



## Review:

# A physical view of computational neurodynamics\*

Jun MA<sup>†1</sup>, Zhuo-qin YANG<sup>2</sup>, Li-jian YANG<sup>3</sup>, Jun TANG<sup>4</sup>

<sup>1</sup>Department of Physics, Lanzhou University of Technology, Lanzhou 730050, China

<sup>2</sup>School of Mathematics and Systems Science, Beihang University, Beijing 100191, China

<sup>3</sup>Department of Physics, Central China Normal University, Wuhan 430079, China

<sup>4</sup>School of Physics, China University of Mining and Technology, Xuzhou 221116, China

<sup>†</sup>E-mail: hyperchaos@163.com; hyperchaos@lut.cn

Received June 19, 2019; Revision accepted July 25, 2019; Crosschecked Aug. 6, 2019

**Abstract:** The nervous system is made of a large number of neurons. Time-varying balance between excitatory and inhibitory neurons is important to activate appropriate modes of electrical activity. A realistic biological neuron is complex, often presenting various electrophysiological activities and diffusive propagation of ions in the cell. Therefore, the physical effects of electromagnetic induction become very important and should be considered when estimating signal encoding and mode selection. Synaptic plasticity and anatomical structure have been developed to enhance the self-adaption of neurons. Thus, the electrical mode with the most effective links and weights can be selected to benefit information encoding and signal propagation between neurons in the network. As a result, the demand for metabolic energy can be greatly reduced. In this review, neuron model setting with biophysical effects, modulation of astrocytes, autapse formation and biological function, synaptic plasticity, memristive synapses, and field coupling between neurons and networks are reviewed briefly to provide guidance in the field of neurodynamics.

**Key words:** Neuron; Neural networks; Autapse; Hamilton energy; Electromagnetic induction  
<https://doi.org/10.1631/jzus.A1900273>

**CLC number:** O59; TN710

## 1 Building a neuron model

Electrophysiological processes in the cell can induce distinct bioelectricities and these charged ions are pumped to flow across the cell membrane via channels embedded in the membrane (Kawato et al., 1984; Stent, 1984; Busciglio et al., 1992). As a result, the balance of ion concentration between the inside and outside of the cell is disturbed and membrane potential becomes time-varying when an external electric stimulus is imposed on the neuron. From a physical viewpoint, static potassium and sodium can

activate an electric field and the field energy can be generated by spatial distribution in the cell. Furthermore, the diffusion and propagation of these charged ions will change the energy distribution and energy propagation. In particular, the continuous current across the membrane channels will change the density distribution of charged ions, energy storage and release. Therefore, capacitance for the cell membrane can be used to estimate the membrane potential, and signal propagation will be modulated (Holmes and Loew, 2008). Biological cells are elastic and their geometry alters to change the capacitance of membranes exposed to external electromagnetic fields due to the effect of polarization and magnetization. Most previous studies seldom considered the elastic properties of cell membranes (Valverde, 1976; Pellionisz, 1989; Tomba et al., 2014) and thus the capacitance was considered fixed as a constant in biological neuron models (Fitzhugh, 1966; Hassard, 1978;

\* Project supported by the National Natural Science Foundation of China (Nos. 11765011 and 11672122) and the Hongliu First-class Disciplines Development Program of Lanzhou University of Technology, China

ORCID: Jun MA, <https://orcid.org/0000-0002-6127-000X>

© Zhejiang University and Springer-Verlag GmbH Germany, part of Springer Nature 2019

Morris and Lecar, 1981; McCormick et al., 2007; Pospischil et al., 2008). These neuron models can be simplified and rebuilt in circuits for nonlinear analysis (Tsumoto et al., 2006; González-Miranda, 2007; Storace et al., 2008; Goldwyn et al., 2011; Mao, 2017). For example, Patel and DeWeerth (1997) designed a very-large-scale integration (VLSI) circuit for producing electrical activity in a Morris-Lecar neuron, which was verified in an analog circuit by Hu et al. (2016). In this paper, we analyze the reasons for tolerance fluctuations, construct a condition-driven adaptive design method for tolerance using a hidden Markov model algorithm, and then present the results of some experiments on the precision stamping process and discuss the feasibility and effectiveness of the adaptive design method.

As the basic functional unit in the nervous system, a reliable neuron model is important to estimate the dynamic properties and predicate possible mode transition in neural activities. The propagation and pumping of ions in the cell can be very complex, but the application of a patch clamp benefits the estimation and detection of the membrane potential of neurons. As a result, implicit and auxiliary variables can be used to build reliable neuron models. Astrocytes (Dani et al., 1992; Parpura et al., 1994; Zonta et al., 2003; Navarrete et al., 2014) play an important role in regulating the concentration of calcium and inositol triphosphate (IP<sub>3</sub>) by adjusting the neurotransmitters adenosine triphosphate (ATP) and glutamic acid. For some interneurons, an auxiliary loop is formed to develop an autapse (van der Loos and Glaser, 1972; Seung et al., 2000; Yue et al., 2017; Song et al., 2018). An autapse is a specific synapse, connecting its own body via a closed loop. It can be developed to enhance signal propagation when the axon of the neuron is injured (Wang CN et al., 2017). Thus, the anatomical structure of neurons and interaction between astrocytes and neurons should be considered. Guo et al. (2017) proposed that an astrocyte-neuron network driven by autapses can detect the possible occurrence of mode selection and firing pattern in the electrical activities. For isolated neurons, the involvement of an autapse connection can enhance the self-adaption of mode selection and response in electrical activities (Song et al., 2015; Ren et al., 2017; Uzun, 2017; Zhao and Gu, 2017; Xu Y et al., 2018a). Furthermore, setting an appropriate

distribution of autapse connections in the network can induce coherence resonance (Uzun et al., 2017; Yang et al., 2017) and enhance the realization of synchronization (Ma et al., 2015b) and pattern selection (Ma et al., 2015a) in the network.

More importantly, physical effects should be considered in building reliable neuron models. From a dynamic point of view, a variety of neural circuits (Tamaševičius et al., 2015; Wei et al., 2017; Carro-Pérez et al., 2018; Wang RB et al., 2018; Bao et al., 2019) can be designed to generate different firing patterns by adjusting the parameters or applying an appropriate external stimulus. Thus, spiking, bursting, and even chaotic series can be produced to match the nonlinear properties of neural activities. In physical and mechanical systems, continuous energy supply is critical to support stable oscillation. In biological systems, sufficient metabolic energy is needed to maintain normal electrical activities. An energy model for estimating the electrical activities of neurons has recently been proposed (Wang ZY et al., 2015; Zheng et al., 2016; Wang YH et al., 2017; Wang and Wang, 2018). Results from the model are important for estimating the relation between blood flow, energy supply, and mode transition. Inspired by the Helmholtz theorem (Kobe, 1986), scale transformation is often applied to neuronal models and nonlinear circuits. The Hamilton energy (Wang Y et al., 2017; Wu et al., 2018a; Zhang et al., 2018) can be calculated to estimate the dependence of energy on oscillation modes, chaos control, and nonlinearity in the system. For example, a neuron will maintain lower Hamilton energy in a bursting and/or chaotic state than in a spiking state. The occurrence of multi-scroll attractors also presents lower Hamilton energy in chaotic systems. Most neuron models emphasize the occurrence and fluctuation of membrane potential induced by the channel current and an external electric stimulus, while the intrinsic physical field effect is missed. Potassium, sodium, and calcium are known to be kept and transmitted in and out of the cell membrane. Any slight spatial change or flow of these charged ions will induce a complex electromagnetic field in the cell and thus the successive transmission and pumping of ions will be changed to modulate electrical activity.

Therefore, Ma and Tang (2015) and Wu et al. (2017) suggested that magnetic flux can be added as

new variable to existing neuron models. The law of electromagnetic induction indicates that an equivalent induction current can be imposed to approach the effect of an induced electromotive force, which shows certain weight modulation on the membrane potential. Indeed, a memristor can be magnetic flux-controlled or charge-controlled, and the memristive function is dependent on the relation between magnetic flux and charges. At least two variables (membrane potential and current) should be used to estimate the electrical activity of a neuron. Ma and Tang (2015) introduced magnetic flux as a new additive variable to the three-variable neuron model, and the same magnetic flux was introduced into the four-variable neuron model. As a result, the effect of electromagnetic induction is estimated by supplying additive current to the membrane. The ability of a two-variable neuron model to describe the field effect resulting from electromagnetic induction can be improved by using three variables. For building a generic and simple neuron model, the effect of electromagnetic induction and radiation on neural activity can be estimated as

$$\begin{cases} C \frac{du}{dt} = f(u, i, p) + I_{\text{ext}} + i_{\text{induct}}, \\ L \frac{di}{dt} = g(u, i, p), \\ \frac{d\varphi}{dt} = k_1 u - k_2 \varphi + \varphi_{\text{ext}}, \\ i_{\text{induct}} = \frac{dq}{dt} = \frac{dq}{d\varphi} \frac{d\varphi}{dt} = k_0 \rho(\varphi) u, \end{cases} \quad (1)$$

where  $u$  represents the membrane potential,  $i$  is the channel current,  $q$  is the charge,  $\varphi$  represents the magnetic flux,  $p$  is the intrinsic parameter, and  $k_0$ ,  $k_1$ , and  $k_2$  are parameters associated with the media. When a neuron is considered as an excitable media, its physical parameters can be approached by using equivalent capacitance  $C$  and inductance  $L$ .  $t$  denotes time.  $I_{\text{ext}}$  denotes an external stimulus, and  $i_{\text{induct}}$  the induction current resulting from electromagnetic induction.  $\varphi_{\text{ext}}$  is used to describe different types of electromagnetic radiations.  $\rho(\varphi)$  calculates the memductance of the memristor. This kind of electromagnetic induction can also be considered in the Hindmarsh-Rose, Hodgkin-Huxley (HH), and other

neuron models (Ma and Tang, 2015; Wu et al., 2017) by including the magnetic flux variable and memristive function. In particular, when this effect is estimated in cardiac tissue, which is often described by two-variable reaction-diffusion equations, two kinds of death mechanisms (Wu et al., 2016; Ma et al., 2017) of heart tissue from electromagnetic radiation can be explained by a breakup of spiral waves and blocking of the propagation of target waves.

$$\begin{cases} \frac{\partial u}{\partial t} = f(u, v) + D\nabla^2 u + i_{\text{induct}} + I_{\text{ext}}, \\ \frac{\partial v}{\partial t} = g(u, v), \\ \frac{\partial \varphi}{\partial t} = k_1 u - k_2 \varphi + \varphi_{\text{ext}}, \\ i_{\text{induct}} = \frac{dq}{dt} = \frac{dq}{d\varphi} \frac{d\varphi}{dt} = k_0 \rho(\varphi) u, \end{cases} \quad (2)$$

where  $\nabla^2$  is the Laplace operation, and  $D$  represents the diffusion coefficient. The variables  $u$  and  $v$  often describe the activator such as membrane potential, and inhibitor such as recovery variable for current, respectively. In the cardiac tissue of a healthy heart, the sinoatrial node can emit a continuous electrical signal and maintain a stable target wave (Wu et al., 2016; Ma et al., 2017). A higher intensity of electromagnetic radiation can block the propagation of a target wave, thereby suppressing the blood pump in the heart (Qu et al., 2014). On the other hand, in the case of arrhythmia and tachycardia, when some spiral waves can be detected in the cardiac tissue, electromagnetic radiation can induce breakup of spiral waves, and ventricular fibrillation is induced leading to final rapid death of the heart. Inspired by model setting for neural activity (Ma and Tang, 2015; Wu et al., 2017), extensive studies have been carried out to investigate the collective behavior of neural networks and wave propagation in cardiac tissue in the presence of electromagnetic induction and radiation (Mvogo et al., 2017; Zhan and Liu, 2017; Ge et al., 2018b; Rostami et al., 2018; Takembo et al., 2018; Xu Y et al., 2018b; Lv et al., 2019; Mostaghimi et al., 2019).

It is accepted that an induction current can be used to estimate the effect of electromagnetic induction resulting from a time-varying concentration of charged ions under transmission and exchange. Also,

by generating memristive currents it can give helpful clues to help understand the function of memristive synapses. Therefore, when two neurons are connected by a memristive synapse (Park et al., 2015; Covi et al., 2016; Azghadi et al., 2017; Xu F et al., 2018), the synapse current for the coupled neurons is estimated by

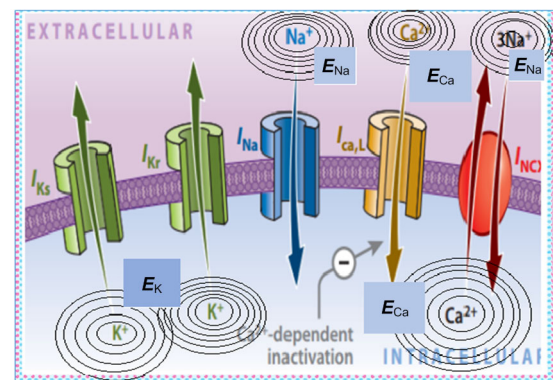
$$I_m = k\rho(\varphi)(u_1 - u_2), \quad (3)$$

where  $u_1$  and  $u_2$  describe the membrane potential for each of the two neurons respectively,  $\rho(\varphi)$  is dependent on the synapse property, and  $k$  is the coupling intensity. According to the difference in function mechanism, an electrical synapse is often considered as a gap junction coupling between neurons, while chemical synapse coupling is activated between neurons by the release of neurotransmitters. From a physical viewpoint, electrical synapse coupling could account for voltage coupling via a resistor, while chemical synapse coupling can be recognized as field coupling (Perea et al., 2009; Ma SY et al., 2019; Wu et al., 2019) because any release of neurotransmitter can induce a time-varying electromagnetic field and a change in the distribution of charged ions in the cell. The field variable is not presented in this model, though its field effect is considered by using an induction current. Therefore, a variable  $E$  is introduced to describe the effect of the electric field in the neuron, that is, the continuous exchange of ions across the membrane channels can be considered as placing a certain distribution of charges on the cell membrane. For simplicity, a two-variable nonlinear circuit is used and the intrinsic electric field  $E$  of the capacitor is estimated. As a result, its generic form and the relations of physical variables are estimated as follows (Ma J et al., 2019):

$$\begin{cases} C \frac{dV}{dt} = f(V, i, p), \\ L \frac{di}{dt} = g(V, i) + rE, \\ \frac{dE}{dt} = ki + E_{\text{ext}}, \end{cases} \quad (4)$$

where  $V$  and  $E$  describe the membrane potential and inner and outer electric fields of the cell membrane,

respectively.  $C$ ,  $L$ ,  $r$ ,  $k$ , and  $p$  are normalized parameters for equivalent capacitance, inductance, the size of the membrane and intrinsic properties of the media.  $E_{\text{ext}}$  is the external static or the time-varying electric field, and the propagation of an ion flow will be regulated to modulate the channel current. When more than two neurons are exposed to the external electric field, the electric field between the neurons will be activated to adjust the synapse coupling. As suggested in open problems (Ma SY et al., 2019), the Hindmarsh-Rose and HH neuron models can be used to describe the nonlinearity in Eq. (4). Thus, field coupling-induced synchronization and pattern formation can be further investigated. From physical and chemical viewpoints, the neuron cell can be regarded as an excitable media in which the inner propagation of charged ions and external electromagnetic field will change the distribution of magnetic and electrical fields, as shown in Fig. 1. As a result, the transient distribution and flow of ions across the membrane will have a distinct impact on membrane potential. Therefore, it is useful to include appropriate variables to estimate the membrane potential, channel current, magnetic field, and electrical field. Wu et al. (2019) proposed a new neuron model with four physical variables: membrane voltage, current, charge number on the membrane, and magnetic flux. Their paper includes a detailed description of the dynamical response induced by electromagnetic radiation.



**Fig. 1 Ion propagation in a neuron cell. Calcium, potassium, and sodium ions can trigger spatial distribution of electric fields  $E_{Ca}$ ,  $E_K$ , and  $E_{Na}$**

Continuous movement and propagation of ions can induce a time-varying magnetic field which can be described by magnetic flux

## 2 Contribution of astrocytes

Astrocytes were thought to be passive elements playing merely nutritional and structural roles in the central nervous system of mammals. However, new evidence suggests that they can have a great effect on neuron function (Perea et al., 2009). It was suggested that astrocytes implement a feedback control to neural activity through synapses by regulating neurotransmitter release (Volterra and Meldolesi, 2005; Wang et al., 2009; Halassa and Haydon, 2010; Henneberger et al., 2010; Khakh and Sofroniew, 2015; Poskanzer and Yuste, 2016). This feedback may involve different biochemical pathways (de Pittà et al., 2012), among which, the astrocytic calcium modulation pathway is most involved. Astrocytes are not electrically excitable cells, but they can sense and respond to synaptic activity by adjusting their  $Ca^{2+}$  concentration. When the neurotransmitter glutamate is released from a presynaptic neuron, the astrocyte  $Ca^{2+}$  concentration will be increased. Furthermore, astrocytes can release some active gliotransmitters which can modulate the excitability and synaptic plasticity of pre- and postsynaptic neurons (Araque et al., 2014). Indeed, astrocytes are connected via gap junctions, and any increase of  $Ca^{2+}$  concentration in one astrocyte may induce a  $Ca^{2+}$  wave in nearby astrocytes (Newman and Zahs, 1997). As a result, neurons connected to those astrocytes will be excited and may have epileptic activity in the whole neuron-astrocyte network.

Due to the limitations of experimental technology, most studies on astrocyte  $Ca^{2+}$  signaling are performed in vitro (Bezprozvanny et al., 1991; Höfer et al., 2002; Sloan and Barres, 2014; Manninen et al., 2018). Thus, the reliability of these results may be dependent on the method applied and the context selection. Therefore, a reliable computational method is often required to understand the complexity of different astrocyte  $Ca^{2+}$  signals and estimate the dynamics of astrocytic  $Ca^{2+}$ . The most popular astrocyte  $Ca^{2+}$  models are the DeYoung-Keizer (de Young and Keizer, 1992) and Li-Rinzel (Li and Rinzel, 1994) models, and the Höfer model (Höfer et al., 2002), which includes a calcium ( $Ca^{2+}$ )-induced  $Ca^{2+}$  release (CICR) mechanism (Höfer et al., 2002). The Höfer model was proposed specifically to simulate astrocytes, and comprises the dynamics of the cytoplasmic  $Ca^{2+}$  concentration ( $C_{Ca^{2+}}$ ), endoplasmic reticulum

(ER) store  $Ca^{2+}$  concentration ( $C_{Ca^{2+},ER}$ ),  $IP_3$  concentration ( $C_I$ ), and active fraction of  $IP_3R$  ( $C_R$ ). The nonlinear relation between these variables can be calculated by

$$\left\{ \begin{aligned} \frac{\partial C_{Ca^{2+}}}{\partial t} &= v_{rel} - v_{SERCA} + v_{in} - v_{out} \\ &\quad + D_{Ca} \left( \frac{\partial^2 C_{Ca^{2+}}}{\partial x^2} + \frac{\partial^2 C_{Ca^{2+}}}{\partial y^2} \right), \\ \frac{\partial C_{Ca^{2+},ER}}{\partial t} &= \beta(v_{SERCA} - v_{rel}), \\ \frac{\partial C_R}{\partial t} &= v_{rec} - v_{inact}, \\ \frac{\partial C_I}{\partial t} &= v_{PLC\beta} + v_{PLC\delta} - v_{deg} \\ &\quad + D_{IP_3} \left( \frac{\partial^2 C_I}{\partial x^2} + \frac{\partial^2 C_I}{\partial y^2} \right), \end{aligned} \right. \quad (5)$$

where  $v_{rel}$ ,  $v_{SERCA}$ ,  $v_{in}$ , and  $v_{out}$  represent the  $Ca^{2+}$  release from the ER store,  $Ca^{2+}$  pump into the ER store,  $Ca^{2+}$  transmembrane influx, and transmembrane outflow, respectively.  $v_{rec}$  and  $v_{inact}$  are the recovery and inactivate rates, respectively, of the  $IP_3$  receptor.  $v_{PLC\beta}$  and  $v_{PLC\delta}$  are the activation rates of  $IP_3$  mediated by  $PLC\beta$  and  $PLC\delta$ , respectively, and  $v_{deg}$  is the  $IP_3$  degradation rate. The diffusion of  $Ca^{2+}$  and  $IP_3$  inside the astrocyte cell is described by the last diffusive terms, where  $D_{Ca}$  and  $D_{IP_3}$  are the diffusion coefficients. The Höfer model assumes that  $IP_3$  is the key messenger that mediates information communication between cells. Based on this model, many studies of  $Ca^{2+}$  signaling in astrocytes (Lavrentovich and Hemkin, 2008; Zeng et al., 2009; Toivari et al., 2011) have been discussed. The DeYoung-Keizer and Li-Rinzel models, in which the neurotransmitter is taken into account at the same time (Gibson et al., 2007; Bennett et al., 2008; Chander and Chakravarthy, 2012; Witthoft and Karniadakis, 2012; Hadfield et al., 2013; Witthoft et al., 2013; Kenny et al., 2018), have also been selected for detecting signal propagation in and between cells.

A pioneering model named the ‘dressed neuron’ model, which includes a neuron and an astrocyte, was proposed by Nadkarni and Jung (2003). They adapted

the HH model (Hodgkin and Huxley, 1952) to simulate the action of a neuron, and the Li-Rinzel model to simulate the  $\text{Ca}^{2+}$  dynamics in an astrocyte. When the neuron is triggered to produce an action potential, neurotransmitter is released into the synaptic cleft and binds to the transmitter receptor on the astrocyte. Thus, the intracellular  $\text{IP}_3$  is released as follows (de Young and Keizer, 1992; de Pittà et al., 2012):

$$\frac{dC_{\text{IP}_3}}{dt} = \frac{1}{\tau_{\text{IP}_3}} (C_{\text{IP}_3}^* - C_{\text{IP}_3}) + r_{\text{IP}_3} \Theta(v - 50 \text{ mV}), \quad (6)$$

where  $C_{\text{IP}_3}$  represents the concentration of  $\text{IP}_3$  in the astrocyte cell, and  $C_{\text{IP}_3}^*$  is the equilibrium value of  $\text{IP}_3$ .  $\tau_{\text{IP}_3}$  determines the time scale of chemical transmission of  $\text{IP}_3$ . A step function  $\Theta(x)$  is used to simulate the  $\text{IP}_3$  release triggered by action potential, and  $r_{\text{IP}_3}$  represents the strength of coupling between the neuron and the astrocyte. The feedback of astrocytes to the HH neuron is estimated by introducing the current  $I_{\text{astro}}$ , which can be obtained by fitting the experimental data (de Young and Keizer, 1992; de Pittà et al., 2012):

$$I_{\text{astro}} = 2.11 \Theta(\ln y) \ln y, \quad (7)$$

$$y = (C_{\text{Ca}^{2+}} - 196.69) \text{ nmol/L}.$$

Note that the time scale of the action potential of the neuron is millisecond, whereas the  $\text{Ca}^{2+}$  dynamics simulated by the Li-Rinzel model is on a much slower time scale of seconds. Some results confirmed that with a stronger coupling strength, i.e. a high value of  $r_{\text{IP}_3}$  that is related to the density of mGlu receptors on the astrocyte membrane, seizure-like oscillations emerge without an external stimulus (Nadkarni and Jung, 2003). Therefore, Nadkarni and Jung (2003) suggested that higher expression of mGlu receptors may be one of the physiological reasons for epilepsy. Based on a similar modeling scheme, Tang et al. (2013, 2016) obtained a tripartite synapse model and discussed in detail the information transmission between neurons. They found that the presence of an astrocyte facilitates the occurrence of episodic spikes (ESs) in both presynaptic and postsynaptic neurons.

Furthermore, the noise, originating from random open-close transitions of calcium ion channels in the endoplasmic reticulum membrane of the astrocyte, can change the firing patterns of two neurons and facilitate the occurrence of ESs in both neurons during neuronal information transmission.

In addition to the HH model, other neuron models, such as the Pinsky-Rinzel Model (Nadkarni and Jung, 2007), FitzHugh-Nagumo model (Postnov et al., 2007), and leaky integrate-and-fire model (Wade et al., 2012; Nazari et al., 2017), can be effective for examining information transmission between neurons and astrocytes. These models can reproduce the phenomena of hyperexcitability, plasticity, and  $\text{Ca}^{2+}$  oscillation observed in experiments. Other evidence has indicated that astrocytes can be organized into networks (Halassa and Haydon, 2010), interconnected through gap junction channels. These are regulated by extracellular and intracellular signals that enable the effective exchange of information. For exploring two networks, the concept of 'astroglial networks' was suggested in a recent review paper (Giaume et al., 2010). Network models including multiple neurons and astrocytes have been developed to study information transmission in the cortex (Allegrini et al., 2009; Liu and Li, 2013; Chan et al., 2017; Tang et al., 2017), hippocampus (Amiri et al., 2012a, 2013; Mesiti et al., 2015), and other parterres in the brain (Amiri et al., 2012b, 2012c; Yang and Yeo, 2015). The main issues discussed in those models are synchronization, information transfer, and hyperexcitability. For example, Amiri et al. (2012a, 2013) constructed a neural network model to study the effect of astrocytes on synchronization reach. It is thought that astrocytes are capable of changing the threshold value of transition from synchronous to asynchronous behavior among neurons. Tang et al. (2017) constructed a chain-type neuron-astrocyte network model to study the correlation between an astrocytic calcium wave and seizure-like behavior in a neuron network. They concluded that calcium wave propagation in astrocytes dominates the propagation of seizure-like discharges (SDs) in coupled neurons (Tang et al., 2017). As reviewed by Manninen et al. (2018), calcium signaling models in astrocytes can be categorized into four groups: isolated astrocyte models, astrocyte network models, neuron-astrocyte synapse models, and neuron-astrocyte network models.

### 3 Synaptic plasticity

Elastic media, such as cardiac tissue and muscle, can capture and slow down external mechanical pressure by inducing an appropriate deformation orientation. Complex electrophysiological activity occurs in the cardiac tissue and nervous system, and an appropriate response will be triggered when an external electric stimulus is applied. Synapses behave synchronously as receptors and transmitting terminals. In particular, the input signal will be encoded by a synapse and then converted to an equivalent transmembrane potential. Furthermore, intracellular and extracellular ion exchange will be regulated to induce a variety of firing patterns. External stimuli, including a signal from the post-synapse from an adjacent neuron, can change the polarization properties of a synapse. As a result, the impedance of the synapse will be changed under an external stimulus. When two or more neurons are coupled via synapses, the time-varying exchange of charged ions and fluctuation of membrane potentials can propagate electric signal along the axon to emit changeable signals to the pre-synapse of another neuron. Therefore, the impedance of the coupling synapse is changed to present different intensities. The nervous system has distinct self-adaption due to synaptic plasticity (Zucker, 1989; Bliss and Collingridge, 1993; Abraham and Bear, 1996; Abbott and Nelson, 2000; Zucker and Regehr, 2002). The connection intensity of synapses can be changeable, and the property or phenomenon of relatively permanent alteration in the morphology and function of synapses is confirmed. That is, the synapse current can be enhanced by an increase in involvement in the processing electrical activities, while its intensity is reduced by a decrease in involvement in signal encoding.

Synaptic plasticity can be short-term or long-term. Potentiation, depression, and facilitation are the main aspects of short-term synaptic plasticity. Short-duration synaptic plasticity (Salin et al., 1996; Buonanno, 2000; Junge et al., 2004; Pan and Zucker, 2009; Tutkun et al., 2010) is an important form of synaptic plasticity and plays an important role in activating the normal function of nervous systems. Synaptic short-duration plasticity can strengthen the certainty of synaptic transmission, and regulate the balance between the cortical excitation and inhibition.

As a result, a switch between excitation and inhibition enables neurons to select the most suitable response and firing patterns. In a word, its biological function is to form the temporal and spatial characteristics of neural activities, and to enhance and regulate the synchronous oscillation of the cortical thalamic network. Synaptic short-duration plasticity is involved in the realization of higher nervous system functions such as attention, priming effect, sleep rhythm, learning, and memory. Short-duration synaptic plasticity can be divided into short-duration enhancement and short-duration inhibition. Long-term synaptic plasticity (Bear and Malenka, 1994; Engert and Bonhoeffer, 1999; Nestler, 2001; Yuste and Bonhoeffer, 2001; Trachtenberg et al., 2002) is characterized by long-term potentiation (LTP) and long-term depression (LTD). Long-term potentiation is also called the long-term gain effect, and is a persistent potentiation phenomenon in signal transmission resulting from synchronous stimulation of two neurons. This is one of several phenomena associated with synaptic plasticity, the ability of synapses to change strength (Bliss and Collingridge, 1993). Since memory is thought to be encoded by changes in synaptic strength, LTP is widely regarded as one of the major molecular mechanisms underlying learning and memory.

In 1966, LTP was discovered in the hippocampus of rabbits by Terje Lomo and has long been a hot topic of research. Many modern LTP studies attempt to better understand its biological rationale, while others aim to explore the causal relationship between LTP and behavioral learning. Some researchers are developing ways to improve learning and memory by enhancing LTP, for example, by injecting drugs. LTP is also the subject of clinical research in areas such as Alzheimer's disease and addiction medicine. LTP has several characteristics, including input specificity, relevance, synergy, and persistence. LTD is known as long-term depression and long-term depotentiation, which refers to the inhibitory behavior of nerve synapses lasting for several hours to several days. Strong synaptic stimulation (cerebellar Purkinje cells) or long-term weak synaptic stimulation (hippocampus) can lead to long-term inhibition, which is thought to be induced by changes in postsynaptic receptor density. However, changes in presynaptic release may also have an effect on inhibition. Long-term inhibition

of the cerebellum is assumed to play an important role in motor learning, and long-term suppression of the hippocampus can be effective in erasing past memories. LTD of the hippocampus/cerebral cortex is controlled by the N-methyl-D-aspartate (NMDA) receptor, mGluR, or the endocannabinoid.

The coupling intensity and equivalent synapse current are dependent on the involvement of synapses such that external input-induced polarization can be balanced and less energy can be consumed. As a result, the synchronization between neurons and model selection can be controlled completely. For example, Wang JY et al. (2018) constructed a modular neuronal network with modified Oja's learning rule, and used it to eliminate the pathological synchronized rhythm of interacting bursting neurons. They found that synaptic plasticity with a high learning rate can effectively suppress bursting synchronization among strongly synchronous neurons in a modular neural network by applying a specific range of coupling intensity. Based on an Izhikevich neuron within a subthreshold excitatory population, these individual neurons can exhibit noise-induced bursts with increasing coupling intensity. The neuronal population has adaptive dynamic synaptic strength governed by spike-timing-dependent plasticity (STDP), and the neuron cannot fire spontaneously without noise. Kim and Lim (2018) investigated the effect of additive STDP on stochastic burst synchronization (SBS) by changing the noise intensity in a Barabási–Albert scale-free network (SFN). They explained a Matthew effect in synaptic plasticity which occurs due to a positive feedback process. Furthermore, perfect burst synchronization (with a high bursting measure) improves with LTP of synaptic strength, while non-perfect burst synchronization (with a low bursting measure) deteriorates with LTD. Tarai et al. (2019) discussed the neurobiological mechanisms of stress and mood disorders with the aim of enhancing the pharmacological effects of antidepressants and mood stabilizers. They found that regulation of neurotrophic factors can blockade stress and enhance neuronal survival, even though limbic regions can be paralyzed. Neurotrophic factors and molecular agents also adjust behavioral and synaptic plasticity in addiction and stress disorders. Short-term synaptic depression mainly reveals the depletion of the readily releasable pool (RRP) of quanta. Bui and Glavinović (2013) used patterned stimulation on the

Schaeffer collateral fiber pathway and model-fitting of the excitatory postsynaptic currents (EPSCs) recorded from CA1 neurons in rat hippocampal slices. Ursino et al. (2018) implemented new synaptic learning rules to take into account the role of partially shared features and distinctive features with different saliency. The trained network handled word recognition and task naming tasks in an effective way, and the different roles of salient versus marginal features in concept identification were exploited. Li (2014) studied dendritic and synaptic integration with different spatial distributions of synapses on the dendrites of a biophysically-detailed layer 5 pyramidal neuron model. They found that temporally synchronous and spatially clustered synaptic inputs make dendrites perform a highly nonlinear integration. Lu et al. (2019) analyzed the propagation and fidelity of a subthreshold EPSC signal in a feed-forward neural network composed of five layers by using the spike timing precision and power norm and the EPSC signal imposed on the Hodgkin–Huxley neurons of the first layer. They found that background noise contributed to the propagation of subthreshold EPSC signal in the feed-forward neural network and the fidelity between the system's response and subthreshold EPSC signal was preserved. Sun et al. (2019) discussed the dependence of signal detection on coupling strength and network topologies in small-world neuronal networks. They confirmed that the shorter the average path length, the better the signal detection under intermediate coupling strengths.

Synaptic plasticity, involving changes in synaptic strength observed *in vivo* or *in vitro* after learning, is one of the mechanisms underlying memory storage. Long-lasting forms of synaptic plasticity, including both LTP that synaptic strength increases (Bliss and Gardner-Medwin, 1973; Bliss and Lømo, 1973), and LTD that synaptic strength decreases (Ito, 1989), are the cellular bases of learning and memory (Bliss and Collingridge, 1993), which are fundamental mental processes critical for adaptation and survival.

Quantitative computational models have become important for obtaining a deep understanding of complex networks of interacting pathways with convergence, divergence, and positive and negative feedback loops. Some evidence has revealed that a large number of molecules, and complex interactions between them, underlie plasticity (Collins et al., 2005;



Coba et al., 2009). Presynaptic release of glutamate and postsynaptic depolarization are the two crucial features of most induction protocols at excitatory synapses. Biophysical models involving both electrophysiological properties and biochemical reactions (signaling pathways) have been developed to understand the pre- and postsynaptic events in LTP and LTD (Kotaleski and Blackwell, 2010; Manninen et al., 2010). For example, a framework for computational models of signaling pathways was proposed to understand the molecular mechanisms underlying synaptic plasticity of glutamatergic synapses (Kotaleski and Blackwell, 2010). Some computational postsynaptic signal transduction models have been developed to investigate the dependence of synaptic plasticity on species and interactions (Manninen et al., 2010).

An elevation in intracellular calcium in the postsynaptic neuron is crucial for LTP or LTD (Bliss and Collingridge, 1993; Malenka and Bear, 2004), and shows some differences from the molecular mechanisms leading to synaptic plasticity.  $\text{Ca}^{2+}$  can activate protein kinases and phosphatases for inducing phosphorylation–dephosphorylation cycles, LTP and LTD. There are various pre- and postsynaptic mechanisms of changes in synaptic strength, in which cytosolic  $\text{Ca}^{2+}$ /calmodulin-dependent signals play an important role in synaptic plasticity (Lisman and Goldring, 1988a, 1988b). Most mechanistic models typically confirm the role of calcium in synaptic plasticity by detecting calcium dynamics in electrical activity (Schiegg et al., 1985; Gamble and Koch, 1987; Holmes and Levy, 1990). It is claimed that the amplitude of calcium elevation depends on the frequency of synaptic stimulation, while kinases are not directly implicated in plasticity. It remains open why some stimulation protocols produce depression and others produce potentiation, because  $\text{Ca}^{2+}$  can activate multiple processes and enzymes. The interaction of multiple signaling pathways at multiple points leading to kinase activation, as well as neuromodulators, is nonlinear processing, which makes a quantitative understanding more difficult. Indeed, further computational modeling of signaling pathways is needed to investigate these complex interactions and predict important molecular mechanisms, and then to guide researchers to the most valuable experiments.

It is assumed that the number of molecules in deterministic modeling is large enough to be repre-

sented as a concentration. However, many subcellular compartments are so small as to contain finite molecules, which results in stochastic fluctuation in molecule numbers and changes the outcome of signaling pathways. Stochastic simulations have revealed that signaling pathways, such as positive feedback loops with bistable switches modeled deterministically, are no longer bistable. For example, taking an extremely long time to spontaneously switch states resembles the bistability mechanism, and suggests that synapses can exhibit multiple stable states (Hayer and Bhalla, 2005). These spontaneous transitions cause thresholds to be located in some ranges and so switches become either less sensitive to signals or more sensitive to noise (Bhalla, 2004; Hayer and Bhalla, 2005). Deterministic simulations (Bhalla and Iyengar, 1999; Bhalla, 2002; Ajay and Bhalla, 2004) have revealed several emergent properties of a global network of interacting pathways that are not present in individual pathways. Consequently, stochastic effects should be considered when modeling a system.

Spatial gradients of signaling molecules are known to be prominent in neurons with elongated dendritic structures, and the spatial aspect of cell signaling should be estimated. For example, synaptic inputs in one part of the dendrite induce the generation and diffusion of secondary messengers to other parts of the dendrite (Blackwell and Jędrzejewska-Szmek, 2013). When all synapses of a neuron are potentiated in response to synaptic stimulation, the neuron will respond not only to previously learned patterns, but also to any arbitrary spatial pattern of synaptic input from environmental stimuli (Irvine et al., 1994). Modeling this spatial aspect of neurons is a relatively new approach to investigating neuronal plasticity (Ajay and Bhalla, 2007; Neves et al., 2008). Although there are many non-spatial models of signaling pathways, it is necessary to focus on models incorporating significant morphological features of neurons, such as a soma with an elongated dendrite or a dendrite with spines (Blackwell and Jędrzejewska-Szmek, 2013). A more comprehensive, deterministic spatial model of signaling pathways (including mitogen-activated protein kinase (MAPK), protein kinase A (PKA), calcium/calmodulin-dependent protein kinase II (CaMKII), and protein kinase C (PKC)) needs to incorporate a multi-compartmental, multi-channel electrical model of a dendrite.

In summary, computational models of synaptic plasticity have taken into account the involvement of molecules in synaptic plasticity and that some particular molecular mechanisms are responsible for experimental observations. Most models at the molecular level are far from complete due to a lack of knowledge of the biology and exact setting for kinetic parameters. This also raises some challenges for understanding the role of the stochasticity induced by sometimes only dozens of copies of a certain protein in a synaptic spine, and how to estimate the effect of spatial inhomogeneity remains an open question. The scarcity of complete models of synaptic plasticity reflects the complexity of the underlying mechanisms resulting from insufficient information on the quantity and subcellular localization of critical enzymes. Therefore, applying a synthetic and integrative systems-level approach to model setting facilitates a deeper understanding of nonlinear processes of multiple interactions in information processing and memory storage in neurons.

#### 4 Collective behavior in neural networks

Due to the application of nanotechnology, feasible micro circuits and artificial synapses can be designed to build intelligent neuron processors. In a practical sense, the collective responses of neural processors and artificial neurons are worthy of further investigation so that signal propagation and information encoding in nervous systems can be understood. In this way, the occurrence and emergence of neural disease could be predicted, enabling possible suppression or curing. Neurons can be soaked in potassium, sodium, calcium, and even chloride ion solutions. A concentration gradient of ions can be activated to enhance the exchange and pumping of charged ions, thereby changing the membrane potential to trigger a variety of firing modes (Gu and Chen, 2014; Gu et al., 2014a, 2014b; Gu and Pan, 2015a, 2015b). In the nervous system, 20% of neurons can be inhibitory while 80% are kept in excitatory states. Therefore, it is important to consider the balance between excitability and inhibition (Zhao and Gu, 2015; Xiao et al., 2016) of neurons in estimating collective responses and pattern selection in neural networks. The collective behavior of networks de-

pends mainly on the local kinetics of node and topological connection (Ma et al., 2016b; Mei et al., 2016, 2018; Xu et al., 2016; Wei et al., 2018), and even initial setting (Ma et al., 2016a). For example, synchronization transition between neurons can be induced by resetting parameters and initial values (Gu et al., 2015). Multistability emerges in memristive systems involved with memristor-based functions. Therefore, synchronization stability between memristive systems (oscillators) is dependent on the initial values (Wu et al., 2018b, 2018c; Liu Y et al., 2019). As a result, resetting the initial values will induce different types of synchronization.

In biological neurons, chemical and electrical synapses are activated to receive and encode signal inputs and wave propagation between neurons. An electric synapse can couple neurons via a gap junction while a chemical synapse can connect neurons by release of a neurotransmitter. From a dynamic viewpoint, the parameters of many nonlinear circuits or systems can be modulated to trigger quiescent, spiking, bursting, and even chaotic series, which can be consistent with modes of electrical activity in the membrane potential of biological neurons. Any electrical devices such as resistors, capacitors, induction coils, and memristors can be used to connect the nonlinear circuits, and appropriate setting of parameters (resistance, capacitance, inductance, and memductance) for the coupling device can realize phase synchronization and/or complete synchronization. Indeed, gap junction coupling shows properties similar to those of direct voltage coupling via a resistor by consuming a certain Joule heat. This kind of coupling is often used as direct variable coupling. As explained by Liu ZL et al. (2019a) and Xu et al. (2019), capacitor coupling provides effective electric field coupling by balancing and pumping the energy from the connected circuits. The energy pumping in the coupling device is estimated from  $H=0.5C(V_1-V_2)^2$ , where  $C$  is the capacitance of the coupling device, and  $V_1$  and  $V_2$  are the output voltages of the circuits. On the other hand, inductor coupling (Yao et al., 2019) bridges magnetic field coupling by balancing the energy estimated as  $H=0.5LI^2$ , where  $L$  is the inductance and  $I$  the current across the coupling induction coil. A biological tissue such as a synapse can present complex physical properties such as resistance, capacitance, and inductance. Ma J et al. (2019) used a resistor, capacitor,

and induction coil to investigate synchronization realization between memristive circuits. Therefore, hybrid synapses can be more reliable in processing information and signal propagation than a sole chemical or electrical synapse. Liu ZL et al. (2019b) combined a capacitor, resistor, and induction coil to design hybrid coupling devices. By connecting these electrical devices in parallel or series, they found that parallel connection was more effective in stabilizing synchronization than connection in series, and that the intensity threshold via resistance coupling could be reduced and power consumption greatly reduced in the coupling device. For example, Fig. 2 is a diagram representing two artificial neurons connected via a hybrid synapse.

These hybrid synapses bridge the same output ends by balancing the energy flow and energy consumption. Nonlinear circuits can draw out many output ends, and thus more than two coupling channels can be opened for signal and energy exchange. For an isolated neuron, low frequency, high frequency signals and even different kinds of noise can be imposed synchronously, and the intrinsic properties of excitable media account for mode selection and dynamical response in the electrical activities. From a physical viewpoint, continuous or intermittent release of a neurotransmitter can change the propagation and distribution of charged ions. As a result, an electromagnetic field is induced to propagate signals between neurons. Therefore, this kind of field coupling gives physical evidence for understanding the biological function of chemical synapses. In realistic and biological nervous systems, neurons show diversity in excitability and inhibition, and can be considered in different clusters and layers in the networks. As a result, the collective response in hybrid networks with cluster connections and a layered distribution is

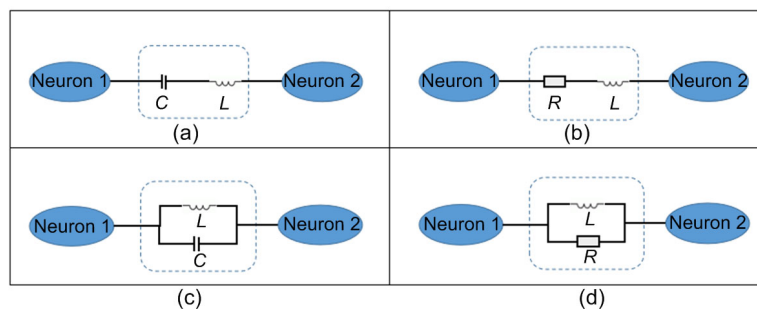
worthy of investigation by activating multiple channel coupling and hybrid synapses. In estimating the degree of spatial regularity and synchronization, the statistical synchronization factor (Qin et al., 2014; Wang and Ma, 2018)  $R$  is calculated using mean field theory, as follows:

$$F = \frac{1}{N} \sum_{i=1}^N x_i, \quad R = \frac{\langle F^2 \rangle - \langle F \rangle^2}{\frac{1}{N} \sum_{i=1}^N (\langle x_i^2 \rangle - \langle x_i \rangle^2)}, \quad (8)$$

where  $x_i$  is any detectable variable of node  $i$  in a network composed of  $N$  nodes, and  $\langle * \rangle$  represents the average value over time with a transient period.  $R \approx 1$  indicates perfect synchronization and the network will show a homogeneous distribution, while  $R \approx 0$  indicates the occurrence of non-perfect synchronization and a regular spatial pattern is formed in the network. In previous studies with the preferred chemical synapse between neurons, the coupling effect was estimated by adding an equivalent forcing current to each neuron. In our view, any winding or facing between synapses of two neurons can be approached by estimating the field coupling effect. Firstly, the gap junction can be thought of as an equivalent capacitor coupling in which a time-varying electric field is induced before reaching complete synchronization in the series for membrane potential. The interaction between synapse ends can be estimated by the induction current as follows:

$$i_c = \pm C' \left( \frac{dv_1}{dt} - \frac{dv_2}{dt} \right), \quad (9)$$

where  $v_1$  and  $v_2$  are the membrane potentials of two neurons (output voltages from the same output end of



**Fig. 2 Neuron connections bridged via hybrid synapses: (a) capacitor connecting inductor in series; (b) resistor connecting inductor in series; (c) capacitor connecting inductor in parallel; (d) resistor connecting inductor in parallel**

the neural circuits), and  $C'$  is the equivalent capacitance for the coupling synapse (or gap junction). The symbols “+” and “-” denote the current term included in the two neurons. As a result, a time-varying electric field is generated in this gap junction and energy is pumped between the two neurons. Considering the random flow and exchange of ions in the cell, a magnetic field also can be generated due to the flow of ions, and the neuron cell has a certain inductance. In this way, a magnetic field coupling can be considered when synapses are twisted together, then an induced electromotive force and induction current will be generated to balance the two neurons. As a result, the induced electromotive force  $\varepsilon$  modulates the membrane potential by imposing the induction current as follows:

$$i_L = \pm \frac{1}{L} \int (v_1 - v_2) dt, \quad (10)$$

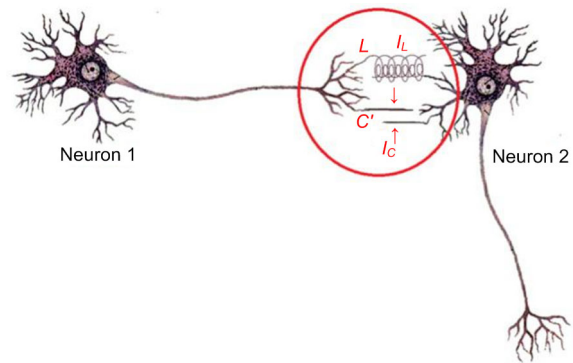
where  $L$  is the equivalent inductance for the coupling synapse, and  $v_1$  and  $v_2$  denote the membrane potentials of the neurons (also the output voltages from the output ends of the neural circuits). According to Eqs. (9) and (10), the capacitance and inductance of the coupling terms enables the ability to pump energy between neurons, and thus the membrane potential to be regulated for reaching synchronization. Furthermore, when the interaction between synapses is considered as field coupling, the winding and twisting between synapses can be explained as building an effective energy harvester, which can capture field energy from external electromagnetic radiation. For example, when the neurons (or neural circuits) are exposed to electromagnetic radiation, the coupling devices or coupled synapses can capture energy as follows:

$$\begin{cases} H_C = 0.5C'(v_1 - v_2)^2 + c_1 H_{\text{ext}}, \\ H_L = 0.5Li_L^2 + c_2 H_{\text{ext}}, \end{cases} \quad (11)$$

where  $H_C$  and  $H_L$  represent the energy in the coupling capacitor and induction coil, respectively.  $H_{\text{ext}}$  represents the energy flow from electromagnetic radiation, and  $c_1$  and  $c_2$  are coefficients for energy absorption. These coefficients are associated with the intrinsic properties of the media and neurons. A schematic

diagram of this kind of field coupling between neurons is shown in Fig. 3.

From a physical viewpoint, each neuron of a multi-layer network is embedded into a certain field node superposed by other neurons, and continuous flow between intracellular and extracellular ions will induce coexistence of a magnetic field and an electric field. Therefore, it is a challenge to discuss the collective behavior and regularity of a network when magnetic and electric field couplings between neurons are activated with different ratios. Recent studies of signal processing and communication between neurons suggest that synchronization stability and transition become more attractive when synaptic plasticity and memristive synapse function are considered (Zhang and Liao, 2017; Xu Q et al., 2018; Sharma et al., 2019; Zayer et al., 2019).



**Fig. 3 Magnetic field coupling and electric field coupling between neurons whose synapses are twisted together and/or placed in parallel**

## 5 Open problems

It is well known that the nervous system has a large number and diversity of neurons and these neurons can have different biological functions. However, most neuron models consider only a single biological function and the external stimulus is often handled as an equivalent current. Neural activities are often described by a series of membrane potentials and the anatomical structure and functional connectivity can be estimated by supplying appropriate additive current to the membrane potential. From a physical viewpoint, the electromagnetic field is often described by the electric field and magnetic field,

while many neuron models seldom involve the electromagnetic field variables directly. Therefore, it is difficult to estimate the electrical activities of neurons from a physical viewpoint in these neuron models. For an artificial and biological neuron model, auditory, visual, and perceptual effects should be included to describe the responses to acoustic, optical, or piezoelectric signals. In this way, multiple channel inputs can be processed to guide the body morphology and movements. Furthermore, when synapse coupling is explained as field coupling, it is interesting to investigate the collective responses of neural networks and the weight of the contribution of the electromagnetic field of each neuron is kept open. Noise is known to play an important role in mode transition and information encoding of neural activities. From a physical viewpoint, external noise in nonlinear and neural circuits can be considered as a stochastic disturbance resulting from electromagnetic radiation, which can be estimated by low frequency, high frequency, Gaussian white noise, and color noise. Based on our neuron model (Ma and Tang, 2015) with electromagnetic induction, electromagnetic radiation can be described by adding an appropriate magnetic flux function to the dynamical equation for magnetic flux (Lu et al., 2017; Ge et al., 2018a; Jin et al., 2019). That is, the external magnetic field can change the magnetic flux across the membrane and then the induction current can be used to estimate the electromagnetic induction. However, the physical electric field cannot be estimated directly when the magnetic field is changed.

In a summary, building a reliable neuron model with biophysical effects is critical for estimating the mode transition and coexistence of multiple modes of electrical activity. Considering the difference in perceptive function, the photoelectric, piezoelectric, and acoustoelectric conversion can be estimated to build a multifunctional neuron model. Thus, a stimulus of light, sound, or mechanical stress can be included for designing intelligent neuron sensors and processors. Also, the dependent and independent relations between different sense functions should be recognized. From the viewpoint of complex networks, path optimization in one-layer, multi-layer, and cluster networks should be reconsidered so that signal can be propagated in the most effective way, while minimizing energy consumption. The involvement of field

coupling in multiscale networks gives new insights and guidance for exploring neurodynamics.

### Contributors

Jun MA designed the research and presented the discussion on model setting, collective behavior of neural networks, and open problems. Zhuo-qin YANG presented the discussion on synaptic plasticity. Li-jian YANG and Jun TANG presented the discussion on astrocytes. Jun MA revised and edited the final version.

### Conflict of interest

Jun MA, Zhuo-qin YANG, Li-jian YANG, and Jun TANG declare that they have no conflict of interest.

### References

- Abbott LF, Nelson SB, 2000. Synaptic plasticity: taming the beast. *Nature Neuroscience*, 3(11):1178-1183. <https://doi.org/10.1038/81453>
- Abraham WC, Bear MF, 1996. Metaplasticity: the plasticity of synaptic plasticity. *Trends in Neurosciences*, 19(4):126-130. [https://doi.org/10.1016/S0166-2236\(96\)80018-X](https://doi.org/10.1016/S0166-2236(96)80018-X)
- Ajay SM, Bhalla US, 2004. A role for ERKII in synaptic pattern selectivity on the time-scale of minutes. *European Journal of Neuroscience*, 20(10):2671-2680. <https://doi.org/10.1111/j.1460-9568.2004.03725.x>
- Ajay SM, Bhalla US, 2007. A propagating ERKII switch forms zones of elevated dendritic activation correlated with plasticity. *HFSP Journal*, 1(1):49-66. <https://doi.org/10.2976/1.2721383/10.2976/1>
- Allegrini P, Fronzoni L, Pirino D, 2009. The influence of the astrocyte field on neuronal dynamics and synchronization. *Journal of Biological Physics*, 35(4):413-423. <https://doi.org/10.1007/s10867-009-9166-8>
- Amiri M, Bahrami F, Janahmadi M, 2012a. Functional contributions of astrocytes in synchronization of a neuronal network model. *Journal of Theoretical Biology*, 292:60-70. <https://doi.org/10.1016/j.jtbi.2011.09.013>
- Amiri M, Bahrami F, Janahmadi M, 2012b. Modified thalamocortical model: a step towards more understanding of the functional contribution of astrocytes to epilepsy. *Journal of Computational Neuroscience*, 33(2):285-299. <https://doi.org/10.1007/s10827-012-0386-8>
- Amiri M, Bahrami F, Janahmadi M, 2012c. On the role of astrocytes in epilepsy: a functional modeling approach. *Neuroscience Research*, 72(2):172-180. <https://doi.org/10.1016/j.neures.2011.11.006>
- Amiri M, Hosseinmardi N, Bahrami F, et al., 2013. Astrocyte-neuron interaction as a mechanism responsible for generation of neural synchrony: a study based on modeling and experiments. *Journal of Computational Neuroscience*, 34(3):489-504. <https://doi.org/10.1007/s10827-012-0432-6>

- Araque A, Carmignoto G, Haydon PG, et al., 2014. Gliotransmitters travel in time and space. *Neuron*, 81(4):728-739.  
<https://doi.org/10.1016/j.neuron.2014.02.007>
- Azghadi MR, Linares-Barranco B, Abbott D, et al., 2017. A hybrid CMOS-memristor neuromorphic synapse. *IEEE Transactions on Biomedical Circuits and Systems*, 11(2):434-445.  
<https://doi.org/10.1109/TBCAS.2016.2618351>
- Bao H, Liu WB, Chen M, 2019. Hidden extreme multistability and dimensionality reduction analysis for an improved non-autonomous memristive FitzHugh–Nagumo circuit. *Nonlinear Dynamics*, 96(3):1879-1894.  
<https://doi.org/10.1007/s11071-019-04890-1>
- Bear MF, Malenka RC, 1994. Synaptic plasticity: LTP and LTD. *Current Opinion in Neurobiology*, 4(3):389-399.  
[https://doi.org/10.1016/0959-4388\(94\)90101-5](https://doi.org/10.1016/0959-4388(94)90101-5)
- Bennett MR, Farnell L, Gibson WG, 2008. Origins of blood volume change due to glutamatergic synaptic activity at astrocytes abutting on arteriolar smooth muscle cells. *Journal of Theoretical Biology*, 250(1):172-185.  
<https://doi.org/10.1016/j.jtbi.2007.08.024>
- Bezprozvanny I, Watras J, Ehrlich BE, 1991. Bell-shaped calcium-response curves of Ins(1,4,5)P<sub>3</sub>- and calcium-gated channels from endoplasmic reticulum of cerebellum. *Nature*, 351(6329):751-754.  
<https://doi.org/10.1038/351751a0>
- Bhalla US, 2002. Mechanisms for temporal tuning and filtering by postsynaptic signaling pathways. *Biophysical Journal*, 83(2):740-752.  
[https://doi.org/10.1016/S0006-3495\(02\)75205-3](https://doi.org/10.1016/S0006-3495(02)75205-3)
- Bhalla US, 2004. Signaling in small subcellular volumes. II. Stochastic and diffusion effects on synaptic network properties. *Biophysical Journal*, 87(2):745-753.  
<https://doi.org/10.1529/biophysj.104.040501>
- Bhalla US, Iyengar R, 1999. Emergent properties of networks of biological signaling pathways. *Science*, 283(5400):381-387.  
<https://doi.org/10.1126/science.283.5400.381>
- Blackwell KT, Jedrzejewska-Szmek J, 2013. Molecular mechanisms underlying neuronal synaptic plasticity: systems biology meets computational neuroscience in the wilds of synaptic plasticity. *Wiley Interdisciplinary Reviews: Systems Biology and Medicine*, 5(6):717-731.  
<https://doi.org/10.1002/wsbm.1240>
- Bliss TVP, Lomo T, 1973. Long-lasting potentiation of synaptic transmission in the dentate area of the anaesthetized rabbit following stimulation of the perforant path. *The Journal of Physiology*, 232(2):331-356.  
<https://doi.org/10.1113/jphysiol.1973.sp010273>
- Bliss TVP, Gardner-Medwin AR, 1973. Long-lasting potentiation of synaptic transmission in the dentate area of the unanaesthetized rabbit following stimulation of the perforant path. *The Journal of Physiology*, 232(2):357-374.  
<https://doi.org/10.1113/jphysiol.1973.sp010274>
- Bliss TVP, Collingridge GL, 1993. A synaptic model of memory: long-term potentiation in the hippocampus. *Nature*, 361(6407):31-39.  
<https://doi.org/10.1038/361031a0>
- Bui L, Glavinović MI, 2013. Synaptic activity slows vesicular replenishment at excitatory synapses of rat hippocampus. *Cognitive Neurodynamics*, 7(2):105-120.  
<https://doi.org/10.1007/s11571-012-9232-y>
- Buonomano DV, 2000. Decoding temporal information: a model based on short-term synaptic plasticity. *Journal of Neuroscience*, 20(3):1129-1141.  
<https://doi.org/10.1523/JNEUROSCI.20-03-01129.2000>
- Busciglio J, Lorenzo A, Yankner BA, 1992. Methodological variables in the assessment of beta amyloid neurotoxicity. *Neurobiology of Aging*, 13(5):609-612.  
[https://doi.org/10.1016/0197-4580\(92\)90065-6](https://doi.org/10.1016/0197-4580(92)90065-6)
- Carro-Pérez I, Sánchez-López C, González-Hernández HG, 2018. Experimental verification of a memristive neural network. *Nonlinear Dynamics*, 93(4):1823-1840.  
<https://doi.org/10.1007/s11071-018-4291-1>
- Chan SC, Mok SY, Ng DWK, et al., 2017. The role of neuron–glia interactions in the emergence of ultra-slow oscillations. *Biological Cybernetics*, 111(5-6):459-472.  
<https://doi.org/10.1007/s00422-017-0740-z>
- Chander BS, Chakravarthy VS, 2012. A computational model of neuro-glio-vascular loop interactions. *PLoS One*, 7(11):e48802.  
<https://doi.org/10.1371/journal.pone.0048802>
- Coba MP, Pocklington AJ, Collins MO, et al., 2009. Neurotransmitters drive combinatorial multistate postsynaptic density networks. *Science Signaling*, 2(68):ra19.  
<https://doi.org/10.1126/scisignal.2000102>
- Collins MO, Yu L, Coba MP, et al., 2005. Proteomic analysis of in vivo phosphorylated synaptic proteins. *The Journal of Biological Chemistry*, 280(7):5972-5982.  
<https://doi.org/10.1074/jbc.M411220200>
- Covi E, Brivio S, Serb A, et al., 2016. Analog memristive synapse in spiking networks implementing unsupervised learning. *Frontiers in Neuroscience*, 10:482.  
<https://doi.org/10.3389/fnins.2016.00482>
- Dani JW, Chernjavsky A, Smith SJ, 1992. Neuronal activity triggers calcium waves in hippocampal astrocyte networks. *Neuron*, 8(3):429-440.  
[https://doi.org/10.1016/0896-6273\(92\)90271-E](https://doi.org/10.1016/0896-6273(92)90271-E)
- de Pittà M, Volman V, Berry H, et al., 2012. Computational quest for understanding the role of astrocyte signaling in synaptic transmission and plasticity. *Frontiers in Computational Neuroscience*, 6:98.  
<https://doi.org/10.3389/fncom.2012.00098>
- de Young GW, Keizer J, 1992. A single-pool inositol 1,4,5-trisphosphate-receptor-based model for agonist-stimulated oscillations in Ca<sup>2+</sup> concentration. *Proceedings of the National Academy of Sciences of the United States of America*, 89(20):9895-9899.  
<https://doi.org/10.1073/pnas.89.20.9895>
- Engert F, Bonhoeffer T, 1999. Dendritic spine changes associated with hippocampal long-term synaptic plasticity.

- Nature*, 399(6731):66-70.  
<https://doi.org/10.1038/19978>
- Fitzhugh R, 1966. Theoretical effect of temperature on threshold in the Hodgkin-Huxley nerve model. *The Journal of General Physiology*, 49(5):989-1005.  
<https://doi.org/10.1085/jgp.49.5.989>
- Gamble E, Koch C, 1987. The dynamics of free calcium in dendritic spines in response to repetitive synaptic input. *Science*, 236(4806):1311-1315.  
<https://doi.org/10.1126/science.3495885>
- Ge MY, Xu Y, Zhang ZK, et al., 2018a. Autaptic modulation-induced neuronal electrical activities and wave propagation on network under electromagnetic induction. *The European Physical Journal Special Topics*, 227(7-9):799-809.  
<https://doi.org/10.1140/epjst/e2018-700141-7>
- Ge MY, Jia Y, Xu Y, et al., 2018b. Mode transition in electrical activities of neuron driven by high and low frequency stimulus in the presence of electromagnetic induction and radiation. *Nonlinear Dynamics*, 91(1):515-523.  
<https://doi.org/10.1007/s11071-017-3886-2>
- Giaume C, Koulakoff A, Roux L, et al., 2010. Astroglial networks: a step further in neuroglial and gliovascular interactions. *Nature Reviews Neuroscience*, 11(2):87-99.  
<https://doi.org/10.1038/nrn2757>
- Gibson WG, Farnell L, Bennett MR, 2007. A computational model relating changes in cerebral blood volume to synaptic activity in neurons. *Neurocomputing*, 70(10-12):1674-1679.  
<https://doi.org/10.1016/j.neucom.2006.10.071>
- Goldwyn JH, Imenov NS, Famulare M, et al., 2011. Stochastic differential equation models for ion channel noise in Hodgkin-Huxley neurons. *Physical Review E*, 83(4):041908.  
<https://doi.org/10.1103/PhysRevE.83.041908>
- González-Miranda JM, 2007. Complex bifurcation structures in the Hindmarsh-Rose neuron model. *International Journal of Bifurcation and Chaos*, 17(9):3071-3083.  
<https://doi.org/10.1142/S0218127407018877>
- Gu HG, Chen SG, 2014. Potassium-induced bifurcations and chaos of firing patterns observed from biological experiment on a neural pacemaker. *Science China Technological Sciences*, 57(5):864-871.  
<https://doi.org/10.1007/s11431-014-5526-0>
- Gu HG, Pan BB, 2015a. A four-dimensional neuronal model to describe the complex nonlinear dynamics observed in the firing patterns of a sciatic nerve chronic constriction injury model. *Nonlinear Dynamics*, 81(4):2107-2126.  
<https://doi.org/10.1007/s11071-015-2129-7>
- Gu HG, Pan BB, 2015b. Identification of neural firing patterns, frequency and temporal coding mechanisms in individual aortic baroreceptors. *Frontiers in Computational Neuroscience*, 9:108.  
<https://doi.org/10.3389/fncom.2015.00108>
- Gu HG, Pan BB, Chen GR, et al., 2014a. Biological experimental demonstration of bifurcations from bursting to spiking predicted by theoretical models. *Nonlinear Dynamics*, 78(1):391-407.  
<https://doi.org/10.1007/s11071-014-1447-5>
- Gu HG, Pan BB, Xu J, 2014b. Experimental observation of spike, burst and chaos synchronization of calcium concentration oscillations. *EPL (Europhysics Letters)*, 106(5):50003.  
<https://doi.org/10.1209/0295-5075/106/50003>
- Gu HG, Pan BB, Li YY, 2015. The dependence of synchronization transition processes of coupled neurons with co-existing spiking and bursting on the control parameter, initial value, and attraction domain. *Nonlinear Dynamics*, 82(3):1191-1210.  
<https://doi.org/10.1007/s11071-015-2226-7>
- Guo SL, Tang J, Ma J, et al., 2017. Autaptic modulation of electrical activity in a network of neuron-coupled astrocyte. *Complexity*, 2017:4631602.  
<https://doi.org/10.1155/2017/4631602>
- Hadfield J, Plank MJ, David T, 2013. Modeling secondary messenger pathways in neurovascular coupling. *Bulletin of Mathematical Biology*, 75(3):428-443.  
<https://doi.org/10.1007/s11538-013-9813-x>
- Halassa MM, Haydon PG, 2010. Integrated brain circuits: astrocytic networks modulate neuronal activity and behavior. *Annual Review of Physiology*, 72:335-355.  
<https://doi.org/10.1146/annurev-physiol-021909-135843>
- Hassard B, 1978. Bifurcation of periodic solutions of the Hodgkin-Huxley model for the squid giant axon. *Journal of Theoretical Biology*, 71(3):401-420.  
[https://doi.org/10.1016/0022-5193\(78\)90168-6](https://doi.org/10.1016/0022-5193(78)90168-6)
- Hayer A, Bhalla US, 2005. Molecular switches at the synapse emerge from receptor and kinase traffic. *PLoS Computational Biology*, 1(2):e20.  
<https://doi.org/10.1371/journal.pcbi.0010020>
- Henneberger C, Papouin T, Oliet SHR, et al., 2010. Long-term potentiation depends on release of D-serine from astrocytes. *Nature*, 463(7278):232-236.  
<https://doi.org/10.1038/nature08673>
- Hodgkin AL, Huxley AF, 1952. A quantitative description of membrane current and its application to conduction and excitation in nerve. *The Journal of Physiology*, 117(4):500-544.  
<https://doi.org/10.1113/jphysiol.1952.sp004764>
- Höfer T, Venance L, Giaume C, 2002. Control and plasticity of intercellular calcium waves in astrocytes: a modeling approach. *Journal of Neuroscience*, 22(12):4850-4859.  
<https://doi.org/10.1523/JNEUROSCI.22-12-04850.2002>
- Holmes RM, Loew LM, 2008. Geometry shapes cell signaling network output. *Chemistry & Biology*, 15(6):523-524.  
<https://doi.org/10.1016/j.chembiol.2008.06.001>
- Holmes WR, Levy WB, 1990. Insights into associative long-term potentiation from computational models of NMDA receptor-mediated calcium influx and intracellular calcium concentration changes. *Journal of Neurophysiology*, 63(5):1148-1168.  
<https://doi.org/10.1152/jn.1990.63.5.1148>

- Hu XY, Liu CX, Liu L, et al., 2016. An electronic implementation for Morris–Lecar neuron model. *Nonlinear Dynamics*, 84(4):2317-2332.  
<https://doi.org/10.1007/s11071-016-2647-y>
- Irvine JM, Blackwell KT, Alkon DL, et al., 1994. Angular separation in neural networks. *Journal of Artificial Neural Networks*, 1(1):169-182.
- Ito M, 1989. Long-term depression. *Annual Review of Neuroscience*, 12:85-102.  
<https://doi.org/10.1146/annurev.ne.12.030189.000505>
- Jin WY, Wang A, Ma J, et al., 2019. Effects of electromagnetic induction and noise on the regulation of sleep wake cycle. *Science China Technological Sciences*, in press.  
<https://doi.org/10.1007/s11431-018-9423-x>
- Junge HJ, Rhee JS, Jahn O, et al., 2004. Calmodulin and Munc13 form a  $\text{Ca}^{2+}$  sensor/effector complex that controls short-term synaptic plasticity. *Cell*, 118(3):389-401.  
<https://doi.org/10.1016/j.cell.2004.06.029>
- Kawato M, Hamaguchi T, Murakami F, et al., 1984. Quantitative analysis of electrical properties of dendritic spines. *Biological Cybernetics*, 50(6):447-454.  
<https://doi.org/10.1007/BF00335202>
- Kenny A, Plank MJ, David T, 2018. The role of astrocytic calcium and TRPV4 channels in neurovascular coupling. *Journal of Computational Neuroscience*, 44(1):97-114.  
<https://doi.org/10.1007/s10827-017-0671-7>
- Khakh BS, Sofroniew MV, 2015. Diversity of astrocyte functions and phenotypes in neural circuits. *Nature Neuroscience*, 18(7):942-952.  
<https://doi.org/10.1038/nn.4043>
- Kim SY, Lim W, 2018. Effect of spike-timing-dependent plasticity on stochastic burst synchronization in a scale-free neuronal network. *Cognitive Neurodynamics*, 12(3):315-342.  
<https://doi.org/10.1007/s11571-017-9470-0>
- Kobe DH, 1986. Helmholtz's theorem revisited. *American Journal of Physics*, 54(6):552-554.  
<https://doi.org/10.1119/1.14562>
- Kotaleski JH, Blackwell KT, 2010. Modelling the molecular mechanisms of synaptic plasticity using systems biology approaches. *Nature Reviews Neuroscience*, 11(4):239-251.  
<https://doi.org/10.1038/nrn2807>
- Lavrentovich M, Hemkin S, 2008. A mathematical model of spontaneous calcium(II) oscillations in astrocytes. *Journal of Theoretical Biology*, 251(4):553-560.  
<https://doi.org/10.1016/j.jtbi.2007.12.011>
- Li XM, 2014. Signal integration on the dendrites of a pyramidal neuron model. *Cognitive Neurodynamics*, 8(1):81-85.  
<https://doi.org/10.1007/s11571-013-9252-2>
- Li YX, Rinzel J, 1994. Equations for  $\text{InsP}_3$  receptor-mediated  $[\text{Ca}^{2+}]_i$  oscillations derived from a detailed kinetic model: a Hodgkin-Huxley like formalism. *Journal of Theoretical Biology*, 166(4):461-473.  
<https://doi.org/10.1006/jtbi.1994.1041>
- Lisman J, Goldring M, 1988a. Evaluation of a model of long-term memory based on the properties of the  $\text{Ca}^{2+}$ /calmodulin-dependent protein kinase. *Journal de Physiologie*, 83(3):187-197.
- Lisman J, Goldring M, 1988b. Feasibility of long-term storage of graded information by the  $\text{Ca}^{2+}$ /calmodulin-dependent protein kinase molecules of the postsynaptic density. *Proceedings of the National Academy of Sciences of the United States of America*, 85(14):5320-5324.  
<https://doi.org/10.1073/pnas.85.14.5320>
- Liu Y, Li CG, 2013. Stochastic resonance in feedforward-loop neuronal network motifs in astrocyte field. *Journal of Theoretical Biology*, 335:265-275.  
<https://doi.org/10.1016/j.jtbi.2013.07.007>
- Liu Y, Ren GD, Zhou P, et al., 2019. Synchronization in networks of initially independent dynamical systems. *Physica A: Statistical Mechanics and Its Applications*, 520:370-380.  
<https://doi.org/10.1016/j.physa.2019.01.030>
- Liu ZL, Ma J, Zhang G, et al., 2019a. Synchronization control between two Chua's circuits via capacitive coupling. *Applied Mathematics and Computation*, 360:94-106.  
<https://doi.org/10.1016/j.amc.2019.05.004>
- Liu ZL, Wang CN, Zhang G, et al., 2019b. Synchronization between neural circuits connected by hybrid synapse. *International Journal of Modern Physics B*, 33(16):1950170.  
<https://doi.org/10.1142/S0217979219501704>
- Lu LL, Jia Y, Liu WH, et al., 2017. Mixed stimulus-induced mode selection in neural activity driven by high and low frequency current under electromagnetic radiation. *Complexity*, 2017:7628537.  
<https://doi.org/10.1155/2017/7628537>
- Lu LL, Jia Y, Kirunda JB, et al., 2019. Effects of noise and synaptic weight on propagation of subthreshold excitatory postsynaptic current signal in a feed-forward neural network. *Nonlinear Dynamics*, 95(2):1673-1686.  
<https://doi.org/10.1007/s11071-018-4652-9>
- Lv M, Ma J, Yao YG, et al., 2019. Synchronization and wave propagation in neuronal network under field coupling. *Science China Technological Sciences*, 62(3):448-457.  
<https://doi.org/10.1007/s11431-018-9268-2>
- Ma J, Tang J, 2015. A review for dynamics of collective behaviors of network of neurons. *Science China Technological Sciences*, 58(12):2038-2045.  
<https://doi.org/10.1007/s11431-015-5961-6>
- Ma J, Qin HX, Song XL, et al., 2015a. Pattern selection in neuronal network driven by electric autapses with diversity in time delays. *International Journal of Modern Physics B*, 29(1):1450239.  
<https://doi.org/10.1142/S0217979214502397>
- Ma J, Song XL, Tang J, et al., 2015b. Wave emitting and propagation induced by autapse in a forward feedback neuronal network. *Neurocomputing*, 167:378-389.  
<https://doi.org/10.1016/j.neucom.2015.04.056>
- Ma J, Xu Y, Wang CN, et al., 2016a. Pattern selection and self-organization induced by random boundary initial



- values in a neuronal network. *Physica A: Statistical Mechanics and Its Applications*, 461:586-594.  
<https://doi.org/10.1016/j.physa.2016.06.075>
- Ma J, Xu Y, Ren GD, et al., 2016b. Prediction for breakup of spiral wave in a regular neuronal network. *Nonlinear Dynamics*, 84(2):497-509.  
<https://doi.org/10.1007/s11071-015-2502-6>
- Ma J, Wu FQ, Hayat T, et al., 2017. Electromagnetic induction and radiation-induced abnormality of wave propagation in excitable media. *Physica A: Statistical Mechanics and Its Applications*, 486:508-516.  
<https://doi.org/10.1016/j.physa.2017.05.075>
- Ma J, Zhang G, Hayat T, et al., 2019. Model electrical activity of neuron under electric field. *Nonlinear Dynamics*, 95: 1585-1598.  
<https://doi.org/10.1007/s11071-018-4646-7>
- Ma SY, Yao Z, Zhang Y, et al., 2019. Phase synchronization and lock between memristive circuits under field coupling. *AEU-International Journal of Electronics and Communications*, 105:177-185.  
<https://doi.org/10.1016/j.aeue.2019.04.018>
- Malenka RC, Bear MF, 2004. LTP and LTD: an embarrassment of riches. *Neuron*, 44(1):5-21.  
<https://doi.org/10.1016/j.neuron.2004.09.012>
- Manninen T, Hituri K, Kotaleski JH, et al., 2010. Postsynaptic signal transduction models for long-term potentiation and depression. *Frontiers in Computational Neuroscience*, 4: 152.  
<https://doi.org/10.3389/fncom.2010.00152>
- Manninen T, Havela R, Linne ML, 2018. Computational models for calcium-mediated astrocyte functions. *Frontiers in Computational Neuroscience*, 12:14.  
<https://doi.org/10.3389/fncom.2018.00014>
- Mao XC, 2017. Complicated dynamics of a ring of nonidentical FitzHugh–Nagumo neurons with delayed couplings. *Nonlinear Dynamics*, 87(4):2395-2406.  
<https://doi.org/10.1007/s11071-016-3198-y>
- McCormick DA, Shu YS, Yu YG, 2007. Neurophysiology: Hodgkin and Huxley model—still standing? *Nature*, 445(7123):E1-E2.  
<https://doi.org/10.1038/nature05523>
- Mei GF, Wu XQ, Ning D, et al., 2016. Finite-time stabilization of complex dynamical networks via optimal control. *Complexity*, 21(S1):417-425.  
<https://doi.org/10.1002/cplx.21755>
- Mei GF, Wu XQ, Wang YF, et al., 2018. Compressive-sensing-based structure identification for multilayer networks. *IEEE Transactions on Cybernetics*, 48(2):754-764.  
<https://doi.org/10.1109/TCYB.2017.2655511>
- Mesiti F, Floor PA, Balasingham I, 2015. Astrocyte to neuron communication channels with applications. *IEEE Transactions on Molecular, Biological and Multi-Scale Communications*, 1(2):164-175.  
<https://doi.org/10.1109/TMBMC.2015.2501743>
- Morris C, Lecar H, 1981. Voltage oscillations in the barnacle giant muscle fiber. *Biophysical Journal*, 35(1):193-213.  
[https://doi.org/10.1016/S0006-3495\(81\)84782-0](https://doi.org/10.1016/S0006-3495(81)84782-0)
- Mostaghimi S, Nazarimehr F, Jafari S, et al., 2019. Chemical and electrical synapse-modulated dynamical properties of coupled neurons under magnetic flow. *Applied Mathematics and Computation*, 348:42-56.  
<https://doi.org/10.1016/j.amc.2018.11.030>
- Mvogo A, Takembo CN, Ekobena Fouda HP, et al., 2017. Pattern formation in diffusive excitable systems under magnetic flow effects. *Physics Letters A*, 381(28):2264-2271.  
<https://doi.org/10.1016/j.physleta.2017.05.020>
- Nadkarni S, Jung P, 2003. Spontaneous oscillations of dressed neurons: a new mechanism for epilepsy? *Physical Review Letters*, 91(26):268101.  
<https://doi.org/10.1103/PhysRevLett.91.268101>
- Nadkarni S, Jung P, 2007. Modeling synaptic transmission of the tripartite synapse. *Physical Biology*, 4(1):1-9.  
<https://doi.org/10.1088/1478-3975/4/1/001>
- Navarrete M, Díez A, Araque A, 2014. Astrocytes in endocannabinoid signalling. *Philosophical Transactions of the Royal Society B: Biological Sciences*, 369(1654): 20130599.  
<https://doi.org/10.1098/rstb.2013.0599>
- Nazari S, Faez K, Amiri M, 2017. A multiplier-less digital design of a bio-inspired stimulator to suppress synchronized regime in a large-scale, sparsely connected neural network. *Neural Computing and Applications*, 28(2):375-390.  
<https://doi.org/10.1007/s00521-015-2071-0>
- Nestler EJ, 2001. Molecular basis of long-term plasticity underlying addiction. *Nature Reviews Neuroscience*, 2(2): 119-128.  
<https://doi.org/10.1038/35053570>
- Neves SR, Tsokas P, Sarkar A, et al., 2008. Cell shape and negative links in regulatory motifs together control spatial information flow in signaling networks. *Cell*, 133(4):666-680.  
<https://doi.org/10.1016/j.cell.2008.04.025>
- Newman EA, Zahs KR, 1997. Calcium waves in retinal glial cells. *Science*, 275(5301):844-847.  
<https://doi.org/10.1126/science.275.5301.844>
- Pan B, Zucker RS, 2009. A general model of synaptic transmission and short-term plasticity. *Neuron*, 62(4):539-554.  
<https://doi.org/10.1016/j.neuron.2009.03.025>
- Park S, Chu M, Kim J, et al., 2015. Electronic system with memristive synapses for pattern recognition. *Scientific Reports*, 5:10123.  
<https://doi.org/10.1038/srep10123>
- Parpura V, Basarsky TA, Liu F, et al., 1994. Glutamate-mediated astrocyte–neuron signalling. *Nature*, 369(6483): 744-747.  
<https://doi.org/10.1038/369744a0>
- Patel GN, DeWeerth SP, 1997. Analogue VLSI morris-lecar

- neuron. *Electronics Letters*, 33(12):997-998.  
<https://doi.org/10.1049/el:19970686>
- Pellionisz AJ, 1989. Neural geometry: towards a fractal model of neurons. In: Cotterill RMJ (Ed.), *Models of Brain Function*. Cambridge University Press, Cambridge, UK, p.453-464.
- Perea G, Navarrete M, Araque A, 2009. Tripartite synapses: astrocytes process and control synaptic information. *Trends in Neurosciences*, 32(8):421-431.  
<https://doi.org/10.1016/j.tins.2009.05.001>
- Poskanzer KE, Yuste R, 2016. Astrocytes regulate cortical state switching in vivo. *Proceedings of the National Academy of Sciences of the United States of America*, 113(19):E2675-E2684.  
<https://doi.org/10.1073/pnas.1520759113>
- Pospischil M, Toledo-Rodriguez M, Monier C, et al., 2008. Minimal Hodgkin-Huxley type models for different classes of cortical and thalamic neurons. *Biological Cybernetics*, 99(4-5):427-441.  
<https://doi.org/10.1007/s00422-008-0263-8>
- Postnov DE, Ryazanova LS, Sosnovtseva OV, 2007. Functional modeling of neural-glia interaction. *Biosystems*, 89(1-3):84-91.  
<https://doi.org/10.1016/j.biosystems.2006.04.012>
- Qin HX, Ma J, Jin WY, et al., 2014. Dynamics of electric activities in neuron and neurons of network induced by autapses. *Science China Technological Sciences*, 57(5): 936-946.  
<https://doi.org/10.1007/s11431-014-5534-0>
- Qu ZL, Hu G, Garfinkel A, et al., 2014. Nonlinear and stochastic dynamics in the heart. *Physics Reports*, 543(2): 61-162.  
<https://doi.org/10.1016/j.physrep.2014.05.002>
- Ren GD, Zhou P, Ma J, et al., 2017. Dynamical response of electrical activities in digital neuron circuit driven by autapse. *International Journal of Bifurcation and Chaos*, 27(12):1750187.  
<https://doi.org/10.1142/S0218127417501875>
- Rostami Z, Pham VT, Jafari S, et al., 2018. Taking control of initiated propagating wave in a neuronal network using magnetic radiation. *Applied Mathematics and Computation*, 338:141-151.  
<https://doi.org/10.1016/j.amc.2018.06.004>
- Salin PA, Scanziani M, Malenka RC, et al., 1996. Distinct short-term plasticity at two excitatory synapses in the hippocampus. *Proceedings of the National Academy of Sciences of the United States of America*, 93(23):13304-13309.  
<https://doi.org/10.1073/pnas.93.23.13304>
- Schiegg A, Gerstner W, Ritz R, et al., 1985. Intracellular Ca<sup>2+</sup> stores can account for the time course of LTP induction: a model of Ca<sup>2+</sup> dynamics in dendritic spines. *American Physiological Society*, 74(3):1046-1055.  
<https://doi.org/10.1152/jn.1995.74.3.1046>
- Seung HS, Lee DD, Reis BY, et al., 2000. The autapse: a simple illustration of short-term analog memory storage by tuned synaptic feedback. *Journal of Computational Neuroscience*, 9(2):171-185.  
<https://doi.org/10.1023/A:1008971908649>
- Sharma SK, Haobijam D, Singh SS, et al., 2019. Neuronal communication: stochastic neuron dynamics and multi-synchrony states. *AEU-International Journal of Electronics and Communications*, 100:75-85.  
<https://doi.org/10.1016/j.aeue.2019.01.006>
- Sloan SA, Barres BA, 2014. Looks can be deceiving: reconsidering the evidence for gliotransmission. *Neuron*, 84(6): 1112-1115.  
<https://doi.org/10.1016/j.neuron.2014.12.003>
- Song XL, Wang CN, Ma J, et al., 2015. Transition of electric activity of neurons induced by chemical and electric autapses. *Science China Technological Sciences*, 58(6): 1007-1014.  
<https://doi.org/10.1007/s11431-015-5826-z>
- Song XL, Wang HT, Chen Y, 2018. Coherence resonance in an autaptic Hodgkin-Huxley neuron with time delay. *Nonlinear Dynamics*, 94(1):141-150.  
<https://doi.org/10.1007/s11071-018-4349-0>
- Stent GS, 1984. Semantics and neural development. In: Sharma CS (Ed.), *Organizing Principles of Neural Development*. Springer, Boston, USA, p.145-160.  
[https://doi.org/10.1007/978-1-4684-4802-3\\_8](https://doi.org/10.1007/978-1-4684-4802-3_8)
- Storace M, Linaro D, de Lange E, 2008. The Hindmarsh-Rose neuron model: bifurcation analysis and piecewise-linear approximations. *Chaos: An Interdisciplinary Journal of Nonlinear Science*, 18(3):033128.  
<https://doi.org/10.1063/1.2975967>
- Sun XJ, Liu ZF, Perc M, 2019. Effects of coupling strength and network topology on signal detection in small-world neuronal networks. *Nonlinear Dynamics*, 96(3):2145-2155.  
<https://doi.org/10.1007/s11071-019-04914-w>
- Takembo CN, Mvogo A, Ekobena Fouda HP, et al., 2018. Modulated wave formation in myocardial cells under electromagnetic radiation. *International Journal of Modern Physics B*, 32(14):1850165.  
<https://doi.org/10.1142/S0217979218501655>
- Tamaševičius A, Mykolaitis G, Tamaševičiūtė E, et al., 2015. Two-terminal feedback circuit for suppressing synchrony of the FitzHugh-Nagumo oscillators. *Nonlinear Dynamics*, 81(1-2):783-788.  
<https://doi.org/10.1007/s11071-015-2028-y>
- Tang J, Luo JM, Ma J, 2013. Information transmission in a neuron-astrocyte coupled model. *PLoS One*, 8(11): e80324.  
<https://doi.org/10.1371/journal.pone.0080324>
- Tang J, Liu TB, Ma J, et al., 2016. Effect of calcium channel noise in astrocytes on neuronal transmission. *Communications in Nonlinear Science and Numerical Simulation*, 32:262-272.  
<https://doi.org/10.1016/j.cnsns.2015.08.019>

- Tang J, Zhang J, Ma J, et al., 2017. Astrocyte calcium wave induces seizure-like behavior in neuron network. *Science China Technological Sciences*, 60(7):1011-1018.  
<https://doi.org/10.1007/s11431-016-0293-9>
- Tarai S, Mukherjee R, Gupta S, et al., 2019. Influence of pharmacological and epigenetic factors to suppress neurotrophic factors and enhance neural plasticity in stress and mood disorders. *Cognitive Neurodynamics*, 13(3): 219-237.  
<https://doi.org/10.1007/s11571-019-09522-3>
- Toivari E, Manninen T, Nahata AK, et al., 2011. Effects of transmitters and amyloid-beta peptide on calcium signals in rat cortical astrocytes: Fura-2AM measurements and stochastic model simulations. *PLoS One*, 6(3):e17914.  
<https://doi.org/10.1371/journal.pone.0017914>
- Tomba C, Braïni C, Wu BL, et al., 2014. Tuning the adhesive geometry of neurons: length and polarity control. *Soft Matter*, 10(14):2381-2387.  
<https://doi.org/10.1039/C3SM52342J>
- Trachtenberg JT, Chen BE, Knott GW, et al., 2002. Long-term in vivo imaging of experience-dependent synaptic plasticity in adult cortex. *Nature*, 420(6917):788-794.  
<https://doi.org/10.1038/nature01273>
- Tsumoto K, Kitajima H, Yoshinaga T, et al., 2006. Bifurcations in Morris–Lecar neuron model. *Neurocomputing*, 69(4-6):293-316.  
<https://doi.org/10.1016/j.neucom.2005.03.006>
- Tutkun E, Ayyildiz M, Agar E, 2010. Short-duration swimming exercise decreases penicillin-induced epileptiform ECoG activity in rats. *Acta Neurobiologiae Experimentalis*, 70(4):382-389.
- Ursino M, Cuppini C, Cappa SF, et al., 2018. A feature-based neurocomputational model of semantic memory. *Cognitive Neurodynamics*, 12(6):525-547.  
<https://doi.org/10.1007/s11571-018-9494-0>
- Uzun R, 2017. Influences of autapse and channel blockage on multiple coherence resonance in a single neuron. *Applied Mathematics and Computation*, 315:203-210.  
<https://doi.org/10.1016/j.amc.2017.07.055>
- Uzun R, Yilmaz E, Ozer M, 2017. Effects of autapse and ion channel block on the collective firing activity of Newman–Watts small-world neuronal networks. *Physica A: Statistical Mechanics and Its Applications*, 486:386-396.  
<https://doi.org/10.1016/j.physa.2017.05.049>
- Valverde F, 1976. Aspects of cortical organization related to the geometry of neurons with intra-cortical axons. *Journal of Neurocytology*, 5(5):509-529.  
<https://doi.org/10.1007/BF01175566>
- van der Loos H, Glaser EM, 1972. Autapses in neocortex cerebri: synapses between a pyramidal cell's axon and its own dendrites. *Brain Research*, 48:355-360.  
[https://doi.org/10.1016/0006-8993\(72\)90189-8](https://doi.org/10.1016/0006-8993(72)90189-8)
- Volterra A, Meldolesi J, 2005. Astrocytes, from brain glue to communication elements: the revolution continues. *Nature Reviews Neuroscience*, 6(8):626-640.  
<https://doi.org/10.1038/nrn1722>
- Wade J, McDaid L, Harkin J, et al., 2012. Self-repair in a bidirectionally coupled astrocyte-neuron (AN) system based on retrograde signaling. *Frontiers in Computational Neuroscience*, 6:76.  
<https://doi.org/10.3389/fncom.2012.00076>
- Wang CN, Ma J, 2018. A review and guidance for pattern selection in spatiotemporal system. *International Journal of Modern Physics B*, 32(6):1830003.  
<https://doi.org/10.1142/S0217979218300037>
- Wang CN, Guo SL, Xu Y, et al., 2017. Formation of autapse connected to neuron and its biological function. *Complexity*, 2017:5436737.  
<https://doi.org/10.1155/2017/5436737>
- Wang JY, Yang XL, Sun ZK, 2018. Suppressing bursting synchronization in a modular neuronal network with synaptic plasticity. *Cognitive Neurodynamics*, 12(6):625-636.  
<https://doi.org/10.1007/s11571-018-9498-9>
- Wang RB, Wang ZY, Zhu ZY, 2018. The essence of neuronal activity from the consistency of two different neuron models. *Nonlinear Dynamics*, 92(3):973-982.  
<https://doi.org/10.1007/s11071-018-4103-7>
- Wang XH, Takano T, Nedergaard M, 2009. Astrocytic calcium signaling: mechanism and implications for functional brain imaging. In: Hyder F (Ed.), *Dynamic Brain Imaging: Multi-modal Methods and in vivo Applications*. Humana Press, New York, USA, p.93-109.  
[https://doi.org/10.1007/978-1-59745-543-5\\_5](https://doi.org/10.1007/978-1-59745-543-5_5)
- Wang Y, Wang CN, Ren GD, et al., 2017. Energy dependence on modes of electric activities of neuron driven by multi-channel signals. *Nonlinear Dynamics*, 89(3):1967-1987.  
<https://doi.org/10.1007/s11071-017-3564-4>
- Wang YH, Wang RB, Xu XY, 2017. Neural energy supply-consumption properties based on Hodgkin-Huxley model. *Neural Plasticity*, 2017:6207141.  
<https://doi.org/10.1155/2017/6207141>
- Wang YY, Wang RB, 2018. An improved neuronal energy model that better captures of dynamic property of neuronal activity. *Nonlinear Dynamics*, 91(1):319-327.  
<https://doi.org/10.1007/s11071-017-3871-9>
- Wang ZY, Wang RB, Fang RY, 2015. Energy coding in neural network with inhibitory neurons. *Cognitive Neurodynamics*, 9(2):129-144.  
<https://doi.org/10.1007/s11571-014-9311-3>
- Wei H, Bu YJ, Dai DW, 2017. A decision-making model based on a spiking neural circuit and synaptic plasticity. *Cognitive Neurodynamics*, 11(5):415-431.  
<https://doi.org/10.1007/s11571-017-9436-2>
- Wei X, Wu XQ, Chen SH, et al., 2018. Cooperative epidemic spreading on a two-layered interconnected network. *SIAM Journal on Applied Dynamical Systems*, 17(2): 1503-1520.  
<https://doi.org/10.1137/17M1134202>
- Witthoft A, Karniadakis GE, 2012. A bidirectional model for

- communication in the neurovascular unit. *Journal of Theoretical Biology*, 311:80-93.  
<https://doi.org/10.1016/j.jtbi.2012.07.014>
- Witthoft A, Filosa JA, Karniadakis GE, 2013. Potassium buffering in the neurovascular unit: models and sensitivity analysis. *Biophysical Journal*, 105(9):2046-2054.  
<https://doi.org/10.1016/j.bpj.2013.09.012>
- Wu FQ, Wang CN, Xu Y, et al., 2016. Model of electrical activity in cardiac tissue under electromagnetic induction. *Scientific Reports*, 6:28.  
<https://doi.org/10.1038/s41598-016-0031-2>
- Wu FQ, Wang CN, Jin WY, et al., 2017. Dynamical responses in a new neuron model subjected to electromagnetic induction and phase noise. *Physica A: Statistical Mechanics and Its Applications*, 469:81-88.  
<https://doi.org/10.1016/j.physa.2016.11.056>
- Wu FQ, Hayat T, An XL, et al., 2018a. Can Hamilton energy feedback suppress the chameleon chaotic flow? *Nonlinear Dynamics*, 94(1):669-677.  
<https://doi.org/10.1007/s11071-018-4384-x>
- Wu FQ, Zhou P, Alsaedi A, et al., 2018b. Synchronization dependence on initial setting of chaotic systems without equilibria. *Chaos, Solitons & Fractals*, 110:124-132.  
<https://doi.org/10.1016/j.chaos.2018.03.024>
- Wu FQ, Ma J, Ren GD, 2018c. Synchronization stability between initial-dependent oscillators with periodical and chaotic oscillation. *Journal of Zhejiang University-SCIENCE A (Applied Physics & Engineering)*, 19(12):889-903.  
<https://doi.org/10.1631/jzus.A1800334>
- Wu FQ, Ma J, Zhang G, 2019. A new neuron model under electromagnetic field. *Applied Mathematics and Computation*, 347:590-599.  
<https://doi.org/10.1016/j.amc.2018.10.087>
- Xiao WW, Gu HG, Liu MR, 2016. Spatiotemporal dynamics in a network composed of neurons with different excitabilities and excitatory coupling. *Science China Technological Sciences*, 59(12):1943-1952.  
<https://doi.org/10.1007/s11431-016-6046-x>
- Xu F, Zhang JQ, Fang TT, et al., 2018. Synchronous dynamics in neural system coupled with memristive synapse. *Nonlinear Dynamics*, 92(3):1395-1402.  
<https://doi.org/10.1007/s11071-018-4134-0>
- Xu Q, Song Z, Bao H, et al., 2018. Two-neuron-based non-autonomous memristive Hopfield neural network: numerical analyses and hardware experiments. *AEU-International Journal of Electronics and Communications*, 96:66-74.  
<https://doi.org/10.1016/j.aeue.2018.09.017>
- Xu Y, Wang CN, Lv M, et al., 2016. Local pacing, noise induced ordered wave in a 2D lattice of neurons. *Neurocomputing*, 207:398-407.  
<https://doi.org/10.1016/j.neucom.2016.05.030>
- Xu Y, Jia Y, Kirunda JB, et al., 2018a. Dynamic behaviors in coupled neuron system with the excitatory and inhibitory autapse under electromagnetic induction. *Complexity*, 2018:3012743.  
<https://doi.org/10.1155/2018/3012743>
- Xu Y, Jia Y, Ge MY, et al., 2018b. Effects of ion channel blocks on electrical activity of stochastic Hodgkin-Huxley neural network under electromagnetic induction. *Neurocomputing*, 283:196-204.  
<https://doi.org/10.1016/j.neucom.2017.12.036>
- Xu YM, Yao Z, Hobiny A, et al., 2019. Differential coupling contributes to synchronization via a capacitor connection between chaotic circuits. *Frontiers of Information Technology & Electronic Engineering*, 20(4):571-583.  
<https://doi.org/10.1631/FITEE.1800499>
- Yang XL, Yu YH, Sun ZK, 2017. Autapse-induced multiple stochastic resonances in a modular neuronal network. *Chaos: An Interdisciplinary Journal of Nonlinear Science*, 27(8):083117.  
<https://doi.org/10.1063/1.4999100>
- Yang YQ, Yeo CK, 2015. Conceptual network model from sensory neurons to astrocytes of the human nervous system. *IEEE Transactions on Biomedical Engineering*, 62(7):1843-1852.  
<https://doi.org/10.1109/TBME.2015.2405549>
- Yao Z, Ma J, Yao YG, et al., 2019. Synchronization realization between two nonlinear circuits via an induction coil coupling. *Nonlinear Dynamics*, 96(1):205-217.  
<https://doi.org/10.1007/s11071-019-04784-2>
- Yue Y, Liu LW, Liu YJ, et al., 2017. Dynamical response, information transition and energy dependence in a neuron model driven by autapse. *Nonlinear Dynamics*, 90(4):2893-2902.  
<https://doi.org/10.1007/s11071-017-3850-1>
- Yuste R, Bonhoeffer T, 2001. Morphological changes in dendritic spines associated with long-term synaptic plasticity. *Annual Review of Neuroscience*, 24:1071-1089.  
<https://doi.org/10.1146/annurev.neuro.24.1.1071>
- Zayer F, Dghais W, Benabdeladhim M, et al., 2019. Low power, ultrafast synaptic plasticity in 1R-ferroelectric tunnel memristive structure for spiking neural networks. *AEU-International Journal of Electronics and Communications*, 100:56-65.  
<https://doi.org/10.1016/j.aeue.2019.01.003>
- Zeng S, Li B, Chen SQ, 2009. Simulation of spontaneous Ca<sup>2+</sup> oscillations in astrocytes mediated by voltage-gated calcium channels. *Biophysical Journal*, 97(9):2429-2437.  
<https://doi.org/10.1016/j.bpj.2009.08.030>
- Zhan FB, Liu SQ, 2017. Response of electrical activity in an improved neuron model under electromagnetic radiation and noise. *Frontiers in Computational Neuroscience*, 11:107.  
<https://doi.org/10.3389/fncom.2017.00107>
- Zhang G, Wang CN, Alsaedi A, et al., 2018. Dependence of hidden attractors on non-linearity and Hamilton energy in a class of chaotic system. *Kybernetika*, 54(4):648-663.  
<https://doi.org/10.14736/kyb-2018-4-0648>

- Zhang JH, Liao XF, 2017. Synchronization and chaos in coupled memristor-based FitzHugh-Nagumo circuits with memristor synapse. *AEU-International Journal of Electronics and Communications*, 75:82-90.  
<https://doi.org/10.1016/j.aecu.2017.03.003>
- Zhao ZG, Gu HG, 2015. The influence of single neuron dynamics and network topology on time delay-induced multiple synchronous behaviors in inhibitory coupled network. *Chaos, Solitons & Fractals*, 80:96-108.  
<https://doi.org/10.1016/j.chaos.2015.06.017>
- Zhao ZG, Gu HG, 2017. Transitions between classes of neuronal excitability and bifurcations induced by autapse. *Scientific Reports*, 7(1):6760.  
<https://doi.org/10.1038/s41598-017-07051-9>
- Zheng HW, Wang RB, Qu JY, 2016. Effect of different glucose supply conditions on neuronal energy metabolism. *Cognitive Neurodynamics*, 10(6):563-571.  
<https://doi.org/10.1007/s11571-016-9401-5>
- Zonta M, Angulo MC, Gobbo S, et al., 2003. Neuron-to-astrocyte signaling is central to the dynamic control of brain microcirculation. *Nature Neuroscience*, 6(1):43-50.  
<https://doi.org/10.1038/nn980>
- Zucker RS, 1989. Short-term synaptic plasticity. *Annual Review of Neuroscience*, 12:13-31.  
<https://doi.org/10.1146/annurev.ne.12.030189.000305>
- Zucker RS, Regehr WG, 2002. Short-term synaptic plasticity. *Annual Review of Physiology*, 64:355-405.  
<https://doi.org/10.1146/annurev.physiol.64.092501.114547>

## 中文概要

**题目:** 从物理学角度认知计算神经动力学

**目的:** 基于物理学基本原理解释神经元电活动过程中存在的物理效应, 解释突触生物功能活化过程的物理机制, 以及分析神经元建模中的电磁场效应(图1)。探讨神经元建模、胶质细胞调控、突触可塑性和神经元群体电活动的网络效应。

**创新点:** 1. 论证荷控和磁控忆阻器非线性函数在物理神经元模型构建中的作用。2. 提出神经元突触耦合的物理机制就是电场和磁场耦合(图3)。3. 研究神经元电路混合突触耦合的物理实现(图2)以及能量存储与泵浦。

**方法:** 依据物理学电磁感应定律和赫姆霍兹定理论证神经元电活动过程产生的电磁感应效应以及能量运输过程。基于忆阻器物理特性和量纲一致原理来构建物理神经元模型, 从物理角度解释突触功能实现过程的物理机制。

**结论:** 在神经元电活动过程中需考虑电磁感应效应; 场耦合可以调控神经元突触耦合作用; 在神经元网络中信号传递需考虑物理场耦合过程。

**关键词:** 神经元; 神经网络; 自突触; 哈密顿能量; 电磁感应