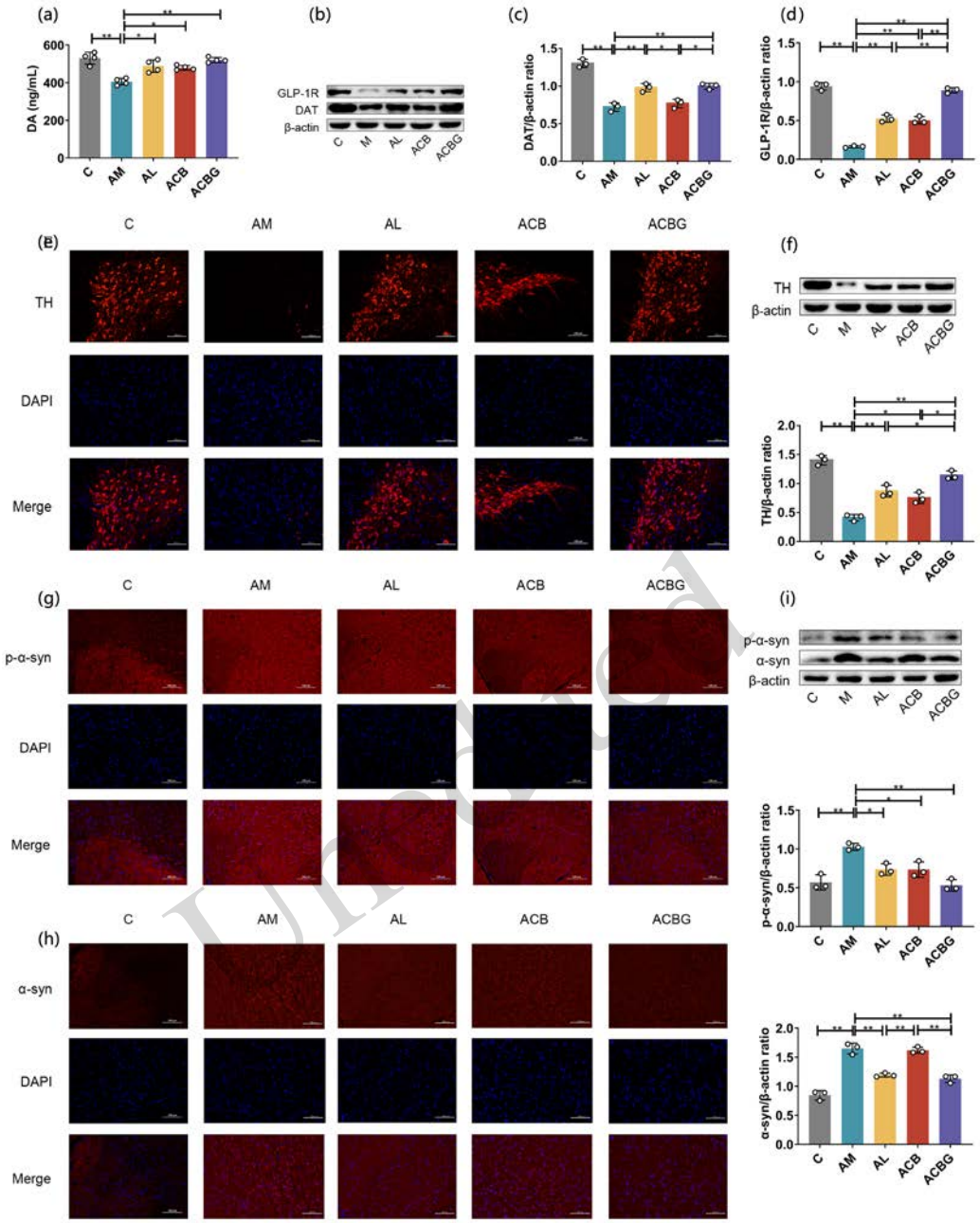

Fig. 4 *C. butyricum*-pMTL007-GLP-1 attenuated neuropathological variations in PD mice. (a) Expression levels of DA in the SN using ELISA analysis (n = 4). (b) Western blot results showing DAT and GLP-1R expression (with β-actin as the internal reference) in the SN. Quantitative analysis of DAT (c) and GLP-1R (d). Immunofluorescence analysis of TH (e), p-α-syn (g), and α-syn (h) (magnification: ×200, scale bar = 100 μm). (f) Western blot analysis showing TH expression. (i) Western blot analysis showing p-α-synuclein and α-synuclein expression. C: Normal mice (n = 3); AM: PD mice (n = 3); AL: Liraglutide-treated PD mice (n = 3); ACB: *C. butyricum*-treated PD mice (n = 3); ACBG: *C. butyricum*-pMTL007-GLP-1-treated PD mice (n = 3). Data were analyzed by one-way analysis of variance (ANOVA) with Tukey's post hoc test for multiple comparisons. Data are presented as mean ± standard deviation (SD). *$p < 0.05$, **$p < 0.01$.

3.5 *C. butyricum*-pMTL007-GLP-1 attenuated neuroinflammation and apoptosis through PI3K/AKT/GSK-3β pathway activation in PD mice

To investigate the anti-neuroinflammatory effects of *C. butyricum*-pMTL007-GLP-1, we performed IF and WB analyses to assess astrocyte initiation (GFAP) and microglia activation (Iba1) in the SN of A53T

α-synuclein transgenic mice (Fig. 5a-5c). Quantitative analyses are presented in Supplementary Fig. S1. Compared to the C group, the AM group displayed significant increases in both GFAP+ astrocytes and Iba1+ microglia in the SN ($p < 0.01$). In particular, *C. butyricum*-pMTL007-GLP-1 treatment more potently attenuated glial activation than either liraglutide or wild-type *C. butyricum* alone, as confirmed by WB quantification of GFAP and Iba1 protein levels. At the molecular level, q-PCR and ELISA analyses demonstrated elevated expression of pro-inflammatory cytokines (IL-1β, IL-6, and TNF-α) in PD mice, which were significantly suppressed by all treatments ($p < 0.05$). The engineered probiotic (ACBG group) exhibited the most robust anti-inflammatory effects (Fig. 5a-5c). Importantly, *C. butyricum*-pMTL007-GLP-1 treatment also restored GLP-1R expression and increased GLP-1 levels (ACBG *vs.* AM: 8.068 pmol/L *vs.* 4.209 pmol/L, $p < 0.01$; Supplementary Fig. S1), suggesting a potential mechanism for its neuroprotective action.

To elucidate the molecular mechanisms underlying the anti-inflammatory effects of C. butyricum-pMTL007-GLP-1, we performed Western blot analysis of the PI3K/AKT/GSK-3β signaling pathway in A53T α-synuclein transgenic mice (Fig. 5d-5h). Compared to the C group, the AM group demonstrated significant reductions in the phosphorylation ratios of PI3K (p-PI3K/PI3K), AKT (p-AKT/AKT), and GSK at $3\beta^{Ser9}$ (p-GSK-$3\beta^{Ser9}$/GSK-3β), along with elevated phosphorylation of p65 (p-p65/p65), indicative of NF-κB pathway activation (all $p < 0.01$ versus C group). Strikingly, *C. butyricum*-pMTL007-GLP-1 treatment not only normalized these aberrant signaling patterns but showed superior efficacy to both wild-type *C. butyricum* and liraglutide treatments. Further investigation of apoptotic regulators revealed that PD mice exhibited prominent dysregulation of apoptosis-related proteins, including increased expression of pro-apoptotic factors (Bax and cleaved-caspase-3) and reduced levels of anti-apoptotic Bcl-2 (Fig. 5i-5l). Treatment with *C. butyricum*-pMTL007-GLP-1 effectively restored the balance of these apoptotic markers, demonstrating comparable therapeutic effects to liraglutide ($p < 0.05$ versus AM group). Collectively, these findings suggest that *C. butyricum*-pMTL007-GLP-1 ameliorated PD pathology in A53T α-synuclein transgenic mice through coordinated activation of the PI3K/AKT/GSK-3β neuroprotective pathway, which concurrently mediated potent anti-inflammatory effects and suppressed apoptotic signaling cascades.

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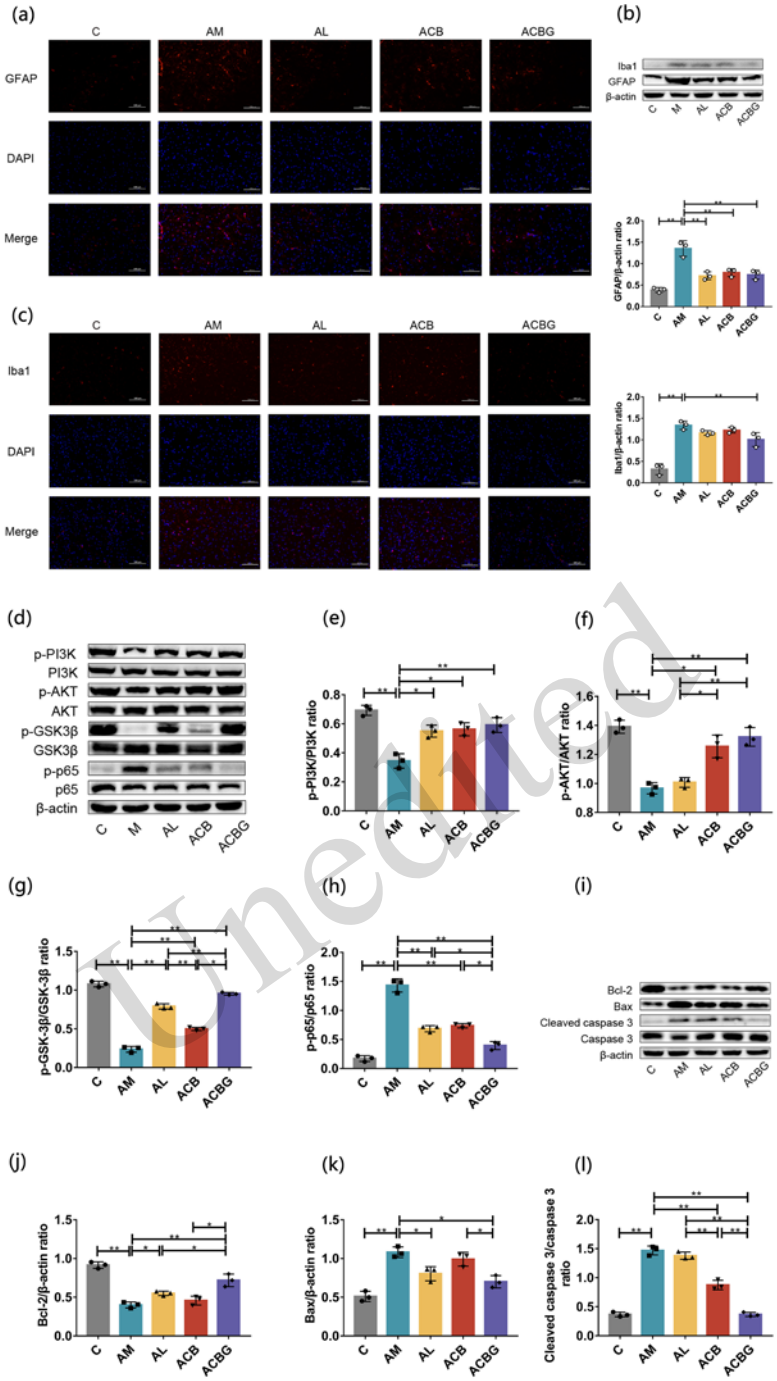

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Fig. 5 *C. butyricum*-pMTL007-GLP-1 activated the PI3K/AKT/GSK-3β pathway to reduce inflammation and apoptosis in PD mice. (a) Immunofluorescence analysis of GFAP (a) and Iba1 (c) in the SN of mice (magnification: ×200, scale bar = 100 μm). (b) Western blot analysis showing GFAP, Iba1, and β-actin expression in the SN. (d) Western blot analysis showing protein expression in the signaling pathway. (e-h) Quantitative analyses of expression levels of p-PI3K/PI3K (e), p-AKT/AKT (f), p-GSK-3β/GSK-3β (g), and p-p65/p65 (h). (i) Western blot analysis showing apoptosis-related protein expression. (j-l) Quantitative analysis of expression levels of Bcl-2 (j), Bax (k), and cleaved-caspase-3/caspase-3 (l). C: Normal mice (n = 3); AM: PD mice (n = 3); AL: Liraglutide-treated PD mice (n = 3); ACB: *C. butyricum*-treated PD mice (n = 3); ACBG: *C. butyricum*-pMTL007-GLP-1-treated PD mice (n = 3). Data were analyzed by one-way analysis of variance (ANOVA) with Tukey's post hoc test for multiple comparisons. Data are presented as mean ± standard deviation (SD). *$p < 0.05$, **$p < 0.01$.

4 Discussion

PD, the second most prevalent neurodegenerative disorder among middle-aged and elderly populations, remains without disease-modifying therapies to effectively halt its progression (Bohnen et al., 2025; Vaughan et al., 2025). Recent advances in microbiome research and genetic engineering technology have enabled the development of novel probiotic-based therapeutic strategies (Zommiti et al., 2020). Our research group has pioneered the development of three genetically engineered probiotic strains for sustained GLP-1 delivery: *L. lactis*-MG1363-pMG36e-GLP-1, *E. coli* Nissle 1917, and *C. butyricum*-pMTL007-GLP-1 (Fang et al., 2020; Wang et al., 2023). While all three strains demonstrated neuroprotective potential in PD models, the first two candidates (*L. lactis* and *E. coli*) present critical limitations such as antibiotic resistance markers, compromised gastrointestinal-tract viability, and formulation instability during storage. In contrast, *C. butyricum*-pMTL007-GLP-1 possesses distinct advantages due to its spore-forming capability, which confers remarkable stability during product storage while also maintaining therapeutic efficacy (Wang et al., 2023). The current study provides systematic evidence that *C. butyricum*-pMTL007-GLP-1 exerts substantial neuroprotective effects in PD mice through concurrent modulation of gut microbiota composition and activation of the PI3K/AKT/GSK-3β neuroprotective signaling pathway. These findings position *C. butyricum*-pMTL007-GLP-1 as a particularly promising translational candidate for PD intervention, combining microbial therapeutic benefits with enhanced pharmaceutical properties.

This study represents significant methodological and conceptual advancements over our previous work through several key innovations. First, while our prior research employed the MPTP-induced PD mouse model — which primarily reflects toxin-mediated dopaminergic neuron degeneration (Mosharov et al., 2009; Wang et al., 2011) — the current investigation employed the A53T α-synuclein transgenic mouse model, which more accurately recapitulates both the genetic and pathological hallmarks of human PD (Smidt et al., 2000; Graham and Sidhu, 2010; Xu et al., 2021). In addition, MPTP models—specifically those that are neurotoxin-induced —exhibit secondary gut dysfunction, such as reduced SCFAs and mild dysbiosis, resulting from acute motor injury. Separately, A53T models, which are α-syn-transgenic, display primary gut pathology, including severe dysbiosis and 2-fold higher LPS levels, due to early intestinal α-syn aggregation. These features better recapitulate PD's "gut-first" axis(Lai et al., 2018; Liang et al., 2022). This strategic model selection substantially enhances the clinical relevance and translational potential of our findings. Second, we have significantly expanded the mechanistic understanding of *C. butyricum*-pMTL007-GLP-1's neuroprotective effects by comprehensively characterizing its activation of the PI3K/AKT/GSK-3β signaling cascade. Our results from this study demonstrate how this engineered probiotic concurrently mediates: (1) suppression of pro-inflammatory cytokine production and (2) inhibition of apoptotic pathways through precise molecular regulation. These findings not only validate our previous observations of therapeutic potential but provide a more rigorous, mechanism-based explanation for this probiotic's neuroprotective efficacy within a genetically relevant PD model.

Our experimental results reveal that A53T α-synuclein transgenic mice treated with *C. butyricum*-pMTL007-GLP-1 show significantly elevated GLP-1 levels in fecal samples and increased GLP-1-positive cell populations in colonic tissues, confirming enhanced intestinal GLP-1 secretion. Mechanistically, the probiotic-induced GLP-1 upregulation appears to attenuate pathological α-synuclein aggregation through phosphorylation of GSK-3β at Ser9 (p-GSK-3β^{Ser9}), effectively reducing both α-synuclein phosphorylation and oligomerization (Su et al., 2022). This regulatory mechanism helps maintain proteostatic balance while preventing the accumulation of neurotoxic α-synuclein species. Considering the well-established fact that gut-derived α-synuclein is propagated to the central nervous system (Liptak et al., 2021; Liu et al., 2021), our data indicate that *C. butyricum*-pMTL007-GLP-1 exerts its therapeutic effects by suppressing colonic α-synuclein and phosphorylated α-synuclein (p-α-synuclein) aggregation through p-GSK-3β^{Ser9} activation, consequently improving intestinal-barrier integrity. By inhibiting α-synuclein aggregation in the gut, this engineered probiotic may effectively block its pathological transmission along the gut-brain axis,

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potentially retarding PD progression. Importantly, the treatment clearly improved characteristic PD-like motor deficits in transgenic mice, including locomotor dysfunction, exploratory behavior impairment, muscle weakness, and balance-coordination deficits (Subbarayan et al., 2020). The comprehensive amelioration of both molecular pathologies and behavioral symptoms provides strong evidence for the neuroprotective efficacy of *C. butyricum*-pMTL007-GLP-1, highlighting its potential as a novel disease-modifying therapy for PD that targets both symptom management and underlying disease progression.

Expanding upon previous findings that demonstrate the efficacy of engineered *C. butyricum*-pMTL007-GLP-1 in modulating gut microbiota composition in MPTP-induced PD rodent models (Wang et al., 2023), we investigated its therapeutic potential in A53T α-synuclein transgenic mice, a genetically relevant PD model. Our results confirm that this engineered probiotic not only restores intestinal microbial diversity but specifically increases the relative abundance of *Prevotella*, a bacterial genus consistently shown to be depleted in both clinical PD populations and preclinical models (Lin et al., 2019). *AF12*, which is typically pro-inflammatory, disappears in PD mice but partially recovers after probiotic treatment, suggesting that PD creates a hostile gut environment. Engineered probiotics promote its non-pathogenic repopulation — a balanced state that is potentially vital for microbiome function and is consistent with reported benefits in metabolism and gut barrier integrity (Lai et al., 2018; Zhao et al., 2020). These findings significantly extend previous work by demonstrating the microbiota-modulating effects of *C. butyricum*-pMTL007-GLP-1 across distinct PD models while validating *Prevotella* as a key microbial marker associated with PD pathology.

The critical involvement of neuroinflammatory mechanisms in PD pathogenesis has been well documented (Tansey and Goldberg, 2010; Wang et al., 2015). Here, we systematically evaluated the anti-neuroinflammatory effects of engineered *C. butyricum*-pMTL007-GLP-1 in PD mouse models. In a study by Rojo et al. (2010), IF and Western blotting analyses revealed marked upregulation of glial activation markers (GFAP and Iba1) in PD mice (AM group), which was significantly attenuated by *C. butyricum*-pMTL007-GLP-1 treatment. According to Yan et al. (2014), the engineered probiotic also effectively suppresses key pro-inflammatory cytokines (IL-1β, IL-6, and TNF-α), demonstrating potent anti-inflammatory activity. Mechanistically, we identified that GLP-1 receptor activation by the engineered bacteria initiates PI3K/AKT signaling, as evidenced by increased p-PI3K and p-AKT levels coupled with reduced p-p65 expression in brain tissues (Cui et al., 2016; Yao et al., 2021). Furthermore, we observed enhanced phosphorylation of GSK-3β at Ser9, a crucial downstream effector of PI3K/AKT signaling known to counteract both inflammatory and apoptotic pathways (Credle et al., 2015). Together, these findings establish that *C. butyricum*-pMTL007-GLP-1 exerts neuroprotective effects in PD through multifaceted mechanisms involving suppression of neuroinflammation, inhibition of apoptotic signaling, and promotion of neuronal survival via the PI3K/AKT/GSK-3β axis.

While this study provides compelling evidence for the neuroprotective effects of engineered *C. butyricum*-pMTL007-GLP-1 in PD mouse models, several limitations should be acknowledged. First, although we identified the PI3K/AKT/GSK-3β pathway as a key mediator of the probiotic's anti-inflammatory and anti-apoptotic effects, the precise molecular interactions between GLP-1 receptor activation and downstream signaling remain incompletely elucidated. Second, while gut microbiota modulation was observed, the direct contribution of specific bacterial taxa (e.g., *Prevotella*) to neuroprotection remains speculative. Finally, the long-term safety and efficacy of engineered probiotics in PD treatment remain unexplored.

5 Conclusions

In summary, our findings demonstrate that *C. butyricum*-pMTL007-GLP-1 exerts comprehensive neuroprotective effects in PD mice through multiple synergistic mechanisms: (1) amelioration of motor dysfunction via reduction of α-synuclein expression and enhancement of intestinal-barrier integrity; (2) restoration of gut microbial homeostasis, including normalization of *Prevotella* abundance; and (3) attenuation of neuropathological changes through decreased nigral p-α-syn accumulation and upregulation of TH, DAT,

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and GLP-1R expression. These therapeutic benefits are mediated through coordinated suppression of proinflammatory cytokines and activation of the PI3K/AKT/GSK-3β signaling cascade, which concurrently inhibits neuroinflammatory responses and promotes neuronal survival. Collectively, our results provide compelling preclinical evidence supporting *C. butyricum*-pMTL007-GLP-1 as a promising multifactorial therapeutic strategy for PD, targeting both gastrointestinal and central nervous-system pathologies through gut-brain-axis modulation.

Data availability statement

The datasets analyzed in this study are available from the corresponding author upon reasonable request. Raw sequences have been deposited in the GenBank database under accession number PRJNA 1091924.

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Author contributions

Xin FANG conceived the study, designed the methodology, and provided financial support. Yun WANG developed the methodology, performed formal analysis, and wrote the original draft. Zhenli LONG, Bin LIAO, Bo WANG and Daojun HONG conducted investigations and data curation. Jie LUO secured funding, supervised the project, and contributed to manuscript revisions. Tingtao CHEN conceptualized the study, provided supervision, and acquired funding. All authors reviewed and approved the final version of the manuscript. All authors read and approved the final manuscript and, therefore, had full access to all the data in the study and take responsibility for the integrity and security of the data.

Compliance with ethics guidelines

Xin FANG, Yun WANG, Zhenli LONG, Bin LIAO, Bo WANG, Daojun HONG, Jie LUO, and Tingtao CHEN declare that they have no conflict of interest.

Animal care protocols and all experimental procedure adhered to National Institutes of Health guidelines and were approved by the Animal Experimental Ethical Inspection Committee of Nanchang Royo Biotechnology Co., Ltd., Nanchang, China (Ethical Approval Number: RyE2021070912).

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Supplementary information:

Table S1-S2; Fig. S1

Supplementary Fig. S1: mRNA expression levels of pro-inflammatory mediators. (a) IL-1β; (b) IL-6; (c) TNF-α; (d) mRNA expression level of GLP-1R in the SN; (e-g) ELISA analysis of concentrations of IL-1β (e), IL-6 (f), and TNF-α (g) in serum; (h) ELISA analysis of GLP-1 concentration in serum. C: Normal mice (n = 3); AM: PD mice (n = 3); AL: Liraglutide-treated PD mice (n = 3); ACB: *C. butyricum*-treated PD mice (n = 3); ACBG: *C. butyricum*-pMTL007-GLP-1-treated PD mice (n = 3). Data were analyzed by one-way analysis of variance (ANOVA) with Tukey's post hoc test for multiple comparisons. Data are presented as mean ± standard deviation (SD). $*p < 0.05$, $**p < 0.01$.

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