



## Review

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# Role of 5-hydroxytryptamine type 3 receptors in the regulation of anxiety reactions

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**Abstract:** 5-Hydroxytryptamine (5-HT) type 3 receptor (5-HT<sub>3</sub>R) is the only type of ligand-gated ion channel in the 5-HT receptor family. Through the high permeability of Na<sup>+</sup>, K<sup>+</sup>, and Ca<sup>2+</sup> and activation of subsequent voltage-gated calcium channels (VGCCs), 5-HT<sub>3</sub>R induces a rapid increase of neuronal excitability or the release of neurotransmitters from axon terminals in the central nervous system (CNS). 5-HT<sub>3</sub>Rs are widely expressed in the medial prefrontal cortex (mPFC), amygdala (AMYG), hippocampus (HIP), periaqueductal gray (PAG), and other brain regions closely associated with anxiety reactions. They have a bidirectional regulatory effect on anxiety reactions by acting on different types of cells in different brain regions. 5-HT<sub>3</sub>Rs mediate the activation of the cholecystokinin (CCK) system in the AMYG, and the  $\gamma$ -aminobutyric acid (GABA) “disinhibition” mechanism in the prelimbic area of the mPFC promotes anxiety by the activation of GABAergic intermediate inhibitory neurons (IINs). In contrast, a 5-HT<sub>3</sub>R-induced GABA “disinhibition” mechanism in the infralimbic area of the mPFC and the ventral HIP produces anxiolytic effects. 5-HT<sub>3</sub>R-mediated regulation of anxiety reactions are also activated by 5-HT<sub>3</sub>R-activated 5-HT release in the HIP and PAG. This provides a theoretical basis for the treatment of anxiety disorders or the production of anxiolytic drugs by targeting 5-HT<sub>3</sub>Rs. However, given the circuit specific modulation of 5-HT<sub>3</sub>Rs on emotion, systemic use of 5-HT<sub>3</sub>R agonism or antagonism alone seems unlikely to remedy anxiety, which deeply hinders the current clinical application of 5-HT<sub>3</sub>R drugs. Therefore, the exploitation of circuit targeting methods or a combined drug strategy might be a useful developmental approach in the future.

**Key words:** 5-Hydroxytryptamine type 3 receptor (5-HT<sub>3</sub>R); Anxiety; Medial prefrontal cortex; Amygdala; Hippocampus; Periaqueductal gray

## 1 Introduction

Anxiety is a defensive emotional response produced by the body when it suffers from various types of stressors including injuries, threatening stimuli, or acute changes in the surrounding environment. However, if the level and duration of stressors are excessive, anxiety can develop into a mental disease, namely anxiety disorder (Crocq, 2015). Anxiety disorder manifests several types of clinical symptoms, including panic disorder (PD), generalized anxiety disorder (GAD), social anxiety disorder (SAD), posttraumatic

stress disorder (PTSD), specific phobia, and obsessive-compulsive disorder (OCD), which are difficult to diagnose and treat (Giacobbe and Flint, 2018; Vu and Conant-Norville, 2021; Lakhtakia and Torous, 2022). Meanwhile, severe psychic irritation, dysfunction of the endocrine system and immune system along with several side-effects like chest tightness, insomnia, diarrhea, and vomiting occur in anxiety disorder. Because of a considerable increase of morbidity and a younger age of onset, anxiety disorder is becoming one of the major mental diseases in clinical medicine and public health (Giacobbe and Flint, 2018). Many animal studies and clinical practices have shown that dysfunction of the endogenous 5-hydroxytryptamine (5-HT) system is closely related to the occurrence and maintenance of anxiety and anxiety disorder. In mice, selective activation of somas of 5-HT neurons in the dorsal raphe nucleus (DRN) and median raphe nucleus (MRN) using optogenetic methods has induced obvious anxiety reactions, including novelty suppressed

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feeding (NSF), marble-burying behavior (MBB), and elevated-plus maze (EPM), in several emotional behavior tests (Challis et al., 2014; Marcinkiewicz et al., 2016; Abela et al., 2020). In contrast, inhibition of 5-HT neural activation in MRN and DRN by pharmacological or genetic manipulation, or direct lesion of these areas by physical methods was found to produce anxiolytic effects (Andrade et al., 2004). Moreover, in patch-clamp studies, the bursting number of action potential of 5-HT neurons in the MRN was significantly increased under an anxiety state (Abela et al., 2020). Meanwhile, abundant clinical data related to selective serotonin reuptake inhibitors (SSRIs) in the treatment of anxiety disorder have been reported recently (Hutchison et al., 2021; Murphy et al., 2021). All this evidence indicates that the mechanisms by which central nerve 5-HT systems modulate anxiety reactions are worthy of detailed study.

In central nervous system (CNS), 5-HT neurons are distributed mainly in the raphe nuclei, and send dense projections to the whole brain. The 5-HT neurotransmitters released from axon terminals of these 5-HT projections subsequently bind to 5-HT receptors (5-HTRs) to execute the modulation of brain function (Sharp and Barnes, 2020; Ślifirski et al., 2021). At least seven subtypes of 5-HTRs have been identified (5-HT<sub>1R</sub>–5-HT<sub>7R</sub>), but unlike other 5-HTRs that belong to G-protein-coupled receptors (GPCRs), 5-HT<sub>3R</sub> is the only cation channel among all these subtypes. The high permeability of 5-HT<sub>3R</sub> to Na<sup>+</sup> and Ca<sup>2+</sup> allows it to trigger an instantaneous depolarization current and therefore mediate more rapid intracellular signaling than other 5-HTR subtypes (Cortes-Altamirano et al., 2018; Zhao et al., 2018). In laboratory studies, 5-HT<sub>3R</sub>s have been suggested to have a regulatory effect on anxiety. Studies focused on 5-HT<sub>3R</sub>-targeted specific brain regions and circuits found that 5-HT<sub>3R</sub>s in different regions and circuits appeared to elicit various, even conflicting, effects on behavioral expression, implying that the role of 5-HT<sub>3R</sub>s on the regulation of anxiety tended to be bidirectional. Meanwhile, although rigorous and precise clinical research has not yet been reported, selective intervention of the function of 5-HT<sub>3R</sub>s by antagonist administration has been shown to play a positive role in the treatment of anxiety disorder (Harrell and Allan, 2003; Bhatt et al., 2013, 2017a, 2017b). By comparing clinical and laboratory studies, the lack of clinical studies seemed to be associated with the complexity of the role of

5-HT<sub>3R</sub>s on the regulation of anxiety in the brain. Therefore, more detailed knowledge of the effects of 5-HT<sub>3R</sub>s on the regulation of anxiety and the mechanisms involved is necessary to assess the potential of 5-HT<sub>3R</sub>s as a new therapeutic target for anxiety. In this paper, we describe the bidirectional role of 5-HT<sub>3R</sub>s in the regulation of anxiety and give a preliminary interpretation by depicting 5-HT<sub>3R</sub> neural networks involved in the regulation of anxiety. This information may provide valuable guidance for more in-depth research on the role of 5-HT<sub>3R</sub>s in the regulation of anxiety in animals, and for exploring the potential medicinal value of 5-HT<sub>3R</sub>s in the clinical treatment of anxiety.

## 2 5-HT<sub>3R</sub> structure and function

5-HT<sub>3R</sub> is the only type of ligand-gated ion channel (LGIC) among all the 5-HTR families. It also belongs to the Cys-loop receptor superfamily because of its Cys-loop structure in the N-terminal domain (Basak et al., 2018). Analysis of its protein conformation showed that 5-HT<sub>3R</sub> is a pseudo-symmetric pentamer formed from five transmembrane protein subunits with a central hole. When 5-HT<sub>3R</sub>s are activated, positive ions can pass through the central hole (Gibbs and Chakrapani, 2021). Five classes of 5-HT<sub>3R</sub> protein subunits, 5-HT<sub>3RA</sub>–5-HT<sub>3RE</sub>, have been isolated, each containing a long extracellular N-terminal domain, four transmembrane domains (TMDs), and a short extracellular C-terminal domain. There is also an intracellular domain between TMD3 and TMD4, regulating both receptor function and channel conductance. Except for 5-HT<sub>3RD</sub>, the subunits contain six amino acid loops close to the N-terminal domain. These amino acid loops include the “major” A–C subunit loops and the “complementary” D–F subunit loops that constitute the binding site of 5-HT<sub>3R</sub> (Celli et al., 2017). 5-HT<sub>3RA</sub> is the key protein subunit of 5-HT<sub>3R</sub> for membrane integration. All the other types of protein subunits assemble with 5-HT<sub>3RA</sub> to form the functional 5-HT<sub>3R</sub> (Barnes et al., 2009; Gibbs and Chakrapani, 2021).

The neurotransmitter 5-HT is a natural agonist of 5-HT<sub>3R</sub>, which is released mainly by 5-HT neurons derived from the MRN and DRN (Lummiss, 2012). 5-HT<sub>3R</sub> is also regulated by other neurotransmitters or

drugs. The main agonists of 5-HT<sub>3</sub>Rs include 5-HT analogues and positive allosteric modulators (PAMs). 2-Methylserotonin and its metal salts represent 5-HT analogues, which can selectively and completely activate 5-HT<sub>3</sub>R to exert its biological effects (Giordano and Gerstmann, 2004). The structurally similar compounds, 1-(3-chlorophenyl)biguanide (mCPBG) and *trans*-3-(4-methoxyphenyl)-*N*-(pentan-3-yl)acrylamide (TMPPAA) are the typical PAMs frequently used in biological and preclinical studies. Both are partial agonists of 5-HT<sub>3</sub>A protein subunits, but have a great ability to activate heterozygous 5-HT<sub>3</sub>Rs containing both 5-HT<sub>3</sub>A and 5-HT<sub>3</sub>B subunits when binding to their interface, and therefore actuate allosteric regulation of 5-HT<sub>3</sub>R responses (Kilpatrick et al., 1990; Campbell et al., 1995; Maksay et al., 2005; Gasiorek et al., 2016). The endogenous neurotransmitter dopamine (DA) has also been shown to partially agonize 5-HT<sub>3</sub>Rs. In electrophysiological studies, the peak responses of 5-HT<sub>3</sub>R induced by 1 mmol/L DA were nearly one-quarter of the maximum responses induced by 5-HT (Solt et al., 2007).

The main antagonists of 5-HT<sub>3</sub>Rs include competitive antagonists and negative allosteric modulators. The competitive antagonists block the function of 5-HT<sub>3</sub>Rs mainly through emulously preempting the binding sites of ligands on 5-HT<sub>3</sub>Rs. These types of antagonists usually have three essential elements: an aromatic ring, a carbonyl-binding group, and a nitrogen atom. The frequently used competitive antagonists, tropisetron, granisetron, bemesetron (MDL7222), and ondansetron, all have the above characteristics. These antagonists have not only been widely used in antagonistic studies of 5-HT<sub>3</sub>Rs, but also clinically used in the treatment of nausea, vomiting, and other diseases related to 5-HT<sub>3</sub>R dysfunction in cancer patients (Bhatt et al., 2021). Other synthetic 5-HT<sub>3</sub>R competitive antagonists have also been synthesized in recent laboratory studies. These include (4-benzylpiperazin-1-yl)(3-methoxyquinoxalin-2-yl)methanone (6g) and *N*-*n*-propyl-3-ethoxyquinoxaline-2-carboxamide (6n), which are both synthesized by condensation and chlorination of *o*-phenylenediamine and ketol diethyl malonate (Bhatt et al., 2013, 2017a, 2017b). Kurhe et al. (2015, 2017) synthesized 3-methoxy-*N*-*p*-tolylquinoxalin-2-carboxamide (QCM-4) based on the structure of 6n. The main negative allosteric modulators of 5-HT<sub>3</sub>R in the natural world include terpene menthol, citronellol, and geraniol.

Chemically separated substances like capsaicin, cannabidiol, and gingerol also show negative allosteric modulation of 5-HT<sub>3</sub>R. They interfere with 5-HT<sub>3</sub>R functions by conformational change (al Kury et al., 2018; el Nebrisi et al., 2020).

Electrophysiological studies have shown that 5-HT<sub>3</sub>Rs have high permeability for cations like Na<sup>+</sup> and K<sup>+</sup> and can quickly regulate the excitability of neurons (Yakel et al., 1990; Machu, 2011; Gibbs and Chakrapani, 2021). Activated 5-HT<sub>3</sub>Rs transform the Na<sup>+</sup>/K<sup>+</sup> permeability ratio of neurons from 0.05 under resting state to 0.92–0.94 under active state, inducing rapid inward cation currents, and therefore directly raising the membrane potential of neurons. These rapid inward cation currents induced by 5-HT<sub>3</sub>Rs can further activate voltage-gated calcium channels (VGCCs) leading to a rapid influx of extracellular Ca<sup>2+</sup> and the elevation of neurotransmitter release from axonal terminals of neurons. These effects have been proven to mediate the quick release of  $\gamma$ -aminobutyric acid (GABA) in inhibitory interneurons (IINs) in hippocampus (HIP) slices (Yakel and Jackson, 1988; Turner et al., 2004). In addition, laser scanning confocal imaging studies showed that 5-HT<sub>3</sub>Rs themselves have a specific permeability to Ca<sup>2+</sup>. Administration of mCPBG onto mouse hippocampal slices increased the intracellular Ca<sup>2+</sup> level directly through 5-HT<sub>3</sub>R functions, and thereby enhanced the release of neurotransmitters at the presynaptic level (Choi et al., 2007; Fawley et al., 2019). In conclusion, the modulation of neuronal activity by 5-HT<sub>3</sub>Rs includes two main aspects. First, 5-HT<sub>3</sub>Rs rapidly up-regulate neuronal excitability through Na<sup>+</sup> influx. Second, 5-HT<sub>3</sub>Rs directly or indirectly (through activation of VGCCs) mediate an increase of the intracellular Ca<sup>2+</sup> level to enhance the release of neurotransmitters in axon terminals, including 5-HT, GABA, and cholinergic and adrenergic neurotransmitters (Zhao et al., 2018). Further research of cellular sublocalization of 5-HT<sub>3</sub>Rs showed that 5-HT<sub>3</sub>Rs expressed on the postsynaptic membrane or cell body mediated mainly the rapid up-regulation of neuronal excitability, while 5-HT<sub>3</sub>Rs expressed on the presynaptic membrane mainly regulated the release of presynaptic neuronal transmitters (Sharp and Barnes, 2020).

5-HT<sub>3</sub>Rs are widely expressed in the central, peripheral, and enteric nervous systems, playing an important role in the regulation of various nervous activities, including gastrointestinal peristalsis, digestive

juice secretion, vomiting, neuroimmunity, visceral reflex, pain response, and emotional response (Cortes-Altamirano et al., 2018; Juza et al., 2020; Irving et al., 2021). In early studies, 5-HT<sub>3</sub>Rs were detected in both the forebrain and brain stem of humans—tissues closely related to anxiety reactions (Kilpatrick et al., 1987; Barnes et al., 1990). Ondansetron injection into the rat brain can significantly increase activation of neurons in these regions, indicating a potential relationship between 5-HT<sub>3</sub>Rs and anxiety (Urzedo-Rodrigues et al., 2014). By using 5-HT<sub>3</sub>AR-green fluorescent protein (GFP) transgenic mice combined with immunofluorescence technologies, Koyama et al. (2017) found that circuit-based distribution of 5-HT<sub>3</sub>R positive signals in several brain areas closely associated with anxiety, such as the prefrontal cortex, HIP, and amygdala (AMYG), providing more direct and precise morphological evidence to support the study of the modulatory effects of 5-HT<sub>3</sub>R on anxiety behavior.

### 3 Bidirectional effects of 5-HT<sub>3</sub>Rs on anxiety behaviors

There have been several studies of the association between the regulation of anxiety and emotional behavior. Although treatment strategies targeting 5-HT<sub>3</sub>Rs are still in the elementary stages, some clinical practices and pharmacological and genetic experiments have shown that 5-HT<sub>3</sub>R antagonists have appreciable or at least auxiliary anxiolytic effects. In clinical studies, several types of 5-HT<sub>3</sub>R competitive antagonists like tropisetron, zatosetron, ondansetron, and granisetron, have been proven to alleviate the unhealthy emotions of patients of different gender and ages suffering from anxiety disorders (Smith et al., 1999; Haus et al., 2000; Harmer et al., 2006). Evaluation of the therapeutic effect on OCD has shown that tropisetron and ondansetron can reverse the compulsive behavior of OCD patients to some extent, and this effect can be enhanced by combinational administration of fluvoxamine (Hewlett et al., 2003; Soltani et al., 2010). In another clinical study, after 12 weeks of oral ondansetron treatment, 64.3% of OCD patients showed reversal of anxiety (Pallanti et al., 2009). Granisetron has also been shown to have a rapid therapeutic effect in moderate or severe OCD patients (Askari et al., 2012; Serata et al., 2015; Sharafkhan et al., 2019). 5-HT<sub>3</sub>R antagonists have also been used

in symptomatic improvement of GAD and SAD patients (Lecrubier et al., 1993; McCann et al., 1997). After administration of tropisetron for GAD or ondansetron for SAD, anxiety symptoms were obviously alleviated. Currently, there is no evidence from clinical studies of other types of anxiety disorder to support a regulatory role of 5-HT<sub>3</sub>Rs, but animal models suggest that 5-HT<sub>3</sub>Rs may be a potential target in the management and treatment of anxiety disorder.

Many animal studies have shown that specific antagonism of 5-HT<sub>3</sub>Rs produces anxiolytic-like reactions. EPM is one of the classic models used to test anxiety level in the laboratory. Because its open arms can directly generate anxiety stimuli in the behavioral testing of mice, the anxiety level of animals can be detected without extra stressors or stimuli in the EPM test. Cutler (1991) and Artaiz et al. (1995) introduced granisetron orally into male DAB rats and by intraperitoneal injection into male Wistar rats. In the EPM test, significant increases in the number of animals entering open arms and the time spent in open arms were observed, indicating anxiolytic effects. Intraperitoneal injection of MDL7222 into ACI/N rats can also increase the staying time and entering frequency in open arms in the EPM test (Hensler et al., 2004). The anxiolytic-like behaviors induced by antagonistic effects on 5-HT<sub>3</sub>R have also been extensively reported in different anxiety-model animals. In studies related to the effects of 6g and 6n on emotional regulation, Bhatt et al. (2013, 2017a, 2017b) tested different types of anxiety disorder models, including lipopolysaccharide (LPS), chronic unpredictable mild stress (CUMS), and traumatic brain injury (TBI), using male Swiss Albino mice. Both 6g and 6n produced a significant reduction of anxiety-like responses on all three models measured by EPM, open-field test (OFT), and light-dark box test (LDT). Kurhe et al. (2017) also detected a significant reduction in anxiety level using QCM-4, a derivative of 6n, in a high-fat diet (HFD)-induced anxiety mouse model. Consistent with the above antagonistic experimental results, mice treated with mCPBG by intracerebroventricular injection to activate cerebral 5-HT<sub>3</sub>Rs produced a significant anxiety-like response in EPM. A significant reduction in the staying time and entering frequency in open arms was induced (Gupta et al., 2016), confirming the anxiolytic effects induced by antagonizing 5-HT<sub>3</sub>Rs from the opposite side. In recent years, genetic evidence for the regulation of anxiety responses by 5-HT<sub>3</sub>Rs



has also been found. On the one hand, when human serotonin type 3 receptor (*Htr3*), the primary gene coding 5-HT<sub>3</sub>Rs, is knocked out, mice showed a significant reduction in the base level of anxiety responses in several testing models including EPM and LDT (Bhatnagar et al., 2004). On the other hand, overexpression of 5-HT<sub>3</sub>Rs in the murine brain through targeted follicle-stimulating hormone (*FSH*) gene knock-out caused the experimental mice to exhibit anxiety-like behaviors (Bi et al., 2020). In conclusion, the above studies have confirmed that 5-HT<sub>3</sub>Rs have promoting effects on anxiety responses from both pharmacological and genetic aspects, i.e., activating 5-HT<sub>3</sub>Rs induces anxiety-like behaviors, whereas antagonizing 5-HT<sub>3</sub>Rs weakens the anxiety response or significantly lowers the base level of response to anxiety stimuli in normal individuals.

However, some studies did not support the boosting effects of 5-HT<sub>3</sub>R on the regulation of anxiety, as they found an anxiolytic-like effect of 5-HT<sub>3</sub>Rs on the modulation of anxiety. For example, by using transgenic mice with overexpressed 5-HT<sub>3</sub>Rs, Harrell and Allan (2003) observed significant reductions in the base level of anxiety in mice tested by EPM. Nowicki et al. (2014) also found that after the administration of ondansetron in the water environment of zebrafishes feeding for a period of time, their base level of anxiety response was significantly improved, indicating that 5-HT<sub>3</sub>Rs produced anxiolytic effects. These contradictory findings were even more pronounced when the effects of 5-HT<sub>3</sub>Rs were restricted to a specific brain region. Considering that 5-HT<sub>3</sub>Rs have extensive distribution in multiple brain regions of the CNS and the complexity of the role of the CNS in the regulation of anxiety (Koyama et al., 2017), the above contradictory findings likely arose because the regulation of the anxiety response by 5-HT<sub>3</sub>Rs involves many neural circuits and cellular mechanisms. Therefore, it is necessary to classify and discuss the neuropharmacological mechanisms of 5-HT<sub>3</sub>R action in different brain regions and neural circuits in relation to anxiety reactions.

#### 4 Mechanism of anxiety regulation in brain circuits by 5-HT<sub>3</sub>R

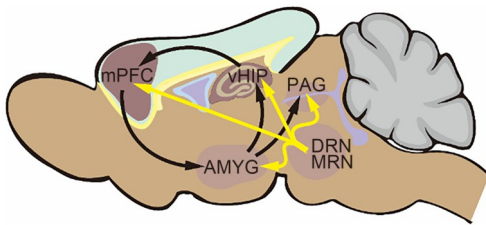
In the CNS, anxiety reactions involve complex neural mechanisms. Firstly, at the anatomical level,

anxiety reactions involve the complicated simultaneous activation of multiple brain regions, including the medial prefrontal cortex (mPFC), ventral HIP (vHIP), basolateral amygdala (BLA), central amygdaloid nucleus (CeA), bed nucleus of the stria terminalis (BNST), hypothalamus, periaqueductal gray (PAG), parabrachial nucleus (PBN), and other nearby regions that have been confirmed to respond to anxiety stimuli in a previous review (Tovote et al., 2015). Enormous neural networks are formed from connective projections between adjacent brain areas involved in the regulation of anxiety reactions. Meanwhile, the interconnections of adjacent brain areas always contain excitatory and inhibitory projections involved in anxiety reactions, giving some brain regions, like the AMYG and mPFC, both anxiogenic and anxiolytic behavioral effects (Tovote et al., 2015). Moreover, functionally inhibitory (largely GABAergic) interneurons in local brain areas also have efficient regulatory effects on both excitatory and inhibitory outputs, and thereby influence anxiety reactions with opposite results (Babaev et al., 2018). Therefore, to discuss the mechanism of 5-HT<sub>3</sub>Rs in regulating anxiety, the brain regions and neural circuits controlling anxiety reactions regulated by 5-HT<sub>3</sub>Rs should first be confirmed. Then, the regulatory role of 5-HT<sub>3</sub>Rs in each specific brain region should be discussed separately. Moreover, the inconsistent regulation of 5-HT<sub>3</sub>Rs in both excitatory and inhibitory projections, and interneurons executing contrary effects of anxiety reactions in some specific brain areas should also be considered.

##### 4.1 Regulatory brain circuits of anxiety reactions related to 5-HT<sub>3</sub>Rs

According to morphological studies by Koyama et al. (2017), the AMYG and neural circuits connecting with this brain region including the vHIP, mPFC, and PAG are considered vital targets of 5-HT<sub>3</sub>R in response to anxiety reactions. The functional activities of these brain areas in anxiety reactions include two main aspects. First, the AMYG receives environmental stimuli from sensory inputs or primary-process emotional signals from the mPFC, integrates and encodes aversive information to downstream effector regions in the brain stem such as the PAG, one of the brainstem structures vital in controlling the “fight to flight” reaction, and thereby generates avoidance and freezing. On the other hand, it sends excitatory

projections to the vHIP and subsequently activates pyramidal neurons projecting to the mPFC, and thus organizes AMYG-vHIP-mPFC neural circuits (Fig. 1) to precisely process and embellish anxiety information at the forebrain level. Serotonin projections, large resources of 5-HTs in the CNS derived from the DRN and MRN, have been clearly shown to generate connections to these brain regions. Therefore, 5-HT<sub>3</sub>R<sub>s</sub> can be directly activated by stimulation of 5-HTs released from terminals of these projections during anxiety reactions to give play to the regulatory roles of anxiety processing. In the following section, we discuss the mechanism of the regulation of anxiety by 5-HT<sub>3</sub>R<sub>s</sub> in the AMYG, vHIP, mPFC, and PAG.



**Fig. 1** Schematic diagram of main brain regions participating in the regulation of anxiety by 5-hydroxytryptamine (5-HT) type 3 receptors (5-HT<sub>3</sub>R<sub>s</sub>). The black arrows represent neuronal connections between brain regions. The yellow arrows represent 5-HT projections from the midbrain. mPFC, medial prefrontal cortex; vHIP, ventral hippocampus; PAG, periaqueductal gray; AMYG, amygdala; DRN, dorsal raphe nucleus; MRN, median raphe nucleus.

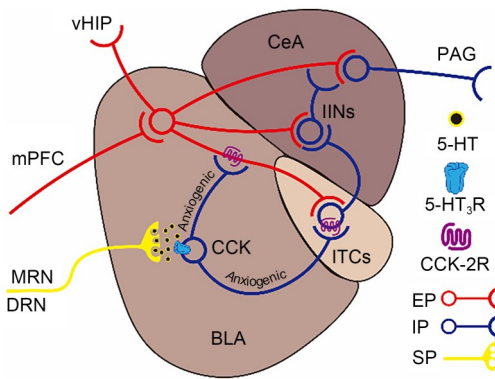
#### 4.2 Mechanism of 5-HT<sub>3</sub>R regulation of anxiety in the AMYG

The AMYG was one of the first brain regions found to be closely associated with anxiety reactions. Human clinical studies and analysis of immediate early genes in rodents have demonstrated a significant increase in AMYG activity under anxiety conditions. Pharmacological studies have also shown that damaging the bioactivity of AMYG neurons leads to a significant anxiolytic response in EPM of animals, which further confirms the important role of the AMYG in the regulation of the anxiety reaction (Sah, 2017). In clinical neuroimaging studies, the AMYG was considered to be closely associated with many types of anxiety disorder. In some magnetic resonance imaging (MRI) studies, abnormally increased volumes of the AMYG were observed in OCD and GAD patients (Szeszko et al., 1999; Millan, 2022). Unusually and selectively activated patterns of the AMYG were

also observed in SAD patients when exposed to fear or aversive conditions (Birbaumer et al., 1998). In PTSD, the AMYG was considered the center of neural circuits involved in the regulation of fear conditioning, while hyperactivation of the AMYG was confirmed by functional brain meta-analyses in adult PTSD (Rogan et al., 1997). In short, the AMYG plays a vital role in the regulation of anxiety. Its abnormal hyperactivation is a commonly observed clinical situation in many types of anxiety disorder.

The AMYG is mainly composed of the BLA and CeA, which establish the main neural pathway for the occurrence and maintenance of the anxiety response in the AMYG. The BLA is the vital region of acceptance of anxiogenic information in the AMYG. It can receive excitatory sensory inputs related to environmental changes and cue-related behaviors from cortical association areas like the mPFC, and then processes and integrates anxiety information on glutamatergic projections in this area with high efficiency. This information can be transmitted into GABAergic inhibitory projections of the CeA to inhibit the activity of downstream brain regions like the PAG and cause fear or disgust (Babaev et al., 2018). In the regulation of loop circuits in the AMYG, the projective glutamatergic neurons of the BLA send projections into IINs in the CeA and a population of intercalated cells (ITCs) that are also GABAergic interneurons located in an intercalated island, a region sandwiched between the BLA and CeA. IINs in the CeA project mainly into GABAergic projection neurons in this region that negatively regulate the excitatory projections derived from the BLA to the CeA. In contrast, ITCs between the BLA and CeA project mainly to IINs located in the CeA to action their feedforward inhibitory regulation. There are also some IINs located in the BLA. Their terminals can administer the projective glutamatergic neurons of the BLA-innervating ITCs and the cell bodies of ITCs in the intercalated island, to inhibit the activity of ITCs. Thus, a complex information transmission network (Fig. 2) of anxiety reactions is formed in the AMYG (Babaev et al., 2018).

In the feedback regulatory network of anxiety reactions in the AMYG, 5-HT<sub>3</sub>R<sub>s</sub> play their anxiety regulatory role mainly through the activation of the cholecystikinin (CCK) system. CCKs are important neuropeptides in the AMYG involved in promoting



**Fig. 2** Schematic paradigm of the regulation of anxiety by 5-hydroxytryptamine (5-HT) type 3 receptors (5-HT<sub>3</sub>R) in the amygdala (AMYG). The BLA receives excitatory inputs encoding anxiogenic information from cortical association areas like the mPFC, and then innervates four main excitatory projections: (1) innervates to the CeA inhibitory projecting neurons to induce inhibition of the CeA on downstream brain regions like the PAG and produces anxiogenic effects; (2) innervates to CeA IINs to inhibit BLA excitatory projections innervating CeA inhibitory projecting neurons and produces anxiolytic effects; (3) innervates to ITCs to inhibit CeA IINs and induces “disinhibitory” effects on BLA excitatory projections innervating CeA inhibitory projecting neurons to evoke anxiogenic effects; (4) innervates the vHIP to become involved in the organization of AMYG-vHIP-mPFC neural circuits. 5-HT<sub>3</sub>Rs are expressed mainly on the CCK positive IINs in the BLA innervated by 5-HT projections derived from the MRN and DRN, which have high efficiency to induce CCK release, thereby activating CCK-2Rs expressed on BLA excitatory projection fibers innervating ITCs and ITCs themselves, i.e., activating ITCs to promote anxiogenic effects. CeA, central amygdaloid nucleus; BLA, basolateral amygdala; mPFC, medial prefrontal cortex; PAG, periaqueductal gray; vHIP, ventral hippocampus; MRN, median raphe nucleus; DRN, dorsal raphe nucleus; CCK, cholecystokinin; CCK-2R, CCK-2 receptor; ITCs, intercalated cells; IINs, inhibitory interneurons; EP, excitatory projection; IP, inhibitory projection; SP, serotonin (5-HT) projection.

and maintaining anxiety responses. Injection of CCK analogs into the AMYG can induce anxiety-like behaviors or enhance anxiety responses in experimental animals, while antagonism of CCK function can significantly reduce anxiety responses (del Boca et al., 2012; Holm et al., 2014). CCK-positive neurons in the AMYG are mainly IINs co-expressed with GABA in the BLA. Immunofluorescence and in situ hybridization studies showed that 5-HT<sub>3</sub>Rs are densely expressed on the cell body and axon terminals of CCK-positive neurons. Meanwhile, these areas also receive 5-HT projections derived from the MRN and DRN

(Mascagni and McDonald, 2007). Consequently, these CCK-positive neurons can directly receive excitatory information from the raphe nucleus during anxiety reactions to activate 5-HT<sub>3</sub>Rs and thereby induce Ca<sup>2+</sup>-dependent release of CCKs (Uvnäs-Moberg et al., 1996). In the CNS, CCKs bind mainly to their specific CCK-2 receptors to play their biological role. CCK-2 receptors are widely distributed in the neurons innervated by IINs in the BLA, including BLA glutamatergic projection fibers innervating ITCs and ITCs themselves in the intercalated island. Thus, CCKs can further induce K<sup>+</sup>-dependent release of GABA at the postsynaptic level through a coordinated mechanism with glutamate receptors by activating CCK-2 receptors on these cells. This results in hyperpolarized potentials from BLA glutamatergic projections innervating ITCs as well as ITCs themselves and decreased excitability of these neurons observed in patch-clamp studies (de la Mora et al., 2007). These CCK system-mediated effects reduce the inhibitory regulation of ITCs on IINs in the CeA, and thus produce a “disinhibitory” effect on the excitatory projections of the BLA to the CeA, i.e., they indirectly enhance the neural transmission of this pathway. Therefore, in the neuronal micro-loop regulation of the AMYG, 5-HT<sub>3</sub>Rs inhibit the inhibitory regulation of ITCs on IINs in the CeA by promoting the release of CCKs and subsequently enhancing CCK-2 receptor-mediated GABA release, thus indirectly enhancing the excitatory projection of the BLA to CeA and promoting the occurrence and development of anxiety reactions (Fig. 2). In a pharmacological behavioral study in vivo, administration of tropisetron or ondansetron into the AMYG of male BKW mice resulted in a significant decrease in the level of anxiety reactions in LDT (Costall et al., 1989). Higgins et al. (1991) also found significant anxiolytic-like responses after injection of ondansetron, tropisetron, and MDL7222 into the AMYG of Lister Hooded rats, proving the efficacy of 5-HT<sub>3</sub>Rs in the promotion of anxiety reactions in the AMYG (Fig. 2). Considering the unidirectional contributory role of 5-HT<sub>3</sub>Rs to anxiety in the AMYG, together with abnormal hyperactivation of the AMYG represented in anxiety disorders, interception of the inhibition of 5-HT<sub>3</sub>Rs in anxiolytic pathways seems to be a relatively effective means to weaken pervasive classes of anxiety disorder, such as PTSD, SAD, GAD, and OCD. Thus, in future the AMYG could become a

hypothetically ideal target in anxiety treatment oriented toward clinical 5-HT<sub>3</sub>R antagonists.

However, it is still unclear if 5-HT<sub>3</sub>R<sub>s</sub> are expressed on IINs or ITCs to directly regulate excitatory anxiogenic projections from the BLA to the CeA, so the morphological and mechanistic aspects of 5-HT<sub>3</sub>R<sub>s</sub> in regulation of anxiety in IINs and ITCs of the AMYG remain to be further explored.

### 4.3 Mechanism of 5-HT<sub>3</sub>R regulation of anxiety in the mPFC

The neurons in the mPFC are involved mainly in processing attention and emotion-related information. Therefore, the mPFC is a critical brain area in the CNS to regulate the fear extinction response and aversive conditioning. In pathology, dysfunction of the mPFC is associated with PTSD. Compared to healthy subjects, PTSD patients show a decreased gray matter volume in the mPFC (Koch et al., 2017). OCD is also linked to the abnormal structure and function of the mPFC. In morphology, a voxel-based morphometry study revealed a significant reduction of gray matter volume in the bilateral mPFC of OCD patients compared with that of healthy subjects (Togao et al., 2010). The mPFC in rodents is composed mainly of the infralimbic cortex (IL) area and prelimbic cortex (PL) area, both of which have close fiber connections with the AMYG. However, they have completely opposite effects on the functional regulation of anxiety reactions. The PL mainly receives excitatory inputs from the vHIP and processes and transmits processed information to activate the downstream BLA, which can further enhance the BLA-CeA-mediated fear/disgust emotional response pathway and induce and promote anxiety reactions. By contrast, the IL contains a large number of ITCs that receive excitatory input from the PL and then send inhibitory projections to the BLA. This plays a regulatory role in the extinction of fear response by inducing a certain degree of anxiolytic effect (Motzkin et al., 2015; Park and Moghaddam, 2017). The utterly different regulatory effects of PL and IL on anxiety reactions lead to the entirely opposite results of 5-HT<sub>3</sub>R in the regulation of the emotional response in the mPFC (Fig. 3).

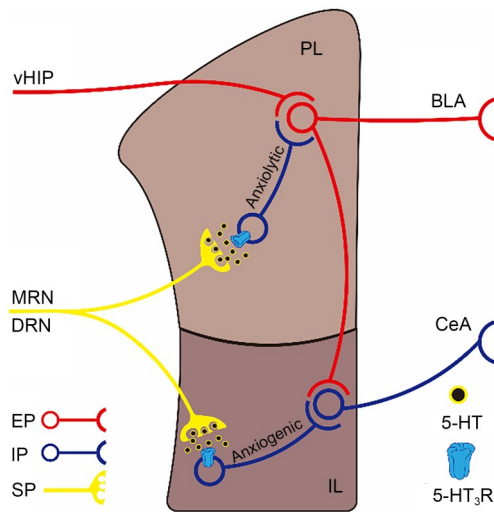
In situ hybridization studies showed that the positive signals of 5-HT<sub>3</sub>R mRNA were extensively co-expressed with GABAergic interneurons in the PL and IL. There was also a high 5-HT input innervated

by the DRN and MRN in these two regions. These findings show that 5-HT systems have the ability to directly modulate GABAergic inter-inhibitory neurons in both the PL and IL via 5-HT<sub>3</sub>R activation (Gui et al., 2010). By enhancing GABA release through activation of 5-HT<sub>3</sub>R, the inhibitory inputs of mPFC pyramidal neurons are enhanced, and thereby hyperpolarization is induced and excitability is reduced (Gui et al., 2010). These molecular effects were further confirmed in the study of Riga et al. (2016), in which administration of vortioxetine and ondansetron into the mPFC significantly increased the excitability of pyramidal neurons and the release of glutamate. These electrophysiological effects were blocked by SR57227A. From the above, corresponding to the opposite effects of the PL and IL on the regulation of anxiety reactions, activation of 5-HT<sub>3</sub>R distributed in the PL can produce anxiolytic effects, whereas activation of 5-HT<sub>3</sub>R distributed in the IL seems to promote anxiety (Fig. 3). Given the above evidence from animal experiments suggesting that 5-HT<sub>3</sub>R<sub>s</sub> expressed on the mPFC elicit bidirectional effects on anxiety, a precise clinical management is required to target mPFC 5-HT<sub>3</sub>R<sub>s</sub>, which seems difficult to achieve under current clinical systems. Thus, more basic information is still needed, especially of the difference in expression of 5-HT<sub>3</sub>R<sub>s</sub> between the PL and IL, and of the level of activation in these two areas under anxiety conditions, to guide the development of precise methods for treating anxiety disorders in clinical practice.

### 4.4 Mechanisms of 5-HT<sub>3</sub>R regulation of anxiety in the vHIP

The HIP is one of brain regions in the CNS that plays an important role in regulating both cognitive learning and emotional responses. It has been well demonstrated that dysfunctions of HIP are obviously linked to anxiety disorders, especially PTSD. Several clinical reports and neuroimaging meta-analyses have indicated that the hippocampal volume of chronic PTSD patients was smaller than that of healthy controls (Bremner et al., 1995, 2021). The HIP can be anatomically divided into the dorsal (dHIP) and ventral (vHIP) parts (Tseilikman et al., 2022). In relevant reviews, Bannerman et al. (2004) and Engin and Treit (2007) have pointed out that the dHIP may be more involved in the processing and disposal of a series of information inputs related to learning and memory,





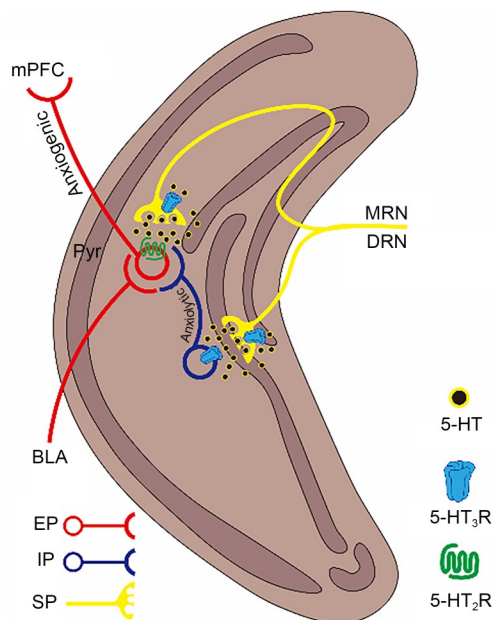
**Fig. 3** Schematic paradigm of the regulation of anxiety by 5-hydroxytryptamine (5-HT) type 3 receptors (5-HT<sub>3</sub>R) in the mPFC. The PL receives excitatory inputs processing anxiogenic information from the vHIP, and then innervates excitatory projecting neurons to the BLA, producing anxiogenic effects. Meanwhile, it also innervates excitatory collaterals to IL inhibitory projections innervating to the CeA inhibitory projecting neurons, to inhibit CeA inhibitory projecting neurons to produce anxiolytic effects. 5-HT<sub>3</sub>R are densely distributed in both the PL and IL IINs innervated by 5-HT projections derived from the MRN and DRN. Activating 5-HT<sub>3</sub>R expressed on PL IINs can promote GABA release to inhibit PL excitatory neurons projecting to the BLA, and thereby produces anxiolytic effects. In contrast, activating 5-HT<sub>3</sub>R expressed on IL IINs will inhibit IL inhibitory neurons projecting to the CeA through the same GABA mechanism, to induce anxiogenic effects. PL, prelimbic cortex; vHIP, ventral hippocampus; BLA, basolateral amygdala; IL, infralimbic cortex; CeA, central amygdaloid nucleus; IINs, intermediate inhibitory neurons; MRN, median raphe nucleus; DRN, dorsal raphe nucleus; GABA,  $\gamma$ -aminobutyric acid; EP, excitatory projection; IP, inhibitory projection; SP, serotonin (5-HT) projection.

while the vHIP may be specifically responsible for the regulation of anxiety-related functions. Therefore, the vHIP is a focus in current research on the mechanism of anxiety reactions. In the neural connections among brain areas during anxiety reactions, the vHIP can receive excitatory inputs from the BLA (Fig. 2), and processes fear or disgust information encoded by the BLA. Then the vHIP sends excitatory projections to the PL in the mPFC, which has downstream excitatory inputs to the BLA (Fig. 4). Thus, as an important relay for the transmission and processing of disgust/fear information, the vHIP has strong positive regulatory effects on anxiety reactions (Tovote et al., 2015).

Like the mPFC, 5-HT<sub>3</sub>R are also expressed mainly on IINs in the vHIP. Thus, activation of 5-HT<sub>3</sub>R in the vHIP leads to Ca<sup>2+</sup>-dependent GABA release from these interneurons, which increases inhibitory inputs and decreases excitability of pyramidal neurons (Fig. 4). Consequently, the excitatory inputs to the PL from the vHIP will be reduced, resulting in anxiolytic effects (Riga et al., 2016). A genetic study by Harrell and Allan (2003) confirmed this view. After overexpression of the gene encoding 5-HT<sub>3</sub>R in the HIP, mice showed greater exploratory behavior in response to stimuli in novel environments and a significant anxiolytic response in the EPM. Likewise, while the expression level of 5-HT<sub>3</sub>R was significantly reduced in the HIP through targeted knockout of neuroplastin 65 protein, mice exhibited a significant reduction in the time that they entered the central area in the OFT test, i.e., the knockout produced anxiety reactions (Li et al., 2019). This study confirmed the anxiolytic effects induced by 5-HT<sub>3</sub>R in the vHIP from the opposite side.

However, some pharmacological behavioral studies do not support the induction of anxiolytic effects by hippocampal 5-HT<sub>3</sub>R. Stefański et al. (1993) did not observe a significant occurrence of anxiety response in OFT tests after injection of ondansetron and tropisetron into the HIP of rats. Meanwhile, Vogel's test showed that rats produced anti-anxiety/depression-like effects after the same operations. Although no studies have directly investigated the cause of this contradiction, 5-HT<sub>3</sub>R-induced 5-HT release could be a possible explanation. In addition to IINs in vHIP local areas, 5-HT<sub>3</sub>R are also expressed on the terminals of 5-HT projections derived from the DRN and MRN to the vHIP. Activation of 5-HT<sub>3</sub>R expressed on these fiber terminals can promote the release of Ca<sup>2+</sup>-dependent 5-HTs (Martin et al., 1992). Although these effects could produce a certain degree of positive feedback on 5-HT<sub>3</sub>R, a large amount of 5-HT<sub>2</sub>R, other subtypes of 5-HT<sub>3</sub>R mediating the excitatory G protein pathway, are expressed in the cell body of pyramidal neurons in the vHIP and could also be positively activated by the release of Ca<sup>2+</sup>-dependent 5-HTs promoted by 5-HT<sub>3</sub>R (Mendelson and McEwen, 1991). If this positive feedback loop is continuously activated, the release of 5-HTs will escalate and excitatory effects on pyramidal neurons mediated by 5-HT<sub>2</sub>R may become dominant. Thus, the rapid increase

in the excitability of pyramidal neurons mediated by 5-HT<sub>2</sub>Rs will positively promote the vHIP-PL-BLA anxiety regulatory pathway and promote the occurrence and development of an anxiety response (Alves et al., 2004). The anxiolytic effects of 5-HT<sub>3</sub>Rs on GABA release in the vHIP may be counterbalanced or even reversed by activation of 5-HT<sub>2</sub>Rs, and thereby produce anxiogenic effects (Fig. 4). In summary, the vHIP might be another target of bidirectional effects of 5-HT<sub>3</sub>Rs involved on anxiety. However, unlike the mPFC, the 5-HT<sub>3</sub>Rs-involved bidirectional effects on anxiety in the vHIP are presumably attributed to the

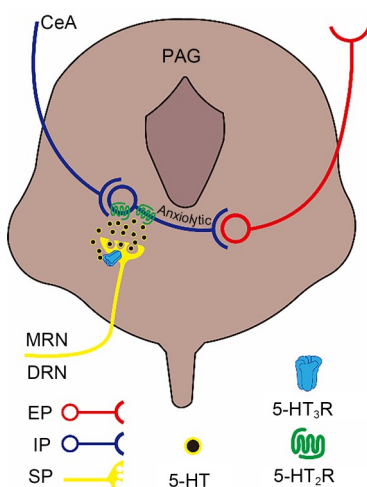


**Fig. 4** Schematic paradigm of the regulation of anxiety by 5-hydroxytryptamine (5-HT) type 3 receptors (5-HT<sub>3</sub>Rs) in the vHIP. The vHIP pyramidal neurons receive excitatory inputs encoding anxiogenic information derived from the BLA, and then send excitatory projections to the PL in the mPFC, thereby becoming involved in the establishment of the AMYG-vHIP-mPFC circuit to promote anxiety reactions. 5-HT<sub>3</sub>Rs are expressed on vHIP IINs and presynaptic terminals of 5-HT projections. Activation of 5-HT<sub>3</sub>Rs expressed on vHIP IINs can promote GABA release to inhibit pyramidal neurons to produce anxiolytic effects. In contrast, activation of 5-HT<sub>3</sub>Rs expressed on presynaptic terminals of 5-HT projections can secondarily promote 5-HT<sub>2</sub>Rs expressed on pyramidal neurons, thereby facilitating the AMYG-vHIP-mPFC circuit to promote anxiety. vHIP, ventral hippocampus; BLA, basolateral amygdala; PL, prelimbic cortex; mPFC, medial prefrontal cortex; IINs, intermediate inhibitory neurons; GABA,  $\gamma$ -aminobutyric acid; AMYG, amygdala; Pyr, pyramidal neuron; MRN, median raphe nucleus; DRN, dorsal raphe nucleus; EP, excitatory projection; IP, inhibitory projection; SP, serotonin (5-HT) projection.

interaction between 5-HT<sub>3</sub>Rs and 5-HT<sub>2</sub>Rs, whereas 5-HT<sub>3</sub>Rs themselves tend to unidirectionally contribute to anxiolysis in the local circuit. Because of the relatively close relationship between the HIP and PTSD, an effective therapeutic strategy targeting 5-HT<sub>3</sub>Rs in the vHIP against PTSD might be a combination of the 5-HT<sub>3</sub>R agonist with the antagonist 5-HT<sub>2</sub>R.

#### 4.5 Mechanism of 5-HT<sub>3</sub>R regulation of anxiety in the PAG

PAG is a gray matter structure surrounding the midbrain aqueduct. In addition to being involved in the transmission and regulation of pain information, PAG has positive regulatory effects on anxiety reactions. In PAG, IINs receive inhibitory projections from the CeA and have a tonic inhibitory effect on excitatory neurons (mostly vesicular glutamate transporter 2 (vGluT2)-positive neurons) in this region (Tovote et al., 2016). Since vGluT2-positive neurons have the ability to promote anxiety reactions (Taylor et al., 2019), the inhibitory projections derived from the CeA to PAG can inhibit the IINs in PAG to “disinhibit” vGluT2-positive neurons, thereby inducing anxiety reactions (Tovote et al., 2016). Whether 5-HT<sub>3</sub>Rs are expressed on IINs in PAG is unknown. Behavioral tests showed that administration of mCPPB into PAG did not cause significant changes in the anxiety level of mice during EPM (Lopes et al., 2022). Thus, the 5-HT<sub>3</sub>Rs may not directly regulate the anxiety reactions in PAG. However, 5-HT<sub>3</sub>R antagonists could significantly inhibit the anxiolytic effect mediated by the 5-HT<sub>2</sub>R agonist 1-(3-chlorophenyl)piperazine (mCPP) (Lopes et al., 2022). The 5-HT<sub>2</sub>Rs are expressed mainly on the soma and dendrites of IINs throughout the PAG, induce GABA release from IINs, and antagonize the action of vGluT2-positive neurons to produce an anxiolytic reaction (Griffiths and Lovick, 2002; Tovote et al., 2016; Vilela-Costa et al., 2021). Meanwhile, 5-HT<sub>3</sub>Rs have a similar distribution of 5-HT projection terminals (from the DRN and MRN) in PAG as in the vHIP (Lopes et al., 2022). Therefore, 5-HT<sub>3</sub>Rs may be involved in anxiolytic responses by enhancing Ca<sup>2+</sup>-dependent 5-HT release in PAG, thereby activating 5-HT<sub>2</sub>Rs and promoting GABA release to induce GABA-controlled anxiolytic responses (Fig. 5). In pathology, PAG is closely associated with PD. Electrical stimulation of the brain using stereotactic neurosurgery in the PAG of humans can induce intense



**Fig. 5** Schematic paradigm of the regulation of anxiety by 5-hydroxytryptamine (5-HT) type 3 receptors (5-HT<sub>3</sub>Rs) in the PAG. PAG IINs receive inhibitory inputs from the CeA to relieve tonic inhibition of their downstream excitatory projections, thereby becoming positively involved in anxiety reactions through a “disinhibition” mechanism. 5-HT<sub>3</sub>Rs are expressed mainly on presynaptic terminals of 5-HT projections here, and efficiently promote activation of 5-HT<sub>2</sub>Rs located in PAG IINs when activated. Activation of 5-HT<sub>2</sub>Rs promotes the inhibition of downstream excitatory projections to produce anxiolytic effects. PAG, periaqueductal gray; IINs, intermediate inhibitory neurons; CeA, central amygdaloid nucleus; MRN, median raphe nucleus; DRN, dorsal raphe nucleus; EP, excitatory projection; IP, inhibitory projection; SP, serotonin (5-HT) projection.

states of fear and/or terror (Zwanzger et al., 2012). Further research is needed to provide a theoretical foundation for exploring the prospect of a potential application to PD treatment targeting 5-HT<sub>3</sub>Rs.

## 5 Conclusions

5-HT<sub>3</sub>Rs have bidirectional regulation effects on anxiety reactions due to the various characteristics of their distribution in different brain regions and cell types in the CNS. In the AMYG, 5-HT<sub>3</sub>R-mediated release of CCK and subsequent “disinhibition” of the BLA-CeA pathway are essential mechanisms for 5-HT<sub>3</sub>R-induced promotion of anxiety reactions. In contrast, 5-HT<sub>3</sub>R-mediated GABA release and inhibition of the vHIP-PL/PL-BLA pathway are the main mechanism responsible for anxiogenic effects in the vHIP and PL, and are also efficient in promoting anxiety in the IL. Besides, the positive regulation of 5-HT release by 5-HT<sub>3</sub>R in the vHIP and PAG, and subsequent

enhancement of 5-HT<sub>2</sub>R functional activity suggest that 5-HT<sub>3</sub>Rs may regulate anxiety reactions through the action of 5-HT<sub>2</sub>Rs, facilitating anxiety in the vHIP and generating anxiogenic effects in PAG. There is evidence that certain anxiety subtypes impact mainly distinct local brain circuitry, which suggests that systemic intervention with 5-HT<sub>3</sub>Rs would have therapeutic potential. For instance, 5-HT<sub>3</sub>R antagonists would be more efficient for GAD and SAD for targeting mainly in the AMYG, whereas 5-HT<sub>3</sub>R agonists could be auxiliaries in PD therapy in view of the anxiolytic effects of 5-HT<sub>3</sub>Rs on PAG. However, the circuit specific and bidirectional regulation of 5-HT<sub>3</sub>Rs in controlling emotions makes it difficult to directly apply 5-HT<sub>3</sub>R agonists or antagonists to generally treat or prevent anxiety. Therefore, exploiting circuit targeting methods and applying a combined medication strategy might be effectual in increasing the therapeutic values of 5-HT<sub>3</sub>Rs on anxiety disorders involving multiple regions of the brain. Furthermore, the expression of 5-HT<sub>3</sub>Rs on a specific subtype of neuron in different brain regions is not yet clear, and whether 5-HT<sub>3</sub>Rs have interactions with other 5-HTR subtypes involved in anxiety is also unknown, and remains to be addressed in future.

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## Author contributions

Yinan DU: conceptualization, writing – original draft, writing – review & editing, and project administration. Zhiwei LI: investigation and visualization. Yukui ZHAO: investigation and visualization. Jing HAN: writing – review & editing and funding acquisition. Weiping HU: supervision and funding acquisition. Zhiqiang LIU: supervision, writing – review & editing, project administration, and funding acquisition. All authors have read and approved the final version.

## Compliance with ethics guidelines

Yinan DU, Zhiwei LI, Yukui ZHAO, Jing HAN, Weiping HU, and Zhiqiang LIU declare that they have no conflict of interest.

This review does not contain any studies with human or animal subjects performed by any of the authors.



## References

- Abela AR, Browne CJ, Sargin D, et al., 2020. Median raphe serotonin neurons promote anxiety-like behavior via inputs to the dorsal hippocampus. *Neuropharmacology*, 168: 107985.  
<https://doi.org/10.1016/j.neuropharm.2020.107985>
- al Kury LT, Mahgoub M, Howarth FC, et al., 2018. Natural negative allosteric modulators of 5-HT<sub>3</sub> receptors. *Molecules*, 23(12):3186.  
<https://doi.org/10.3390/molecules23123186>
- Alves SH, Pinheiro G, Motta V, et al., 2004. Anxiogenic effects in the rat elevated plus-maze of 5-HT<sub>2C</sub> agonists into ventral but not dorsal hippocampus. *Behav Pharmacol*, 15(1):37-43.  
<https://doi.org/10.1097/00008877-200402000-00005>
- Andrade TGCS, Macedo CEA, Zangrossi H, et al., 2004. Anxiolytic-like effects of median raphe nucleus lesion in the elevated T-maze. *Behav Brain Res*, 153(1):55-60.  
<https://doi.org/10.1016/j.bbr.2003.10.036>
- Artaiz I, Romero G, Zazpe A, et al., 1995. The pharmacology of VA<sub>21</sub>B<sub>3</sub>: an atypical 5-HT<sub>3</sub> receptor antagonist with anxiolytic-like properties in animal models. *Psychopharmacology*, 117(2):137-148.  
<https://doi.org/10.1007/bf02245179>
- Askari N, Moin M, Sanati M, et al., 2012. Granisetron adjunct to fluvoxamine for moderate to severe obsessive-compulsive disorder: a randomized, double-blind, placebo-controlled trial. *CNS Drugs*, 26(10):883-892.  
<https://doi.org/10.2165/11635850-000000000-00000>
- Babaev O, Piletti Chatain C, Krueger-Burg D, 2018. Inhibition in the amygdala anxiety circuitry. *Exp Mol Med*, 50(4): 1-16.  
<https://doi.org/10.1038/s12276-018-0063-8>
- Bannerman DM, Matthews P, Deacon RM, et al., 2004. Medial septal lesions mimic effects of both selective dorsal and ventral hippocampal lesions. *Behav Neurosci*, 118(5): 1033-1041.  
<https://doi.org/10.1037/0735-7044.118.5.1033>
- Barnes JM, Barnes NM, Costall B, et al., 1990. Identification and distribution of 5-HT<sub>3</sub> recognition sites within the human brainstem. *Neurosci Lett*, 111(1-2):80-86.  
[https://doi.org/10.1016/0304-3940\(90\)90348-d](https://doi.org/10.1016/0304-3940(90)90348-d)
- Barnes NM, Hales TG, Lummis SCR, et al., 2009. The 5-HT<sub>3</sub> receptor – the relationship between structure and function. *Neuropharmacology*, 56(1):273-284.  
<https://doi.org/10.1016/j.neuropharm.2008.08.003>
- Basak S, Gicheru Y, Samanta A, et al., 2018. Cryo-EM structure of 5-HT<sub>3A</sub> receptor in its resting conformation. *Nat Commun*, 9:514.  
<https://doi.org/10.1038/s41467-018-02997-4>
- Bhatnagar S, Nowak N, Babich L, et al., 2004. Deletion of the 5-HT<sub>3</sub> receptor differentially affects behavior of males and females in the Porsolt forced swim and defensive withdrawal tests. *Behav Brain Res*, 153(2):527-535.  
<https://doi.org/10.1016/j.bbr.2004.01.018>
- Bhatt S, Mahesh R, Devadoss T, et al., 2013. Anxiolytic-like effect of (4-benzylpiperazin-1-yl)(3-methoxyquinoxalin-2-yl)methanone (6g) in experimental mouse models of anxiety. *Indian J Pharmacol*, 45(3):248-251.  
<https://doi.org/10.4103/0253-7613.111923>
- Bhatt S, Mahesh R, Jindal A, et al., 2017a. Neuropharmacological and neurochemical evaluation of N-n-propyl-3-ethoxyquinoxaline-2-carboxamide (6n): a novel serotonergic 5-HT<sub>3</sub> receptor antagonist for co-morbid antidepressant- and anxiolytic-like potential using traumatic brain injury model in rats. *J Basic Clin Physiol Pharmacol*, 28(2):93-100.  
<https://doi.org/10.1515/jbcpp-2016-0057>
- Bhatt S, Mahesh R, Devadoss T, et al., 2017b. Neuropharmacological evaluation of a novel 5-HT<sub>3</sub> receptor antagonist (4-benzylpiperazin-1-yl)(3-methoxyquinoxalin-2-yl)methanone (6g) on lipopolysaccharide-induced anxiety models in mice. *J Basic Clin Physiol Pharmacol*, 28(2): 101-106.  
<https://doi.org/10.1515/jbcpp-2016-0083>
- Bhatt S, Devadoss T, Manjula SN, et al., 2021. 5-HT<sub>3</sub> receptor antagonism: a potential therapeutic approach for the treatment of depression and other disorders. *Curr Neuropharmacol*, 19(9):1545-1559.  
<https://doi.org/10.2174/1570159x18666201015155816>
- Bi WK, Luan SS, Wang J, et al., 2020. FSH signaling is involved in affective disorders. *Biochem Biophys Res Commun*, 525(4):915-920.  
<https://doi.org/10.1016/j.bbrc.2020.03.039>
- Birbaumer N, Grodd W, Diedrich O, et al., 1998. fMRI reveals amygdala activation to human faces in social phobics. *NeuroReport*, 9(6):1223-1226.  
<https://doi.org/10.1097/00001756-199804200-00048>
- Bremner JD, Randall P, Scott TM, et al., 1995. MRI-based measurement of hippocampal volume in patients with combat-related posttraumatic stress disorder. *Am J Psychiatry*, 152(7):973-981.  
<https://doi.org/10.1176/ajp.152.7.973>
- Bremner JD, Hoffman M, Afzal N, et al., 2021. The environment contributes more than genetics to smaller hippocampal volume in Posttraumatic Stress Disorder (PTSD). *J Psychiatr Res*, 137:579-588.  
<https://doi.org/10.1016/j.jpsychires.2020.10.042>
- Campbell AD, Womer DE, Simon JR, 1995. The 5-HT<sub>3</sub> receptor agonist 1-(m-chlorophenyl)-biguanide interacts with the dopamine transporter in rat brain synaptosomes. *Eur J Pharmacol: Mol Pharmacol*, 290(2):157-162.  
[https://doi.org/10.1016/0922-4106\(95\)90029-2](https://doi.org/10.1016/0922-4106(95)90029-2)
- Celli J, Rappold G, Niesler B, 2017. The human serotonin type 3 receptor gene (*HTR3A-E*) allelic variant database. *Hum Mutat*, 38(2):137-147.  
<https://doi.org/10.1002/humu.23136>
- Challis C, Beck SG, Berton O, 2014. Optogenetic modulation of descending prefrontocortical inputs to the dorsal raphe bidirectionally bias socioaffective choices after social defeat. *Front Behav Neurosci*, 8:43.  
<https://doi.org/10.3389/fnbeh.2014.00043>
- Choi IS, Cho JH, Kim JT, et al., 2007. Serotonergic modulation of GABAergic synaptic transmission in developing rat CA3 pyramidal neurons. *J Neurochem*, 103(6):2342-2353.  
<https://doi.org/10.1111/j.1471-4159.2007.04945.x>
- Cortes-Altamirano JL, Olmos-Hernandez A, Jaime HB, et al.,



2018. Review: 5-HT<sub>1</sub>, 5-HT<sub>2</sub>, 5-HT<sub>3</sub> and 5-HT<sub>7</sub> receptors and their role in the modulation of pain response in the central nervous system. *Curr Neuroparmacol*, 16(2): 210-221.  
<https://doi.org/10.2174/1570159x15666170911121027>
- Costall B, Kelly ME, Naylor RJ, et al., 1989. Neuroanatomical sites of action of 5-HT<sub>3</sub> receptor agonist and antagonists for alteration of aversive behaviour in the mouse. *Br J Pharmacol*, 96(2):325-332.  
<https://doi.org/10.1111/j.1476-5381.1989.tb11821.x>
- Crocq MA, 2015. A history of anxiety: from hippocrates to DSM. *Dialogues Clin Neurosci*, 17(3):319-325.  
<https://doi.org/10.31887/DCNS.2015.17.3/macrocq>
- Cutler MG, 1991. An ethological study of the effects of buspirone and the 5-HT<sub>3</sub> receptor antagonist, BRL 43694 (granisetron) on behaviour during social interactions in female and male mice. *Neuropharmacology*, 30(4):299-306.  
[https://doi.org/10.1016/0028-3908\(91\)90053-e](https://doi.org/10.1016/0028-3908(91)90053-e)
- de la Mora MP, Hernández-Gómez AM, Arizmendi-García Y, et al., 2007. Role of the amygdaloid cholecystokinin (CCK)/gastrin-2 receptors and terminal networks in the modulation of anxiety in the rat. Effects of CCK-4 and CCK-8S on anxiety-like behaviour and [<sup>3</sup>H]GABA release. *Eur J Neurosci*, 26(12):3614-3630.  
<https://doi.org/10.1111/j.1460-9568.2007.05963.x>
- del Boca C, Lutz PE, le Merrer J, et al., 2012. Cholecystokinin knock-down in the basolateral amygdala has anxiolytic and antidepressant-like effects in mice. *Neuroscience*, 218:185-195.  
<https://doi.org/10.1016/j.neuroscience.2012.05.022>
- el Nebrisi E, Prytkova T, Lorke DE, et al., 2020. Capsaicin is a negative allosteric modulator of the 5-HT<sub>3</sub> receptor. *Front Pharmacol*, 11:1274.  
<https://doi.org/10.3389/fphar.2020.01274>
- Engin E, Treit D, 2007. The role of hippocampus in anxiety: intracerebral infusion studies. *Behav Pharmacol*, 18(5-6): 365-374.  
<https://doi.org/10.1097/FBP.0b013e3282de7929>
- Fawley JA, Doyle MW, Andresen MC, 2019. 5-HT<sub>3</sub>R-sourced calcium enhances glutamate release from a distinct vesicle pool. *Brain Res*, 1721:146346.  
<https://doi.org/10.1016/j.brainres.2019.146346>
- Gasiorek A, Trattinig SM, Ahring PK, et al., 2016. Delineation of the functional properties and the mechanism of action of TMPPAA, an allosteric agonist and positive allosteric modulator of 5-HT<sub>3</sub> receptors. *Biochem Pharmacol*, 110:111-92-108.  
<https://doi.org/10.1016/j.bcp.2016.04.004>
- Giacobbe P, Flint A, 2018. Diagnosis and management of anxiety disorders. *Continuum (Minneapolis)*, 24(3):893-919.  
<https://doi.org/10.1212/con.0000000000000607>
- Gibbs E, Chakrapani S, 2021. Structure, function and physiology of 5-hydroxytryptamine receptors subtype 3. In: Harris JR, Marles-Wright J (Eds.), *Macromolecular Protein Complexes III: Structure and Function*. Springer, Cham, p.373-408.  
[https://doi.org/10.1007/978-3-030-58971-4\\_11](https://doi.org/10.1007/978-3-030-58971-4_11)
- Giordano J, Gerstmann H, 2004. Patterns of serotonin- and 2-methylserotonin-induced pain may reflect 5-HT<sub>3</sub> receptor sensitization. *Eur J Pharmacol*, 483(2-3):267-269.  
<https://doi.org/10.1016/j.ejphar.2003.10.044>
- Griffiths JL, Lovick TA, 2002. Co-localization of 5-HT<sub>2A</sub>-receptor- and GABA-immunoreactivity in neurones in the periaqueductal grey matter of the rat. *Neurosci Lett*, 326(3):151-154.  
[https://doi.org/10.1016/s0304-3940\(02\)00182-9](https://doi.org/10.1016/s0304-3940(02)00182-9)
- Gui ZH, Zhang QJ, Liu J, et al., 2010. *In vivo* modulation of the firing activity of putative slow- and fast-spiking interneurons in the medial prefrontal cortex by 5-HT<sub>3</sub> receptors in 6-hydroxydopamine-induced Parkinsonian rats. *Neuroscience*, 169(3):1315-1325.  
<https://doi.org/10.1016/j.neuroscience.2010.05.059>
- Gupta D, Thangaraj D, Radhakrishnan M, 2016. A novel 5HT<sub>3</sub> antagonist 4i (N-(3-chloro-2-methylphenyl)quinoxalin-2-carboxamide) prevents diabetes-induced depressive phenotypes in mice: modulation of serotonergic system. *Behav Brain Res*, 297:41-50.  
<https://doi.org/10.1016/j.bbr.2015.10.007>
- Harmer CJ, Reid CB, Ray MK, et al., 2006. 5HT<sub>3</sub> antagonism abolishes the emotion potentiated startle effect in humans. *Psychopharmacology*, 186:18-24.  
<https://doi.org/10.1007/s00213-006-0337-z>
- Harrell AV, Allan AM, 2003. Improvements in hippocampal-dependent learning and decremental attention in 5-HT<sub>3</sub> receptor overexpressing mice. *Learn Mem*, 10(5):410-419.  
<https://doi.org/10.1101/lm.56103>
- Haus U, Varga B, Stratz T, et al., 2000. Oral treatment of fibromyalgia with tropisetron given over 28 days: influence on functional and vegetative symptoms, psychometric parameters and pain. *Scand J Rheumatol*, 29(113): 55-58.  
<https://doi.org/10.1080/030097400446652>
- Hensler JG, Hodge CW, Overstreet DH, 2004. Reduced 5-HT<sub>3</sub> receptor binding and lower baseline plus maze anxiety in the alcohol-preferring inbred fawn-hooded rat. *Pharmacol Biochem Behav*, 77(2):281-289.  
<https://doi.org/10.1016/j.pbb.2003.11.015>
- Hewlett WA, Schmid SP, Salomon RM, 2003. Pilot trial of ondansetron in the treatment of 8 patients with obsessive-compulsive disorder. *J Clin Psychiatry*, 64(9):1025-1030.  
<https://doi.org/10.4088/jcp.v64n0907>
- Higgins GA, Jones BJ, Oakley NR, et al., 1991. Evidence that the amygdala is involved in the disinhibitory effects of 5-HT<sub>3</sub> receptor antagonists. *Psychopharmacology*, 104(4): 545-551.  
<https://doi.org/10.1007/bf02245664>
- Holm L, Liang W, Thorsell A, et al., 2014. Acute effects on brain cholecystokinin-like concentration and anxiety-like behaviour in the female rat upon a single injection of 17β-estradiol. *Pharmacol Biochem Behav*, 122:222-227.  
<https://doi.org/10.1016/j.pbb.2014.04.004>
- Hutchison SM, Mâsse LC, Pawluski JL, et al., 2021. Perinatal selective serotonin reuptake inhibitor (SSRI) and other antidepressant exposure effects on anxiety and depressive behaviors in offspring: a review of findings in humans and rodent models. *Reprod Toxicol*, 99:80-95.  
<https://doi.org/10.1016/j.reprotox.2020.11.013>

- Irving H, Turek I, Kettle C, et al., 2021. Tapping into 5-HT<sub>3</sub> receptors to modify metabolic and immune responses. *Int J Mol Sci*, 22(21):11910.  
<https://doi.org/10.3390/ijms22111910>
- Juza R, Vlcek P, Mezeiova E, et al., 2020. Recent advances with 5-HT<sub>3</sub> modulators for neuropsychiatric and gastrointestinal disorders. *Med Res Rev*, 40(5):1593-1678.  
<https://doi.org/10.1002/med.21666>
- Kilpatrick GJ, Jones BJ, Tyers MB, 1987. Identification and distribution of 5-HT<sub>3</sub> receptors in rat brain using radioligand binding. *Nature*, 330(6150):746-748.  
<https://doi.org/10.1038/330746a0>
- Kilpatrick GJ, Butler A, Burridge J, et al., 1990. 1-(m-Chlorophenyl)-biguanide, a potent high affinity 5-HT<sub>3</sub> receptor agonist. *Eur J Pharmacol*, 182(1):193-197.  
[https://doi.org/10.1016/0014-2999\(90\)90513-6](https://doi.org/10.1016/0014-2999(90)90513-6)
- Koch SBJ, van Zuiden M, Nawijn L, et al., 2017. Decreased uncinate fasciculus tract integrity in male and female patients with PTSD: a diffusion tensor imaging study. *J Psychiatry Neurosci*, 42(5):331-342.  
<https://doi.org/10.1503/jpn.160129>
- Koyama Y, Kondo M, Shimada S, 2017. Building a 5-HT<sub>3A</sub> receptor expression map in the mouse brain. *Sci Rep*, 7:42884.  
<https://doi.org/10.1038/srep42884>
- Kurhe Y, Mahesh R, Devadoss T, 2015. QCM-4, a 5-HT<sub>3</sub> receptor antagonist ameliorates plasma HPA axis hyperactivity, leptin resistance and brain oxidative stress in depression and anxiety-like behavior in obese mice. *Biochem Biophys Res Commun*, 456(1):74-79.  
<https://doi.org/10.1016/j.bbrc.2014.11.036>
- Kurhe Y, Mahesh R, Devadoss T, 2017. Novel 5-HT<sub>3</sub> receptor antagonist QCM-4 attenuates depressive-like phenotype associated with obesity in high-fat-diet-fed mice. *Psychopharmacology*, 234(7):1165-1179.  
<https://doi.org/10.1007/s00213-017-4558-0>
- Lakhtakia T, Torous J, 2022. Current directions in digital interventions for mood and anxiety disorders. *Curr Opin Psychiatry*, 35(2):130-135.  
<https://doi.org/10.1097/ycp.0000000000000772>
- Leclercq Y, Puech AJ, Azcona A, et al., 1993. A randomized double-blind placebo-controlled study of tropisetron in the treatment of outpatients with generalized anxiety disorder. *Psychopharmacology*, 112:129-133.  
<https://doi.org/10.1007/bf02247373>
- Li HH, Liu YT, Gao XQ, et al., 2019. Neuroplastin 65 modulates anxiety- and depression-like behavior likely through adult hippocampal neurogenesis and central 5-HT activity. *FEBS J*, 286(17):3401-3415.  
<https://doi.org/10.1111/febs.14865>
- Lopes LT, Canto-de-Souza L, Baptista-de-Souza D, et al., 2022. The interplay between 5-HT<sub>2C</sub> and 5-HT<sub>3A</sub> receptors in the dorsal periaqueductal gray mediates anxiety-like behavior in mice. *Behav Brain Res*, 417:113588.  
<https://doi.org/10.1016/j.bbr.2021.113588>
- Lummis SCR, 2012. 5-HT<sub>3</sub> receptors. *J Biol Chem*, 287(48):40239-40245.  
<https://doi.org/10.1074/jbc.R112.406496>
- Machu TK, 2011. Therapeutics of 5-HT<sub>3</sub> receptor antagonists: current uses and future directions. *Pharmacol Ther*, 130(3):338-347.  
<https://doi.org/10.1016/j.pharmthera.2011.02.003>
- Maksay G, Biró T, Bugovics G, 2005. Allosteric modulation of 5-HT<sub>3</sub> serotonin receptors. *Eur J Pharmacol*, 514(1):17-24.  
<https://doi.org/10.1016/j.ejphar.2005.03.019>
- Marcinkiewicz CA, Mazzone CM, D'Agostino G, et al., 2016. Serotonin engages an anxiety and fear-promoting circuit in the extended amygdala. *Nature*, 537(7618):97-101.  
<https://doi.org/10.1038/nature19318>
- Martin KF, Hannon S, Phillips I, et al., 1992. Opposing roles for 5-HT<sub>1B</sub> and 5-HT<sub>3</sub> receptors in the control of 5-HT release in rat hippocampus *in vivo*. *Br J Pharmacol*, 106(1):139-142.  
<https://doi.org/10.1111/j.1476-5381.1992.tb14306.x>
- Mascagni F, McDonald AJ, 2007. A novel subpopulation of 5-HT type 3A receptor subunit immunoreactive interneurons in the rat basolateral amygdala. *Neuroscience*, 144(3):1015-1024.  
<https://doi.org/10.1016/j.neuroscience.2006.10.044>
- McCann UD, Morgan CM, Geraci M, et al., 1997. Effects of the 5-HT<sub>3</sub> antagonist, ondansetron, on the behavioral and physiological effects of pentagastrin in patients with panic disorder and social phobia. *Neuropsychopharmacology*, 17(6):360-369.  
[https://doi.org/10.1016/s0893-133x\(97\)00085-7](https://doi.org/10.1016/s0893-133x(97)00085-7)
- Mendelson SD, McEwen BS, 1991. Autoradiographic analyses of the effects of restraint-induced stress on 5-HT<sub>1A</sub>, 5-HT<sub>1C</sub> and 5-HT<sub>2</sub> receptors in the dorsal hippocampus of male and female rats. *Neuroendocrinology*, 54(5):454-461.  
<https://doi.org/10.1159/000125951>
- Millan MJ, 2022. Agomelatine for the treatment of generalized anxiety disorder: focus on its distinctive mechanism of action. *Ther Adv Psychopharmacol*, 12:20451253221105128.  
<https://doi.org/10.1177/20451253221105128>
- Motzkin JC, Philippi CL, Wolf RC, et al., 2015. Ventromedial prefrontal cortex is critical for the regulation of amygdala activity in humans. *Biol Psychiatry*, 77(3):276-284.  
<https://doi.org/10.1016/j.biopsych.2014.02.014>
- Murphy SE, Capitão LP, Giles SLC, et al., 2021. The knowns and unknowns of SSRI treatment in young people with depression and anxiety: efficacy, predictors, and mechanisms of action. *Lancet Psychiatry*, 8(9):824-835.  
[https://doi.org/10.1016/s2215-0366\(21\)00154-1](https://doi.org/10.1016/s2215-0366(21)00154-1)
- Nowicki M, Tran S, Muraleetharan A, et al., 2014. Serotonin antagonists induce anxiolytic and anxiogenic-like behavior in zebrafish in a receptor-subtype dependent manner. *Pharmacol Biochem Behav*, 126:170-180.  
<https://doi.org/10.1016/j.pbb.2014.09.022>
- Pallanti S, Bernardi S, Antonini S, et al., 2009. Ondansetron augmentation in treatment-resistant obsessive-compulsive disorder: a preliminary, single-blind, prospective study. *CNS Drugs*, 23(12):1047-1055.  
<https://doi.org/10.2165/11530240-000000000-00000>
- Park J, Moghaddam B, 2017. Impact of anxiety on prefrontal cortex encoding of cognitive flexibility. *Neuroscience*, 345:193-202.  
<https://doi.org/10.1016/j.neuroscience.2016.06.013>

- Riga MS, Sánchez C, Celada P, et al., 2016. Involvement of 5-HT<sub>3</sub> receptors in the action of vortioxetine in rat brain: focus on glutamatergic and GABAergic neurotransmission. *Neuropharmacology*, 108:73-81. <https://doi.org/10.1016/j.neuropharm.2016.04.023>
- Rogan MT, Stäubli UV, Ledoux JE, 1997. Fear conditioning induces associative long-term potentiation in the amygdala. *Nature*, 390(6660):604-607. <https://doi.org/10.1038/37601>
- Sah P, 2017. Fear, anxiety, and the amygdala. *Neuron*, 96(1):1-2. <https://doi.org/10.1016/j.neuron.2017.09.013>
- Serata D, Kotzalidis GD, Rapinesi C, et al., 2015. Are 5-HT<sub>3</sub> antagonists effective in obsessive-compulsive disorder? A systematic review of literature. *Hum Psychopharmacol*, 30(2):70-84. <https://doi.org/10.1002/hup.2461>
- Sharafkhah M, Alamdar MA, Massoudifar A, et al., 2019. Comparing the efficacy of ondansetron and granisetron augmentation in treatment-resistant obsessive-compulsive disorder: a randomized double-blind placebo-controlled study. *Int Clin Psychopharmacol*, 34(5):222-233. <https://doi.org/10.1097/yic.0000000000000267>
- Sharp T, Barnes NM, 2020. Central 5-HT receptors and their function; present and future. *Neuropharmacology*, 177: 108155. <https://doi.org/10.1016/j.neuropharm.2020.108155>
- Ślifirski G, Król M, Turło J, 2021. 5-HT receptors and the development of new antidepressants. *Int J Mol Sci*, 22(16): 9015. <https://doi.org/10.3390/ijms22169015>
- Smith WT, Londborg PD, Blomgren SL, et al., 1999. Pilot study of zatosetron (LY277359) maleate, a 5-hydroxytryptamine-3 antagonist, in the treatment of anxiety. *J Clin Psychopharmacol*, 19(2):125-131. <https://doi.org/10.1097/00004714-199904000-00006>
- Solt K, Ruesch D, Forman SA, et al., 2007. Differential effects of serotonin and dopamine on human 5-HT<sub>3A</sub> receptor kinetics: interpretation within an allosteric kinetic model. *J Neurosci*, 27(48):13151-13160. <https://doi.org/10.1523/jneurosci.3772-07.2007>
- Soltani F, Sayyah M, Feizy F, et al., 2010. A double-blind, placebo-controlled pilot study of ondansetron for patients with obsessive-compulsive disorder. *Hum Psychopharmacol*, 25(6):509-513. <https://doi.org/10.1002/hup.1145>
- Stefański R, Pałejko W, Bidzinski A, et al., 1993. Serotonergic innervation of the hippocampus and nucleus accumbens septi and the anxiolytic-like action of the 5-HT<sub>3</sub> receptor antagonists. *Neuropharmacology*, 32(10):987-993. [https://doi.org/10.1016/0028-3908\(93\)90063-9](https://doi.org/10.1016/0028-3908(93)90063-9)
- Szeszko PR, Robinson D, Alvir JMJ, et al., 1999. Orbital frontal and amygdala volume reductions in obsessive-compulsive disorder. *Arch Gen Psychiatry*, 56(10):913-919. <https://doi.org/10.1001/archpsyc.56.10.913>
- Taylor NE, Pei JZ, Zhang J, et al., 2019. The role of glutamatergic and dopaminergic neurons in the periaqueductal gray/dorsal raphe: separating analgesia and anxiety. *eNeuro*, 6(1):ENEURO.0018-18.2019. <https://doi.org/10.1523/eneuro.0018-18.2019>
- Togao O, Yoshiura T, Nakao T, et al., 2010. Regional gray and white matter volume abnormalities in obsessive-compulsive disorder: a voxel-based morphometry study. *Psychiatry Res Neuroimaging*, 184(1):29-37. <https://doi.org/10.1016/j.psychres.2010.06.011>
- Tovote P, Fadok JP, Lüthi A, 2015. Neuronal circuits for fear and anxiety. *Nat Rev Neurosci*, 16(6):317-331. <https://doi.org/10.1038/nrn3945>
- Tovote P, Esposito MS, Botta P, et al., 2016. Midbrain circuits for defensive behaviour. *Nature*, 534(7606):206-212. <https://doi.org/10.1038/nature17996>
- Tseilikman V, Akulov A, Shevelev O, et al., 2022. Paradoxical anxiety level reduction in animal chronic stress: a unique role of hippocampus neurobiology. *Int J Mol Sci*, 23(16):9151. <https://doi.org/10.3390/ijms23169151>
- Turner TJ, Mokler DJ, Luebke JI, 2004. Calcium influx through presynaptic 5-HT<sub>3</sub> receptors facilitates GABA release in the hippocampus: *in vitro* slice and synaptosome studies. *Neuroscience*, 129(3):703-718. <https://doi.org/10.1016/j.neuroscience.2004.08.020>
- Urzedo-Rodrigues LS, Ferreira HS, Santana RC, et al., 2014. Blockade of 5-HT<sub>3</sub> receptors in the septal area increases Fos expression in selected brain areas. *Auton Neurosci*, 181:55-68. <https://doi.org/10.1016/j.autneu.2014.01.003>
- Uvnäs-Moberg K, Hillegaart V, Alster P, et al., 1996. Effects of 5-HT agonists, selective for different receptor subtypes, on oxytocin, CCK, gastrin and somatostatin plasma levels in the rat. *Neuropharmacology*, 35(11):1635-1640. [https://doi.org/10.1016/s0028-3908\(96\)00078-0](https://doi.org/10.1016/s0028-3908(96)00078-0)
- Vilela-Costa HH, Maraschin JC, Casarotto PC, et al., 2021. Role of 5-HT<sub>1A</sub> and 5-HT<sub>2C</sub> receptors of the dorsal periaqueductal gray in the anxiety- and panic-modulating effects of antidepressants in rats. *Behav Brain Res*, 404:113159. <https://doi.org/10.1016/j.bbr.2021.113159>
- Vu V, Conant-Norville D, 2021. Anxiety: recognition and treatment options. *Psychiatr Clin North Am*, 44(3):373-380. <https://doi.org/10.1016/j.psc.2021.04.005>
- Yakel JL, Jackson MB, 1988. 5-HT<sub>3</sub> receptors mediate rapid responses in cultured hippocampus and a clonal cell line. *Neuron*, 1(7):615-621. [https://doi.org/10.1016/0896-6273\(88\)90111-0](https://doi.org/10.1016/0896-6273(88)90111-0)
- Yakel JL, Shao XM, Jackson MB, 1990. The selectivity of the channel coupled to the 5-HT<sub>3</sub> receptor. *Brain Res*, 533(1): 46-52. [https://doi.org/10.1016/0006-8993\(90\)91793-g](https://doi.org/10.1016/0006-8993(90)91793-g)
- Zhao HY, Lin Y, Chen SR, et al., 2018. 5-HT<sub>3</sub> receptors: a potential therapeutic target for epilepsy. *Curr Neuropharmacol*, 16(1):29-36. <https://doi.org/10.2174/1570159x15666170508170412>
- Zwanzger P, Domschke K, Bradwejn J, 2012. Neuronal network of panic disorder: the role of the neuropeptide cholecystokinin. *Depress Anxiety*, 29(9):762-774. <https://doi.org/10.1002/da.21919>