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Purified isolation of vacuoles from *Sedum alfredii* leaf-derived protoplasts

Key words: Hyperaccumulation, *Sedum alfredii*, Protoplast, Vacuole, Isolation, Purification

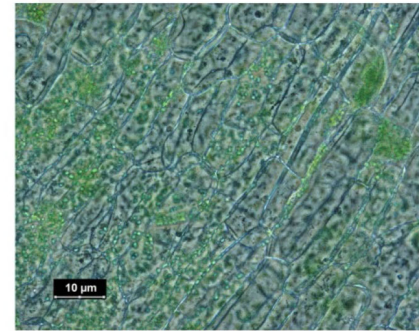
Research Summary

This study aims to develop a method for isolating and purifying protoplasts/vacuoles from fresh leaves of a Cd hyperaccumulator plant species, *Sedum alfredii*.

Main processes

- ◆ Preheating cellulase and macerozyme at **50°C for 5 min** significantly accelerated the cell wall degradation(a).

[A] Control



[B] Preheated



Figure 1 Microscope images of hydrolyzed leaf tissues of *Sedum alfredii* after vibration.

Main processes

◆ For the most optimal conditions for mesophyll protoplast isolation, this was followed by **2-hour-long vibration**.

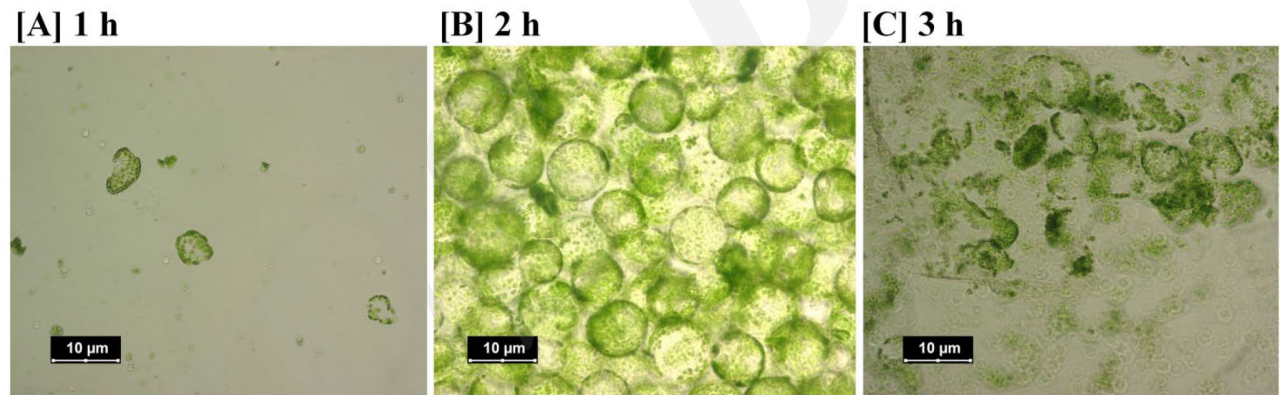


Figure 2 Microscope images of resulted protoplasts (A, B, and C) of *Sedum alfredii* after vibration.

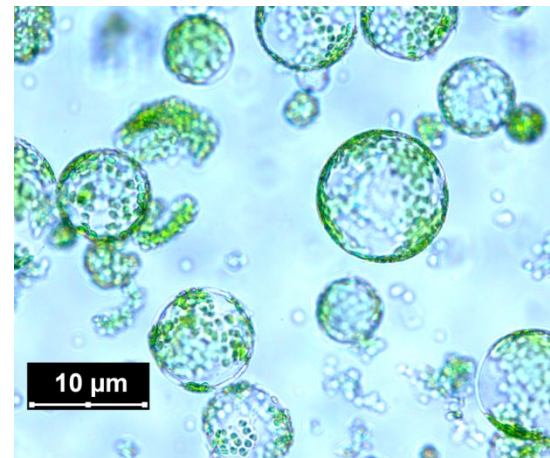
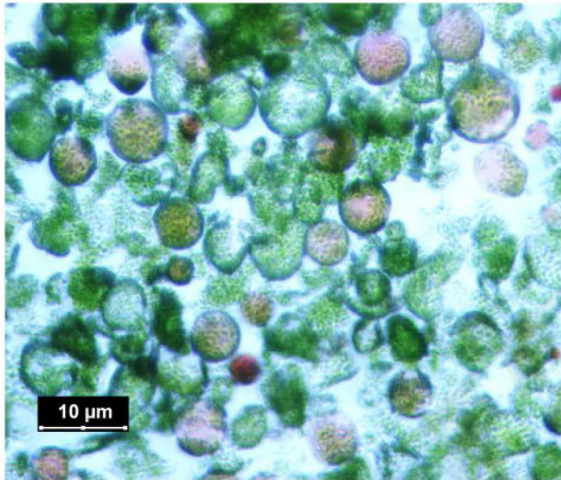


Figure 3 Microscopic image of resulted protoplasts from young leaves of *Sedum alfredii* after centrifugation.

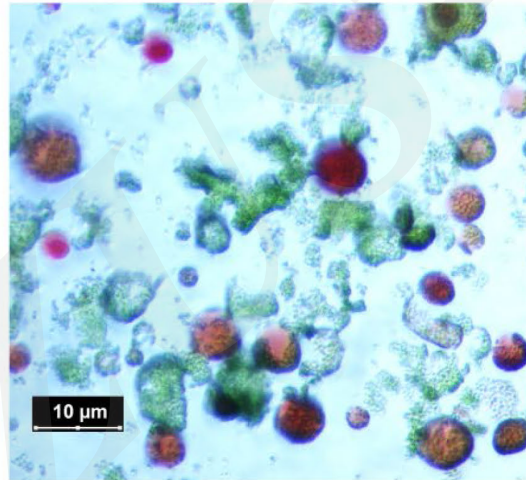
Main processes

- ◆ The protoplast lysate for vacuole isolation was diluted, where by 0.675 mM was identified as the most appropriate CHAPS level.

[A] 0.450 mM CHAPS



[B] 0.675 mM CHAPS



[C] 0.900 mM CHAPS

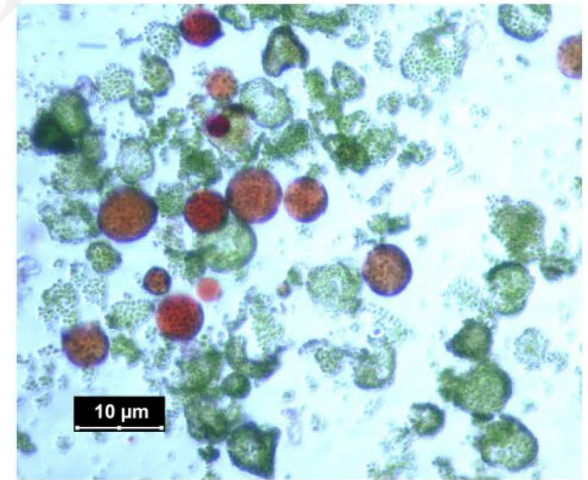
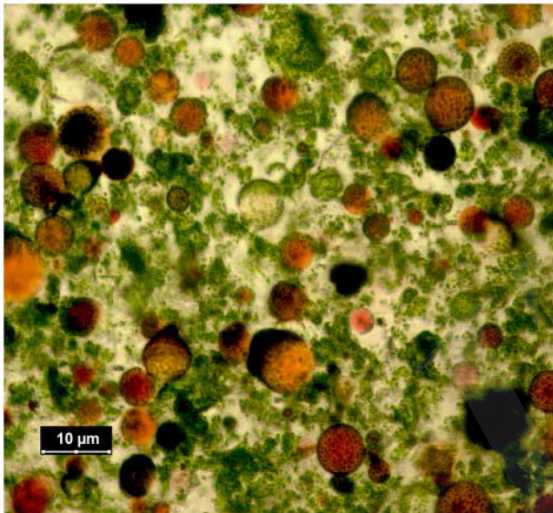


Figure 4 Microscope images of vacuoles and protoplasts after treated with protoplast lysate for 1 hour. The vacuoles were marked using neutral red.

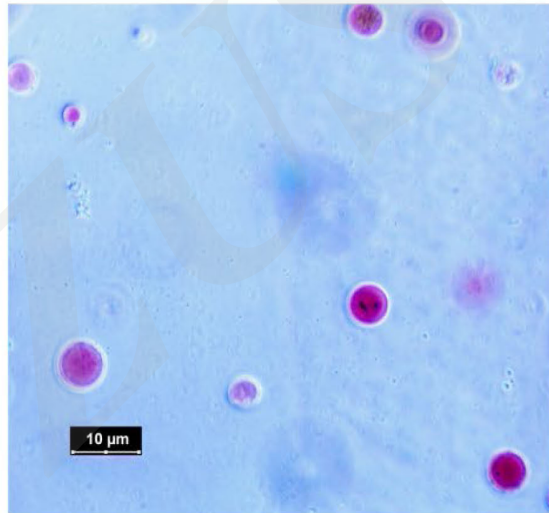
Main processes

◆ For the best vacuole purification results, we established that 0.8 M was the most optimal mannitol level in vacuole buffer in terms of vacuole protection during centrifugation, whereas Ficoll concentration of 10% was adopted in the density gradient centrifugation.

[A] 5% Ficoll



[B] 10% Ficoll



[C] 15% Ficoll

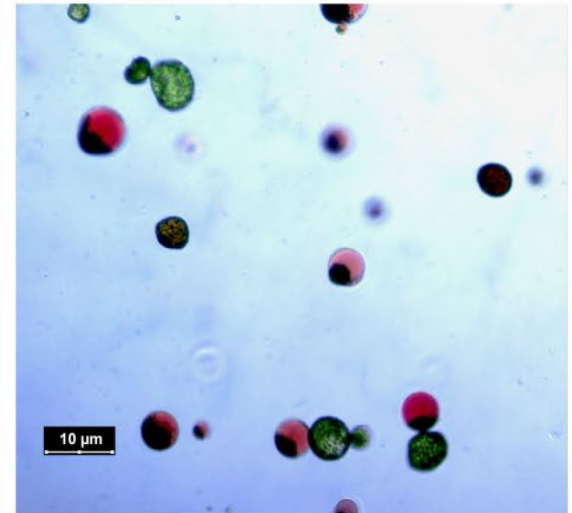


Figure 5 Microscope images of vacuoles after centrifuged at 1500 g and 10°C for 5 min. The vacuoles were marked using neutral red