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Review

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Sense codon reassignment: from global proteome profiling to precision protein engineering

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Abstract: The incorporation of non-canonical amino acids (ncAAs) has expanded the chemical diversity and functional repertoire of proteins beyond the 20 canonical amino acids (cAAs). While stop codon suppression introduces a single orthogonal decoding event, sense codon reassignment actively manipulates endogenous translational competition within the ribosomal decoding center. This strategy enables both proteome-wide stochastic labeling and precise site-specific modification of target proteins. In this review, we highlight recent advances in sense codon recoding strategies and their applications in nascent proteome profiling and precision protein engineering. We first discuss global metabolic labeling using endogenous aminoacyl-transfer RNA (tRNA) synthetases and the programmable stochastic orthogonal recoding of translation (SORT) platform. We then examine site-specific approaches, including competitive rare codon recoding (RCR) in mammalian cells, genome-scale codon compression in *Escherichia coli*, and the creation of blank coding channels through total genome synthesis. These complementary paradigms have enabled high-resolution spatiotemporal mapping of nascent proteomes in live animals, multiplexed incorporation of up to five distinct ncAAs in a single mammalian protein, and the in vivo biosynthesis of complex sequence-defined polymers. By converting translational competition into an engineerable parameter, sense codon reassignment offers new opportunities to transform our ability to decode complex biological networks and develop next-generation molecular medicines.

Key words: Sense codon reassignment; Genetic code expansion (GCE); Stochastic orthogonal recoding of translation (SORT); Rare codon; Orthogonal translation system (OTS)

1 Introduction

The universal genetic code is among the most highly conserved features of biology. It uses 61 sense codons to encode the 20 canonical amino acids (cAAs) and 3 stop codons to terminate translation. Although this decoding system underpins biological inheritance and protein evolution, the chemical functionalities offered by the 20 cAAs are inherently limited (Huang et al., 2025). As modern chemical biology seeks to interrogate complex cellular processes and engineer proteins with new chemical or biophysical properties, expanding the coding capacity of living systems has emerged as a central challenge.

Nature itself provides precedents for genetic code expansion (GCE) through the reassignment of translational termination signals. In certain organisms, the stop codons UGA and UAG are naturally repurposed to encode selenocysteine and pyrrolysine, respectively (Chambers et al., 1986; Srinivasan et al., 2002). Inspired by these natural recoding events, extensive efforts have focused on reprogramming stop codons, particularly the amber codon, to enable the site-specific incorporation of non-canonical amino acids (ncAAs) (Noren et al., 1989; Liu and Schultz, 1999). These stop codon suppression strategies have laid the technological foundation of modern GCE and have driven transformative advances

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in protein engineering (Drienovská and Roelfes, 2020; Zhao et al., 2023), chemical biology (Ding et al., 2024a; Yi et al., 2024), and therapeutic development (Axup et al., 2012; Si et al., 2016). Because stop codon suppression has matured into a well-established field and has been comprehensively reviewed elsewhere (Niu and Guo, 2024; Huang et al., 2025; Osgood et al., 2025), it is not the focus of this review.

Instead, we focus on the emerging field of sense codon reassignment. Beyond the three stop codons, the genetic code contains a much larger and largely untapped coding landscape represented by the 61 sense codons (Jones and Hartman, 2024). Reprogramming sense codons, however, presents a fundamentally distinct challenge. Unlike stop codons, which are normally excluded from translation elongation, sense codons are actively decoded by endogenous aminoacylated transfer RNAs (tRNAs). Consequently, sense codon reassignment requires the active manipulation of translational competition within the ribosomal decoding center. This competition-centric framework has given rise to two complementary paradigms: (1) harnessing endogenous competition to achieve stochastic proteome-wide incorporation of ncAAs for metabolic labeling and system-level interrogation; and (2) overcoming or eliminating endogenous competition to establish dedicated decoding pathways for precise, site-specific protein modification.

In this review, we first examine strategies that exploit endogenous translational competition for global proteome profiling and dynamic interrogation of cellular processes. We then discuss advances in site-specific sense codon reassignment, focusing on competitive rare codon recoding (RCR), genome-scale codon compression, and the development of orthogonal recoding channels for multiplexed ncAA incorporation. Ultimately, we highlight how transforming translational competition from a biological constraint into an engineerable parameter is expanding the functional scope of the genetic code and opening versatile new opportunities for basic biological discovery and next-generation molecular medicine.

2 Sense codon reassignment strategies for global proteome labeling

The cellular proteome dictates cell state and function. Dynamic changes in protein expression drive critical processes such as cell division, differentiation, stress adaptation, and disease progression (Chen et al., 2016; Phillips et al., 2023). Traditional proteomics captures mainly steady-state protein abundance, leaving the low-abundance nascent proteome masked by highly abundant pre-existing proteins (Zhang et al., 2014; Tang and Chen, 2023). Global proteome labeling via sense codon reassignment elegantly overcomes this limitation by chemically tagging actively translating proteins, thereby distinguishing the nascent proteome from the pre-existing pool. Current technologies diverge into two conceptual frameworks depending on how the host translational machinery is manipulated: (1) global metabolic labeling that leverages endogenous systems; and (2) programmable orthogonal translation systems (OTSs) for stochastic recoding (Figs. 1a and 1b).

2.1 Global metabolic labeling via endogenous translation machinery

The earliest approaches relied on selective pressure incorporation (SPI), an auxotroph-based strategy advanced by Budisa et al. (1995) and Kiick et al. (2002). SPI exploits the substrate promiscuity of endogenous aminoacyl-tRNA synthetases (aaRSs), allowing ncAAs to compete with cAAs for incorporation at specific sense codons (Fig. 1a). This framework established endogenous aaRS promiscuity as a practical entry point for residue-specific and proteome-scale ncAA incorporation (Budisa, 2004; Lepthien et al., 2010). Although effective, reliance on natural aaRS promiscuity imposed severe metabolic stress, low incorporation efficiency, and a narrow repertoire of compatible ncAAs. These limitations prompted the engineering of synthetases with enhanced ncAA selectivity, greatly expanding the accessible chemical space and enabling robust labeling under near-physiological conditions.

2.1.1 Global metabolic labeling based on endogenous synthetases

Bioorthogonal non-canonical amino acid tagging (BONCAT) pioneered the efficient profiling of the global nascent proteome by exploiting the substrate promiscuity of wild-type methionyl-tRNA synthetase (MetRS) (Dieterich et al., 2006). Methionine analogues such as azidohomoalanine (AHA) or homopropargylglycine (HPG) are incorporated at AUG codons, enabling large-scale co-translational labeling without complex genetic manipulation of the host (Fig. 2a) (Ngo and Tirrell, 2011).

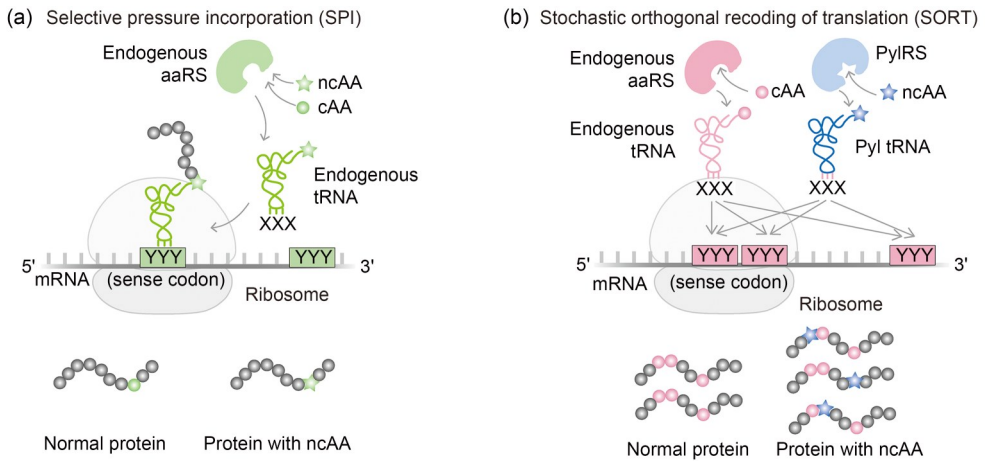


Fig. 1 Strategies for global proteome labeling via sense codon recoding. (a) Selective pressure incorporation (SPI). SPI exploits the promiscuity of endogenous synthetases (or mutated endogenous synthetases) to charge endogenous cognate transfer RNA (tRNA) (with anticodon XXX) with a structurally similar non-canonical amino acid (ncAA) instead of the canonical amino acid (cAA). The ncAA competes directly with the cAA during translation, leading to global substitution at target sense codons (YYY) and yielding a mixture of normal and ncAA-modified proteins. (b) Stochastic orthogonal recoding of translation (SORT). An orthogonal pyrrolysyl-tRNA synthetase (PylRS)/pyrrolysyl-tRNA (Pyl tRNA) pair is introduced and engineered with a mutated anticodon (XXX) to target specific sense codons (YYY). The ncAA-charged orthogonal tRNA competes stochastically with the endogenous cAA-charged tRNA for the same target sense codon, resulting in a proteome-wide statistical mixture of normal and ncAA-modified proteins. aaRS: aminoacyl-tRNA synthetase; mRNA: messenger RNA.

Coupling BONCAT with downstream detection methods has generated powerful multidimensional platforms. Fluorescent non-canonical amino acid tagging (FUNCAT) uses click chemistry with fluorescent probes for in situ spatial visualization of nascent proteins within cellular or tissue microenvironments (Dieterich et al., 2010). Quantitative non-canonical amino acid tagging (QuaNCAT) integrates BONCAT with stable isotope labeling with amino acids in cell culture (SILAC) to selectively enrich and quantify nascent polypeptides while eliminating interference from pre-existing proteins (Howden et al., 2013). These approaches have successfully mapped dynamic proteomic shifts in diverse biological contexts, including pathological cardiac remodeling and embryonic development (Liu et al., 2016).

A major advantage of wild-type-based labeling is its direct applicability in vivo without genetic engineering. Systemic administration of AHA or HPG in wild-type mice achieves whole-animal labeling, including efficient crossing of the placental barrier for high-contrast imaging of nascent proteomes in developing embryonic tissues (Calve et al., 2016).

Nevertheless, wild-type MetRS strongly prefers natural methionine (about a 400-fold substrate preference) (Hinz et al., 2012), necessitating methionine depletion in culture (Anim et al., 2025) or excessively high analogue concentrations in vivo (Calve et al., 2016). These non-physiological conditions perturb cellular metabolic homeostasis, trigger stress responses, alter translation rates, and can distort the very physiological states under investigation (Michels et al., 2021). Prolonged exposure to high concentrations of methionine analogues also induces developmental toxicity in animal and plant models, limiting long-term in vivo utility (Tivendale et al., 2021).

2.1.2 Engineered synthetases for enhanced profiling

To overcome these limitations, researchers engineered the substrate-binding pocket of synthetases to improve ncAA selectivity. A landmark example is the *Escherichia coli* MetRS mutant (NLL-MetRS, harboring L13N/Y260L/H301L), which exclusively recognizes azidonorleucine (ANL) and remains inert to the endogenous host MetRS (Tanrikulu et al., 2009) (Fig. 2b). This cell-selective labeling enabled the specific capture of bacterial virulence effectors during infection of macrophages, revealing distinct secretion profiles of the type III secretion system (T3SS) (Mahdavi et al., 2014).

Moreover, engineered synthetase strategies have been adapted to mammalian systems. A mutant *Mus musculus* MetRS (*Mm*MetRS, L274G) was developed for efficient ANL incorporation in mammalian cell lines under standard culture conditions, completely obviating

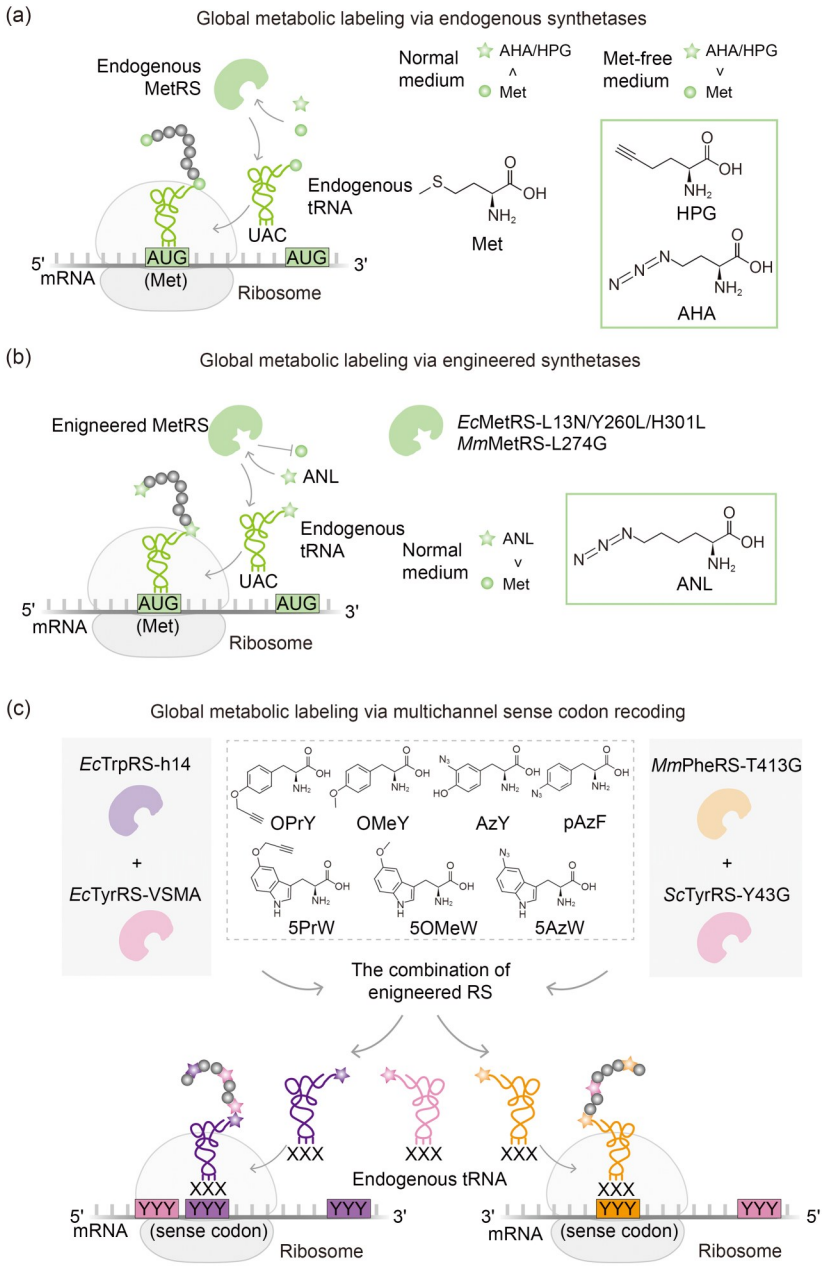


Fig. 2 Global metabolic labeling and multichannel sense codon recoding strategies. (a) Global metabolic labeling via endogenous synthetases. Endogenous methionyl-transfer RNA (tRNA) synthetase (MetRS) charges its cognate tRNA with non-canonical amino acid (ncAA) analogues like azidohomoalanine (AHA) or homopropargylglycine (HPG). Due to competitive inhibition by natural Met, the incorporation of AHA/HPG requires metabolic starvation in a Met-free medium. (b) Global metabolic labeling via engineered synthetases. Mutant synthetases (*Ec*MetRS-L13N/Y260L/H301L and *Mm*MetRS-L274G) specifically recognize and activate analogues like azidonorleucine (ANL), enabling direct ncAA incorporation in normal medium. (c) Global metabolic labeling via multichannel sense codon recoding. Co-expressing combinations of engineered synthetases (*Ec*TrpRS-h14, *Ec*TyrRS-VSMA, *Mm*PheRS-T413G, and *Sc*TyrRS-Y43G) enables the simultaneous targeting of multiple sense codons. The chemical structures of representative ncAAs used for multiplexed labeling, including analogues of tyrosine (OPrY, OMeY, and AzY), phenylalanine (pAzF), tryptophan (5PrW, 5OMeW, and 5AzW), are shown. Met: methionine; OPrY: *O*-propargyl-L-tyrosine; OMeY: *O*-methyl-L-tyrosine; AzY: azido-L-tyrosine; pAzF: para-azido-L-phenylalanine; 5PrW: 5-propargyl-L-tryptophan; 5OMeW: 5-methoxy-L-tryptophan; 5AzW: 5-azido-L-tryptophan; mRNA: messenger RNA; *Ec*MetRS: *Escherichia coli* methionyl-tRNA synthetase; *Mm*MetRS: *Mus musculus* methionyl-tRNA synthetase; *Sc*TyrRS: *Saccharomyces cerevisiae* tyrosyl-tRNA synthetase; *Mm*PheRS: *M. musculus* phenylalanyl-tRNA synthetase; *Ec*TrpRS: *E. coli* tryptophanyl-tRNA synthetase; *Ec*TyrRS-VSMA: *E. coli* tyrosyl-tRNA synthetase harboring Y37V/D182S/F183M/L186A.

the need for methionine depletion (Mahdavi et al., 2016). Building on this foundation, a recently developed transgenic mouse line expressing multiple copies of this mutant enzyme (3×MetRS, L274G) achieves highly efficient global proteome labeling in vivo. By fundamentally mitigating metabolic stress, this engineered synthetase approach broadens the scope of application for studying physiological and pathological homeostasis in complex living organisms (Alvarez-Pardo et al., 2025).

2.1.3 Multiplexed profiling via expanded codon channels

Although AUG-targeted BONCAT has been widely used, relying exclusively on this single sense codon intrinsically limits the comprehensiveness of the captured proteome. Methionine residues are frequently subject to post-translational modifications (PTMs), such as N-terminal excision, which can irreversibly remove the incorporated chemical tags (Giglione et al., 2004; Wang et al., 2008). Furthermore, forcing analogue incorporation exclusively at methionine sites can occasionally perturb local protein folding and stability, causing certain protein subsets to be systematically missed (Yang et al., 2018). Consequently, any single-codon labeling system inevitably captures a biased subproteome, leaving significant blind spots. To achieve unbiased, global proteomic coverage, researchers have had to develop alternative metabolic labeling strategies targeting other sense codons. This pursuit has driven the successful expansion of the labeling repertoire to threonine, tyrosine, phenylalanine, and tryptophan codons (Yang et al., 2018; Ignacio et al., 2023; Loynd et al., 2026), paving the way for both deeper proteome capture and advanced multiplexing.

Parallel to these efforts, engineered synthetases have further expanded the codon pool to capture unique proteomic subsets. Researchers developed a *Saccharomyces cerevisiae* tyrosyl-tRNA synthetase (*Sc*TyrRS, Y43G) and an *M. musculus* phenylalanyl-tRNA synthetase (*Mm*PheRS, T413G) to redirect metabolic labeling toward tyrosine and phenylalanine sense codons (Yang et al., 2018) (Fig. 2c). These variants preferentially activate the corresponding azide-bearing ncAAs over cAAs under the reported labeling conditions, enabling residue-specific bioorthogonal tagging in live tumor models. By labeling the melanoma tumor proteome and plasma secretome in mice, *Sc*TyrRS-Y43G and *Mm*PheRS-T413G achieved the first global proteome labeling in a tumor model and identified plasma factors secreted from specific cell types. Furthermore, integrating these distinct synthetases captured unique proteomic subsets that single channel labeling inherently missed.

Most recently, these expansion efforts have culminated in highly sophisticated spatial and temporal control. Multiplexed BONCAT (mBONCAT) introduced engineered *E. coli* tryptophanyl-(*Ec*TrpRS-h14) and tyrosyl-tRNA synthetases (*Ec*TyrRS-VSMA [Y37V/D182S/F183M/L186A]) (Fig. 2c). A defining advantage of these engineered enzymes is their broad substrate compatibility, accommodating both click-reactive ncAAs and inert analogues (e.g., 5-methoxy-L-tryptophan [5OMeW] or *O*-methyl-L-tyrosine [OMeY]). This distinct property facilitates a precise pulse-chase modality: high-affinity inert ncAAs can competitively terminate the reactive pulse, granting exceptional temporal resolution. Coupled with distinct fluorescent probes, the strict orthogonality of these integrated systems enables the simultaneous tracking of preexisting and nascent protein dynamics within the same cellular environment (Loynd et al., 2026).

Despite these optimization strategies effectively mitigating metabolic stress and expanding targeted codons from AUG to the sense codons of tryptophan, tyrosine, phenylalanine, and threonine, such systems still face fundamental mechanistic constraints. Although engineered synthetases have successfully accommodated a broad array of chemically diverse ncAAs, these approaches remain inextricably tethered to the specific codons dictated by their cognate host tRNAs. They completely lack the modular flexibility required for the customized recoding of arbitrary codons, as altering the anticodon loop of endogenous tRNAs to redirect them to new codons would catastrophically disrupt the host's natural genetic code and global proteostasis.

2.2 SORT as a programmable platform for global proteome labeling

To establish an orthogonal and versatile strategy for ncAA-mediated proteome labeling, Elliott et al. (2014a) repurposed the highly orthogonal pyrrolysyl-tRNA synthetase (PylRS)/tRNA pair, originally developed for site-specific ncAA incorporation, to create stochastic orthogonal recoding of translation (SORT) for global proteomic applications (Fig. 1b). The defining feature of SORT is the largely anticodon-independent

aminoacylation activity of PylRS, which enables pyrrolysyl-tRNA (PylT) anticodons to be reprogrammed toward a broad range of compatible sense codons while retaining aminoacylation activity. In practice, however, successful recoding efficiency is dictated by multiple factors, including endogenous tRNA abundance (Schwark et al., 2020), messenger RNA (mRNA) sequence context (Ding et al., 2024b), aminoacylation efficiency (Bruce and Uhlenbeck, 1982), and necessary tRNA engineering. Upon expression in cells, these optimized orthogonal tRNAs compete with endogenous tRNAs to stochastically decode selected codons with ncAAs during translation.

This unique architecture transformed stochastic proteome labeling from a largely amino acid-restricted strategy into a programmable codon-level recoding platform. By altering the orthogonal tRNA anticodon, ncAA incorporation can be redirected toward distinct codon populations while preserving efficient aminoacylation. This programmability enables flexible tuning of labeling density, proteome coverage, and subproteome selectivity according to the abundance and positional distribution of different amino acids across the proteome.

2.2.1 SORT enabling programmable multi-codon proteome labeling and enrichment

Building on this programmable foundation, the SORT toolkit has rapidly evolved. The following sections detail its expansion from initial global proteome visualization (SORT-M) and deep nascent protein identification (SORT-E) to spatiotemporal mapping in live mammalian disease models, establishing SORT as a robust engine for in vivo chemical proteomics.

Elliott et al. (2014b) introduced SORT with chemoselective modification (SORT-M). In this approach, engineered PylRS/tRNA pairs were used to stochastically incorporate ncAAs bearing bioorthogonal chemical handles into proteins throughout the proteome at designated sense codons. The incorporated handles, including cyclopropene-containing ncAAs, subsequently reacted with tetrazine-conjugated probes through rapid inverse electron-demand Diels–Alder (IEDDA) cycloaddition (Blackman et al., 2008; Patterson et al., 2012). Owing to its exceptionally fast kinetics and high selectivity, this reaction enabled efficient visualization and enrichment of nascent proteins while minimally perturbing cellular physiology.

The versatility of SORT-M was systematically demonstrated across multiple biological systems. Robust proteome-wide labeling with favorable signal-to-noise ratios was achieved in both *E. coli* and human HEK293T cells. To establish the generality of the platform, researchers engineered orthogonal tRNAs targeting a broad range of sense codons, including those encoding Met (AUG), Lys (AAA and AAG), Ser (AGU and AGC), Ala (GCA), Leu (CUG), Val (GUG), Glu (GAG), Tyr (UAU and UAC), and Trp (UGG), all of which successfully supported ncAA incorporation and proteome labeling. Importantly, comprehensive analyses in both bacterial and mammalian systems demonstrated that systemic sense codon recoding under optimized conditions caused minimal cytotoxicity and preserved overall cellular fitness (Elliott et al., 2014b).

The application of SORT was subsequently extended to multicellular organisms through a modular transgenic platform in *Drosophila melanogaster* using the UAS/GAL4 system, a GAL4-dependent transgenic expression system (Brand and Perrimon, 1993). Tissue-specific expression of PylRS enabled spatiotemporally resolved imaging of nascent proteomes in defined cell populations, including ovarian germline cells and wing imaginal discs. For example, expression driven by *nos-vp16-GAL4* produced strong labeling in stage 5 egg chambers with minimal signal in surrounding follicle cells, while causing no detectable developmental abnormalities (Elliott et al., 2014b).

To further enhance nascent proteome identification, researchers subsequently developed SORT with enrichment (SORT-E), which incorporated a cleavable tetrazine diazobenzene biotin (TDB) probe into the workflow (Verhelst et al., 2007; Szychowski et al., 2010; Elliott et al., 2016). Leveraging the ultrafast kinetics of the IEDDA reaction (Dommerholt et al., 2014), TDB enabled direct capture of labeled proteins from crude lysates at low reagent concentrations without precipitation-based processing. This streamlined workflow minimized sample loss and substantially improved recovery of low-abundance proteins that are frequently inaccessible in conventional whole-cell proteomics.

In *E. coli*, SORT-E achieved a highly specific enrichment of tagged proteins, with a specificity exceeding 92%. Quantitative comparisons against comprehensive protein abundance databases (such as PaxDb) (Wang et al., 2012, 2015) confirmed its preferential enrichment of low-abundance proteins frequently lost

in standard whole-cell analyses. Importantly, distinct proteomic subsets exhibit codon-specific enrichment preferences. While earlier methodologies suggested that tagging exclusively at methionine residues might be sufficient for proteome-wide analysis (Yuet and Tirrell, 2014), these codon-specific biases highlight the critical need for multi-codon strategies. By co-expressing multiple orthogonal tRNAs targeting codons for Ala, Ser, and Met, labeling coverage and detection sensitivity were substantially improved, thereby enabling high-resolution proteomic mapping in complex tissues. When applied in vivo to *Drosophila* ovarian germline cells via the *nos-vp16-GAL4* system, this combinatorial approach dramatically increased nascent protein identification compared to single-codon controls (Elliott et al., 2016). Overall, SORT-E offers a powerful and necessary tool for decoding complex, cell-specific proteomic dynamics during development.

2.2.2 Spatiotemporal mapping in complex mammalian organs and disease models

The extension of SORT to live animal models represents a major milestone in in vivo chemical proteomics. Krogager et al. (2018) delivered SORT components via adeno-associated viruses (AAVs) to the mouse brain using tissue-specific promoters (glial fibrillary acidic protein [*GFAP*], human synapsin 1 [*hSyn1*], vesicular GABA transporter [*Vgat*], and vesicular glutamate transporter [*Vglut*]) to drive expression of the engineered translation system in specific neural populations including excitatory neurons, inhibitory GABAergic neurons, and glia (Fig. 3a). Non-invasive administration of the alkynyl ncAA, N^{ε}-(propargyloxycarbonyl)-L-lysine (AlkK), through drinking water enabled in vivo tagging of nascent proteins without perturbing physiological homeostasis (Nguyen et al., 2009; Ernst et al., 2016). This approach identified nearly 2000 proteins from striatal neurons with a high enrichment specificity of 96%. This study established the feasibility of using SORT in live mammals and highlighted its utility for analyzing neural circuits and cell-specific proteomic dynamics.

Building upon the success of AAV-mediated delivery in the nervous system, recent advancements have expanded the platform to address intricate pathological environments. Gu et al. (2025) developed SORT-AC (SORT induced by AAV-delivered Cre) (Fig. 3b). This technology integrated a clustered regularly interspaced short palindromic repeats (CRISPR)-CRISPR-associated protein 9 (Cas9)-mediated knock-in mouse model with AAV-delivered Cre recombinase driven by the liver-specific transthyretin promoter (TTR-Cre), enabling precise and inducible expression of an orthogonal translation system, AlkKRS/PylT_{KASM}, in the live liver. To optimize the labeling of the nascent membrane proteome, researchers used a rational codon selection strategy (KASM). The combination of Lys (K), Ala (A), Ser (S), and Met (M) was selected based on their high abundance, enrichment in transmembrane regions, and preferential distribution on protein surfaces, which minimizes steric hindrance during subsequent chemical labeling.

By simultaneously targeting these four codons and optimizing hepatic click chemistry, the system successfully generated a high-resolution nascent membrane proteome map, termed SORT-AC Print, where membrane proteins accounted for about 50% of the newly identified liver proteome (Fig. 3c). In a model of alcohol-associated liver disease, SORT-AC further revealed the regulatory role of *Phb1/2* in alcohol metabolism and identified *Acsl1/5* as potential therapeutic targets for lipid accumulation (Gu et al., 2025). This evolution toward multi-codon rational design provides a robust toolkit for identifying low-abundance regulatory factors and screening therapeutic interventions in various disease states.

3 Sense codon reassignment strategies for site-specific incorporation

While the SPI and SORT platform provides a powerful mechanism for proteome-wide reassignment, it is inherently unsuitable for the targeted modification of a specific protein of interest (POI), as its global deployment would lead to massive, unpredictable off-target incorporation. However, achieving precise, site-specific incorporation of ncAAs is crucial (Table 1). It has revolutionized protein engineering by endowing proteins with novel chemical functionalities, enabling the real-time visualization of dynamic biological processes, the targeted regulation of protein functions, and the accelerated development of next-generation biotherapeutics.

Historically, achieving such site-directed incorporation has relied mainly on stop codon suppression (particularly the amber codon, UAG). However, this approach is intrinsically limited by intense competition with endogenous release factors, the potential disruption of natural translational termination, and a restricted

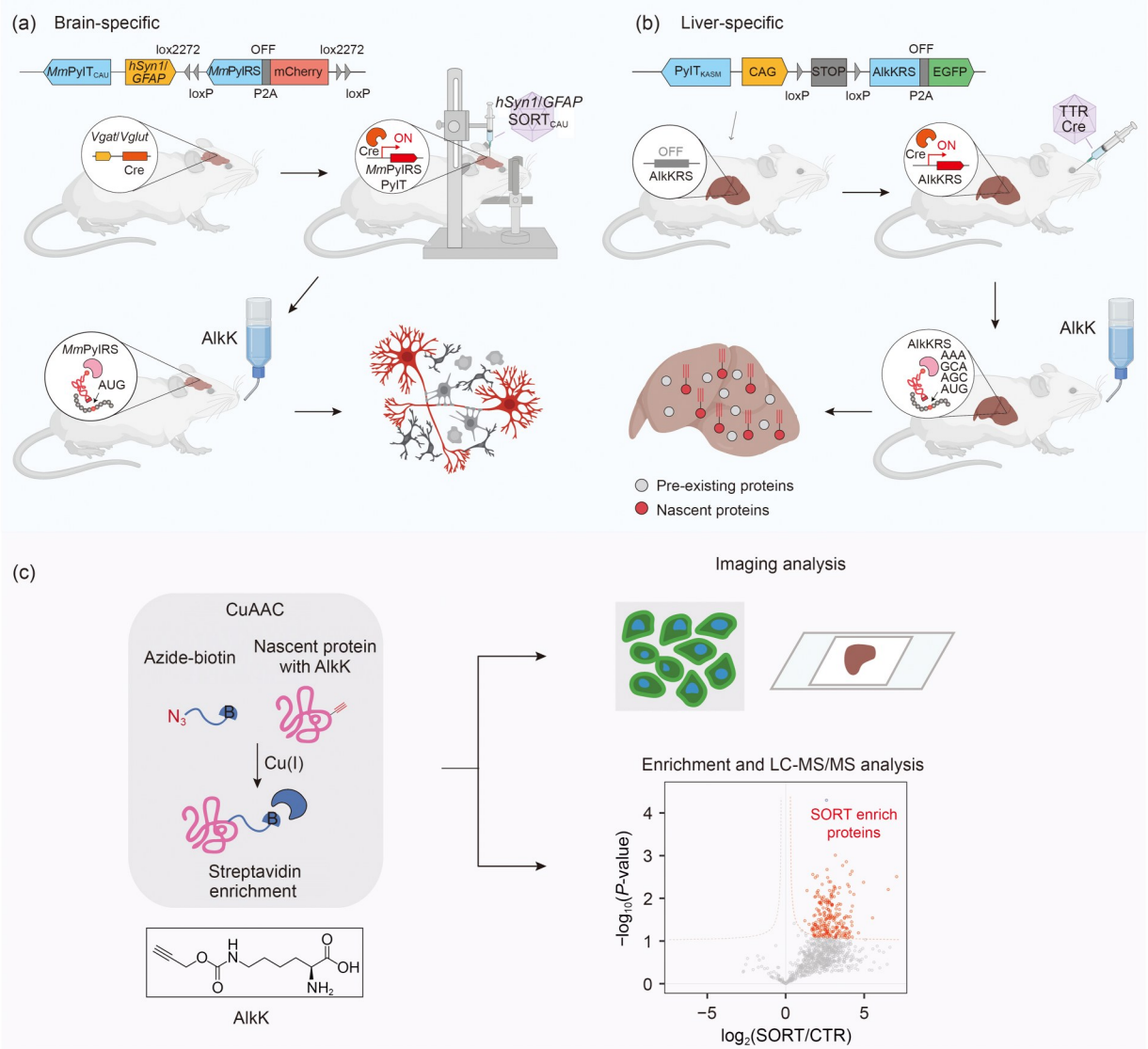


Fig. 3 In vivo tissue- and cell-type-specific nascent proteome labeling utilizing stochastic orthogonal recoding of translation (SORT) in live mammalian systems. (a) Brain-specific proteome tagging. Cre-expressing transgenic mice (*Vgat*/*Vglut*-Cre) receive stereotaxic injections of an adeno-associated virus (AAV) vector carrying Cre-dependent flip-excision (FLEX) SORT. Cre-mediated recombination of the FLEX cassette irreversibly activates *Methanosarcina mazei* pyrrolysyl-tRNA synthetase (*Mm*PylRS) and the red fluorescent reporter mCherry expression. Subsequent oral *N*$^{\varepsilon}$-(propargyloxycarbonyl)-L-lysine (AlkK) administration enables stochastic incorporation of AlkK at Met codons (AUG) within targeted neural populations. (b) Liver-specific proteome tagging. Transgenic mice harboring a genomically integrated, loxP-STOP-loxP-controlled SORT cassette receive an orbital injection of AAV-TTR-Cre. Cre-mediated excision of the STOP cassette selectively activates the engineered AlkK-specific pyrrolysyl-tRNA synthetase (AlkKRS) and enhanced green fluorescent protein (EGFP) expression in hepatocytes. Upon AlkK administration, the optimized AlkKRS/tRNA$_{KASM}$ system incorporates AlkK across four sense codons (AAA, GCA, AGC, and AUG). (c) Bioorthogonal labeling and downstream analysis. Harvested tissue lysates undergo copper-catalyzed azide-alkyne cycloaddition (CuAAC) click chemistry to conjugate azide-biotin probes to AlkK-tagged proteins. The biotin-tagged proteome supports high-resolution in situ imaging alongside subsequent streptavidin enrichment. Enriched nascent proteomes are subsequently quantified via liquid chromatography-tandem mass spectrometry (LC-MS/MS) to identify upregulated newly synthesized proteins (red dots in the volcano plot). STOP: transcriptional termination cassette; loxP: Cre-recognition site from bacteriophage P1; lox2272: mutant loxP site; TTR: transthyretin; PylT$_{KASM}$: four engineered pyrrolysyl tRNAs targeting lysine (K, AAA codon), alanine (A, GCA codon), serine (S, AGC codon), and methionine (M, ATG codon) codons; CAG promoter: composite cytomegalovirus enhancer/chicken β-actin promoter; P2A: self-cleaving 2A peptide from *Porcine teschovirus*-1; *GFAP*: glial fibrillary acidic protein; *hSyn1*: human synapsin 1; *Vgat*: vesicular GABA transporter; *Vglut*: vesicular glutamate transporter; CTR: control.

Table 1 Mechanistic comparison of sense codon reassignment strategies

Strategy	Underlying mechanism	Target codon	Required components	Host system	Incorporation mode	Site specificity	Off-target risk	Major limitations	Applications
BONCAT/SPI	Residue-level analogue incorporation via endogenous/engineered synthetases	Typically AUG (Met), or specific codons dictated by endogenous host tRNAs	Analogue (e.g., AHA/HPG/ANL)+ corresponding endogenous or engineered aaRS	Bacteria, mammalian cells, zebrafish, mice, and transgenic/viral animal model	Global metabolic labeling	Global/stochastic (residue-specific but not protein-specific)	Not applicable	Metabolic stress or amino acid starvation; residue-level bias; limited codon programmability	Nascent proteome labeling; cell-type-specific proteomics; in vivo metabolic profiling
SORT	Stochastic codon competition through engineered orthogonal aaRS/tRNAs	Programmable across various sense codons by altering orthogonal tRNA anticodons	Orthogonal aaRS/tRNA pair (e.g., PylRS/tRNA)+ ncAAs	*Escherichia coli*, HEK293T, *Drosophila*, and mouse models	Proteome-wide stochastic labeling	Proteome-wide (codon-specific but stochastic across the proteome)	Not applicable	Context-dependency of recoding efficiency	Programmable proteome-wide labeling; low-abundance protein enrichment; in vivo proteome mapping
RCR	Site-specific competitive reassignment at inherently rare codons	Rare sense codons with the lowest genomic usage and low endogenous tRNA abundance (e.g., TCG in mammalian cells)	Orthogonal aaRS/tRNA pairs+ncAAs	Mammalian cells (where whole-genome synthesis is currently impractical)	Highly efficient target incorporation overriding weak endogenous competition	High (can be promoted by context-sequence optimization)	Moderate (residual background due to endogenous tRNA competition)	Context-dependency of recoding efficiency; residual background in sensitive systems	Precision protein engineering; multiplexed ncAA incorporation
Genome-recoded organisms	True blank-codon reassignment after genome recoding and cognate tRNA deletion	Compressed/liberated sense codons completely removed from the genome (e.g., TCG, TCA in Syn61)	Synthetic genome+ deleted endogenous tRNAs+ orthogonal aaRS/tRNA+ ncAAs	Prokaryotes (e.g., highly engineered *E. coli* variants like Syn61)	Absolute incorporation without endogenous competition	Absolute (the reassigned codon is absent from the host genome and is introduced solely into the target mRNA)	None (completely interference-free)	Unattainable in complex mammalian systems due to vast scale and regulatory complexity	Programmable biosynthesis of complex sequence-defined polymers and macrocycles

BONCAT: bioorthogonal non-canonical amino acid tagging; SPI: selective pressure incorporation; SORT: stochastic orthogonal recoding of translation; ncAA: non-canonical amino acid; AHA: azidohomoalanine; HPG: homopropargylglycine; ANL: azidonorleucine; Syn61: an *E. coli* strain with a 61-codon genome, designed to enable GCE by providing blank codons for ncAAs; tRNA: transfer RNA; aaRS: aminoacyl-tRNA synthetase; PylRS: pyrrolysyl-tRNA synthetase; mRNA: messenger RNA; Met: methionine; RCR: rare codon recoding.

coding space that severely hinders the simultaneous encoding of multiple distinct ncAAs. To circumvent these bottlenecks, sense codon reassignment offers a compelling alternative. This strategy is fundamentally underpinned by the degeneracy of the genetic code. Because 18 of the 20 cAAs are encoded by multiple synonymous codons (e.g., six synonymous codons for serine and six for arginine), reassigning one specific sense codon provides a vast potential codon reservoir for site-specific ncAA encoding without abolishing the natural amino acid's incorporation across the proteome.

Despite this potential, sense codon reassignment faces three main challenges. First, many sense codons occur frequently in the genome, so their reassignment can lead to off-target ncAA incorporation at endogenous protein sites. Second, at the engineered codon in the target mRNA, the orthogonal tRNA must compete with endogenous isoacceptor tRNAs that normally decode the same codon. Third, maintaining aminoacylation orthogonality becomes problematic in multiplexed ncAA encoding. Because the repertoire of tRNA identity elements that can be used to build mutually orthogonal pairs is limited (Saint-Léger et al., 2016), different orthogonal aaRS/tRNA systems may cross-react with one another or with endogenous translation components. Furthermore, engineering aaRS substrate-recognition pockets to accommodate ncAAs can inadvertently broaden substrate specificity or compromise catalytic efficiency, resulting in mischarging with cAAs or unintended ncAAs.

Targeting rare sense codons offers a practical strategy to overcome these decoding related barriers: their low genomic usage and reliance on low-abundance endogenous tRNAs naturally reduce proteome-wide background and native decoding competition. However, ensuring aminoacylation orthogonality remains a distinct prerequisite for efficient and faithful multiplexed ncAA incorporation, necessitating further investigation into its mechanistic basis and engineering strategies. The following sections discuss how these principles have been implemented in *E. coli*, expanded via genome recoding and total synthesis, and ultimately adapted to complex mammalian systems.

3.1 Establishment and evolution of sense codon reassignment in *E. coli*

Before delving into the mechanistic strategies of sense codon reassignment, it is crucial to establish a robust evaluation methodology. Unlike traditional nonsense suppression, where successful incorporation is easily indicated by full-length protein expression versus a truncated byproduct, sense codon reassignment presents a unique challenge: a sense codon inevitably yields a full-length protein, regardless of whether it is decoded by the introduced ncAA or an endogenous cAA. Therefore, accurately distinguishing targeted ncAA encoding from the cAA background is foundational. To address this challenge, a general screen was developed using permissive active site mutations in green fluorescent protein (GFP). When a critical fluorophore-maturation residue is mutated to a target sense codon, the resulting fluorescence becomes strictly dependent on the successful incorporation of the specific ncAA, thereby providing a direct optical readout for reassignment efficiency (Biddle et al., 2015). Furthermore, the development of a fluorescence-activated cell sorting (FACS) dual-fluorescence pipeline has significantly advanced this process. This platform uses two distinct reporters to simultaneously monitor global protein expression and specific ncAA incorporation, facilitating the high-throughput screening and directed evolution of highly active orthogonal aaRSs (Biddle et al., 2022). Together, these platforms provide the essential quantitative tools required to rigorously evaluate the desired target incorporation.

Equipped with these quantitative screening tools, researchers can effectively dissect and manipulate the core mechanism underlying sense codon reassignment: translational competition. Fundamentally, the introduced OTS must actively compete against the endogenous host translational machinery to decode a specific mRNA codon. Early proof-of-concept studies successfully validated this competitive strategy by targeting rare codons. For instance, by engineering the anticodon of the PylRS/tRNA and coupling it with highly optimized cultivation conditions, such as using specific minimal media (GMML, M9 minimal medium with 1% [volume fraction] glycerol and 0.3 mmol/L leucine) and precise induction timing, the translation of the rare arginine codon AGG was forcefully hijacked (Zeng et al., 2014). Following a similar paradigm, another rare arginine codon, AGA, was also successfully reassigned using an engineered Pyl system under optimized cultivation (Wang and Tsao, 2016).

As these competitive strategies yielded initial successes, the underlying biological mechanisms governing

this competition were further elucidated. For AGG reassignment, Lee et al. (2015) demonstrated that deletion of *argW* (gene for $tRNA^{Arg}_{CCU}$) removed the major endogenous $tRNA_{CCU}$ that normally decodes AGG. Endogenous AGG decoding then depends mainly on the weaker wobble pairing by $tRNA_{UCU}$, giving the engineered orthogonal tRNA a stronger competitive advantage and thereby improving ncAA incorporation efficiency. This observation highlighted a critical strategic principle that reducing the abundance or activity of competing endogenous tRNAs in the host system significantly enhances the competitive advantage of the OTS. This paradigm was further refined by the demonstration that endogenous tRNA abundance, rather than mere codon rarity, is the definitive quantitative determinant of competitive success (Schwark et al., 2020). While codon rarity correlates with lower global toxicity, the local decoding competition is directly dictated by the cellular concentration of the competing endogenous tRNAs; theoretically, a lower abundance allows for higher ncAA incorporation efficiency. Beyond passively selecting low-competition codons, actively attenuating the decoding capacity of host's endogenous translation machinery offers a powerful alternative. For example, knocking out the host enzyme responsible for queuosine (Q) modification on endogenous tRNAs effectively reduces their decoding efficiency, thereby attenuating the endogenous machinery and shifting the translational balance in favor of the OTS (Tittle et al., 2022).

Empowered by this deeper mechanistic understanding of translational competition, researchers have rapidly expanded the boundaries of sense codon reassignment far beyond rare codons. Demonstrating the robust potential of this competitive strategy, a series of highly frequent codons in *E. coli*, including AGU, AGC, and UCG (encoding serine), as well as CUG (encoding leucine), were successfully reassigned by massively overexpressing the OTS, proving that extreme competitive pressure can overcome even highly abundant endogenous tRNAs (Ho et al., 2016). Expanding the scope further, the target was shifted from the elongation phase to the initiation phase, successfully reprogramming the initiator codon (AUG) to achieve N-terminal ncAA incorporation (Tharp et al., 2021). Remarkably, by combining sense codon reassignment with traditional stop codon suppression, the co-translational incorporation of up to three distinct ncAAs within a single complex protein has been realized (Cui et al., 2017; Tharp et al., 2021).

In summary, driven by increasingly sophisticated competitive strategies, a diverse array of codons, including AGG, AGA, AGU, AGC, UCG, CUG, and the initiator AUG, have been successfully reassigned to ncAAs in *E. coli*. However, while optimizing competitive dynamics can maximize targeted incorporation efficiency, it cannot fundamentally eliminate the proteome-wide background misincorporation and associated systemic toxicity that arise from the pervasive distribution of sense codons throughout the host genome.

3.2 Overcoming endogenous competition through genomic recoding and total synthesis

Due to the widespread distribution of sense codons across the genome, achieving targeted ncAA encoding via sense codon reassignment inevitably results in the erroneous misincorporation of ncAAs at natural synonymous sites throughout the host proteome. To address this fundamental limitation, genomic recoding and total synthesis have been developed. These approaches systematically remove all instances of a target codon from the host genome, creating a vacant coding channel that can be reassigned without interference. The technical feasibility of large-scale genomic recoding was first demonstrated through the total replacement of all 321 UAG stop codons with synonymous UAA codons in the *E. coli* genome (Lajoie et al., 2013). By successfully removing every annotated occurrence of UAG, release factor 1 (RF1) was deleted, converting UAG into a blank coding slot with zero competition from the host machinery. Building upon this, the exploration expanded toward sense codons, which present greater complexity due to their high frequency. A systematic evaluation of synonymous codon compression schemes established critical design rules for sense codon replacement (Wang et al., 2016). It was determined that while many synonymous substitutions are well-tolerated, specific positions are highly sensitive to changes that disrupt mRNA secondary structure, induce ribosome stalling, or perturb other regulatory elements, ultimately providing the essential blueprint and multi-omics troubleshooting framework for whole-genome synthesis (Nyerges et al., 2024).

Guided by these compression rules, genomic recoding progressed from localized editing to bottom-up

total synthesis. This monumental shift was realized with the creation of Syn61, an *E. coli* variant harboring a fully synthetic 4-megabase genome (Fredens et al., 2019). In this synthetic genome, over 18 000 target codons were deterministically replaced, successfully eliminating two sense codons (UCG and UCA, encoding serine) and one stop codon (UAG). Crucially, the mere removal of these codons from the genome is insufficient for orthogonal reassignment; the corresponding decoding machinery must also be eliminated. Therefore, the endogenous tRNAs responsible for decoding UCG and UCA were subsequently deleted. This dual knockout strategy, erasing both the genomic codons and their cognate tRNAs, yielded pure, blank sense codons. By introducing OTSs, these emancipated sense codons were harnessed to achieve viral resistance and the efficient, zero-background synthesis of complex sequence-defined polymers and macrocycles comprising multiple distinct non-canonical monomers in vivo (Robertson et al., 2021).

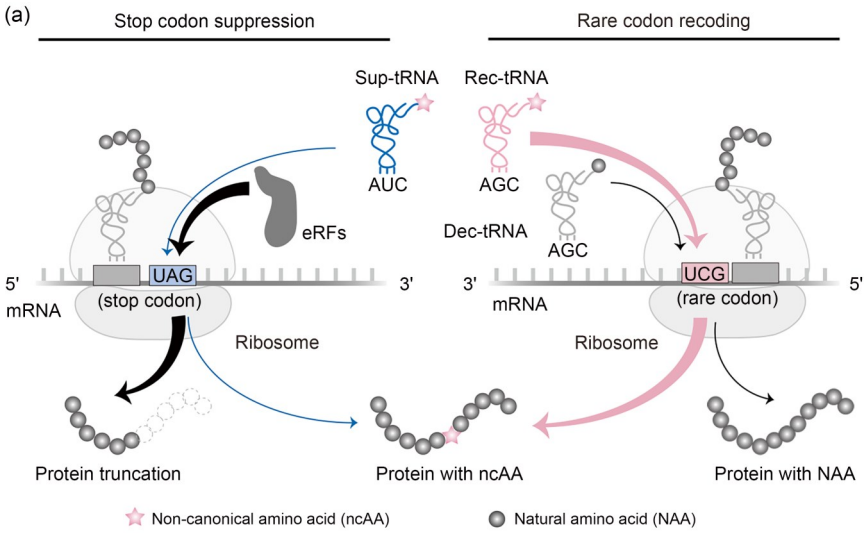
While Syn61 provided three blank codons, the demand for increasingly complex chemically modified biologics necessitates a broader repertoire of orthogonal channels. Consequently, efforts have focused on pushing the limits of genetic code compression. Recent advancements have demonstrated the feasibility of compressing the three natural stop codons into a single UAA stop codon, completely liberating the remaining two for genetic isolation and reassignment (Grome et al., 2025). Pushing this compression to its absolute current limit, a highly engineered *E. coli* strain operating on a 57-codon genetic code was successfully synthesized by removing seven distinct codons from the genome (Ostrov et al., 2016; Robertson et al., 2025). The emergence of such highly compressed genomes marks a profound paradigm shift in sense codon reassignment. Rather than struggling against endogenous competition for a single target site, researchers will soon have up to seven entirely interference-free blank codons. This expansive orthogonal coding space will empower the programmable biosynthesis of highly complex proteins and novel materials bearing extensive, multiplexed chemical modifications.

3.3 Implementation of sense codon reassignment in complex mammalian systems

While total genome synthesis and compression have successfully liberated sense codons in prokaryotes, these disruptive strategies have largely failed to translate into mammalian systems due to the vast scale and regulatory complexity of the eukaryotic genome. Unlike the highly programmable nature of synthetic bacterial genomes, the mammalian translational landscape is governed by a dense network of endogenous tRNAs and intricate mRNA structures, making de novo genome synthesis currently impractical for generating blank coding channels. This limitation has prompted a methodological transition from comprehensively rewriting the host genome to exploiting the inherent non-uniformity of natural codon usage through a strategy known as RCR (Fig. 4a).

Translating this concept into practice, RCR was first successfully implemented in mammalian cells by leveraging the natural scarcity of the TCG codon. Because TCG exhibits both the lowest genomic usage frequency and the lowest abundance of its cognate endogenous tRNA, it serves as an ideal candidate to achieve high-efficiency ncAA incorporation into a target POI alongside minimal background misincorporation across the proteome. The development of a systematic engineering framework and a big-data model, specifically the sequence context for TCG recoding (SCTR) model, allowed for the precise prediction and optimization of ncAA incorporation sites (Ding et al., 2024b). By screening mutually orthogonal aaRS/tRNA pairs and optimizing their expression levels, this approach achieved recoding efficiencies comparable to wild-type protein expression. Ultimately, this breakthrough demonstrated that even without massive genomic alterations, specific rare sense codons could be repurposed into highly selective and efficient vehicles for ncAA incorporation.

Building upon this foundational TCG recoding framework, recent advancements have pushed the technology toward the simultaneous incorporation of multiple distinct ncAAs. By systematically evaluating additional rare codons and engineering highly orthogonal translation machinery, it is now possible to incorporate up to five distinct ncAAs into a single protein within mammalian cells (Fig. 4b) (Fang et al., 2026). This multiplexed recoding strategy achieves high fidelity and site-specificity, enabling the creation of complex protein architectures with diverse chemical functionalities. This progress reveals the redefinable nature of the eukaryotic genetic code and provides a robust platform for engineering sophisticated biological tools.



(b) Genetic code table

Recoded with ncAAs Capable for ncAAs

UUU	Phe	UCU	Ser	UAU	Tyr	UGU	Cys
UUC		UCC		UAC		UGC	
UUA	Leu	UCA		UAA	Stop	UGA	Stop
UUG		UCG		UAG	Stop	UGG	Trp
CUU	Leu	CCU	Pro	CAU	His	CGU	Arg
CUC		CCC		CAC		CGC	
CUA		CCA		CAA	Gln	CGA	
CUG		CCG		CAG		CGG	
AUU	Ile	ACU	Thr	AAU	Asn	AGU	Ser
AUC		ACC		AAC		AGC	
AUA		ACA		AAA	Lys	AGA	Arg
AUG	Met	ACG		AAG		AGG	
GUU	Val	GCU	Ala	GAU	Asp	GGU	Gly
GUC		GCC		GAC		GGC	
GUA		GCA		GAA	Glu	GGA	
GUG		GCG		GAG		GGG	

Fig. 4 Strategies for expanding the genetic code via rare codon recoding (RCR). (a) Schematic of the RCR strategy for integrating ncAAs in mammalian cells. Within this framework, Rec-tRNA refers to the engineered tRNA for RCR. Sup-tRNA represents the tRNA used in conventional stop codon suppression. Dec-tRNA designates the native endogenous tRNA that decodes natural amino acids. In stop codon suppression, the decoding of aminoacylated Sup-tRNA is heavily impeded by the translation termination machinery (such as eRFs), limiting ncAA incorporation. Conversely, the RCR approach enables the aminoacylated Rec-tRNA to successfully outcompete the inherently weak endogenous Dec-tRNA, thereby facilitating highly efficient ncAA incorporation. The width of the lines visually indicates the intensity of the process flow. (b) Codon table schematic illustrating the expansion of available coding channels for multi-type ncAA incorporation. Experimentally recoded codons are highlighted in blue, and candidate codons with potential for future ncAA reassignment are highlighted in light green. ncAAs: non-canonical amino acids; tRNA: transfer RNA; eRFs: eukaryotic release factors; Rec-tRNA: recoding transfer RNA; Dec-tRNA: endogenous decoding transfer RNA; Sup-tRNA: stop-codon suppressor transfer RNA; Phe: phenylalanine; Leu: leucine; Ile: isoleucine; Met: methionine; Val: valine; Ser: serine; Pro: proline; Thr: threonine; Ala: alanine; Tyr: tyrosine; His: histidine; Gln: glutamine; Asn: asparagine; Lys: lysine; Asp: aspartate; Glu: glutamate; Cys: cysteine; Trp: tryptophan; Arg: arginine; Gly: glycine; mRNA: messenger RNA.

Ultimately, the successful implementation of such multiplexed sense codon reassignment in mammalian cells expands the scope of site-specific protein engineering. This technology facilitates the precise introduction of bioorthogonal handles, structural probes, or PTM mimics into selected POI. In future studies, these engineered systems may help probe defined protein states and interactions in carefully validated settings. As the repertoire of available rare codons expands, this platform may support the engineering of therapeutic proteins with defined chemical modifications and enable more targeted studies of selected disease-relevant protein functions.

4 Conclusions and outlook

In conclusion, sense codon reassignment leverages selected reassignment-permissive codons to expand control over protein composition and function. This is achieved by exploiting the redundancy of the genetic code: 61 sense codons encode 20 cAAs, and theoretical analyses suggest that roughly 30–40 codons may suffice to encode an organism's proteome, leaving over 20 codons as potential ncAA-reassignment targets (Krishnakumar et al., 2013; Budisa, 2025). By actively manipulating translational competition within the ribosomal decoding center, this paradigm has successfully evolved into two complementary technological routes. The first route harnesses endogenous competition for global proteome profiling, facilitating dynamic imaging and deep coverage of the nascent proteome under physiological conditions. The second route focuses on establishing dedicated decoding pathways for the precise, site-specific, and multiplexed incorporation of ncAAs into target proteins. Importantly, the efficiency and fidelity of both routes are not determined solely by competition at the ribosomal decoding center, but also by aaRS/tRNA orthogonality, endogenous tRNA abundance and modification status, aminoacylation kinetics, elongation factor compatibility, mRNA and codon-pair context, ncAA uptake, and cellular toxicity. In prokaryotes, coupling this strategy with genomic compression and total synthesis has generated dedicated blank codons, empowering the in vivo programmable synthesis of complex sequence-defined polymers and macrocycles. Meanwhile, in mammalian systems, rare sense codon recoding has enabled the incorporation of up to five distinct ncAAs into a single protein. In addition, future studies may leverage rare sense codon recoding to install defined chemical probes or PTM mimics into selected proteins, providing a useful platform for hypothesis-driven dissection and manipulation of protein function.

Despite these advancements, the broader implementation of this technology encounters several critical limitations. First, simultaneous incorporation of multiple ncAAs can lead to inter-system crosstalk and cross-reactions with the endogenous translation machinery, resulting in deacylation or misreading. Second, in mammalian systems where whole-genome synthesis remains currently unattainable, strategies that rely on competition with endogenous tRNAs inevitably produce background incorporation at the proteome-wide level. This noise can interfere with biological observations in sensitive cell lines or delicate in vivo microenvironments. Third, orthogonal components derived from prokaryotes or archaea may misfold within eukaryotic environments or display suboptimal substrate affinity. Furthermore, their prolonged expression risks inciting immunogenicity or cellular stress.

To resolve these constraints, future research must pivot toward several strategic breakthroughs. A primary imperative is the development of more multiplexed OTSs in mammalian cells. Through directed evolution and rational design, mutually orthogonal pairs could be constructed that do not crosstalk with each other or with the endogenous system, thereby supporting the coordinated encoding of multiple distinct ncAAs (Willis and Chin, 2018; Beattie et al., 2023; Fang et al., 2026). Concurrently, using CRISPR-mediated genome editing to ablate or replace endogenous tRNA genes corresponding to target rare codons will establish a pristine genetic background, effectively eradicating competitive recognition (Ding et al., 2024b). Furthermore, integrating machine learning algorithms with sequence-context models is essential to systematically decipher the determinants of recoding efficiency. While RCR efficiencies are generally robust, ranging from 40% to over 90%, they are profoundly dictated by the flanking nucleotide sequences (Ding et al., 2024b). Integrating structural biology, high-throughput assays, and deep learning will elucidate the molecular mechanisms governing this context dependency. This integration will drive the development of universal predictive tools capable of pre-evaluating and optimizing

flanking sequences for any target site, elevating recoding efficiency to its theoretical maximum while maintaining a low off-target background. Finally, the field must transition from global overexpression paradigms toward precise spatiotemporal and localized regulation (Koehler et al., 2020). Deploying chemically induced dimerization systems (Jiang et al., 2023) or optogenetic switches (Charette et al., 2025) can strictly limit the incorporation time window, while anchoring orthogonal translation components to specific subcellular compartments or transcripts can effectively confine ncAA incorporation to the desired spatial domains (Reinkemeier et al., 2019; Ding et al., 2024b).

Supported by the precise regulatory capabilities afforded by these technical advancements, sense codon reassignment presents a broad landscape of transformative applications. In basic life sciences, researchers could precisely introduce photo-crosslinking groups and bioorthogonal handles into distinct domains of a single protein, facilitating a sequential capture and labeling methodology to identify transient interacting partners. In translational medicine and next-generation biotherapeutic development, this technology provides a foundation for engineering advanced safety circuits in chimeric antigen receptor T (CAR-T) cells. By rendering anti-apoptotic proteins and metabolic enzymes strictly dependent on distinct ncAAs encoded by separate rare codons, cellular activation can be restricted to a precise logical AND-gate condition. Moreover, embedding rare codons into essential sites of viral structural proteins can generate highly safe vaccines that absolutely require dual ncAAs for packaging, mechanistically eliminating the risk of reversion mutations. Ultimately, sense codon reassignment has the potential to become a versatile tool for decoding the complex molecular logic of life to significantly advance the future landscape of synthetic biology and precision molecular medicine.

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Author contributions

Yiru ZHANG, Wei YU, and Wenlong DING contributed to writing the original draft and editing the manuscript. Wenlong DING and Shixian LIN conceived the review topic, developed its framework, and critically reviewed and edited the manuscript. All authors have read and approved the final manuscript.

Compliance with ethics guidelines

Yiru ZHANG, Wei YU, Wenlong DING, and Shixian LIN declare that they have no conflicts of interest.

This review does not contain any studies with human or animal subjects performed by any of the authors.

Declaration on the use of generative AI tools

During the preparation of this work, the authors used ChatGPT for language polishing, readability enhancement, and grammar checking. The generated output was thoroughly reviewed and edited by the authors to ensure accuracy, originality, and compliance with journal standards. The authors take full responsibility for the final content of this manuscript.

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